

No. 2019-1270

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**UNITED STATES COURT OF APPEALS  
FOR THE FEDERAL CIRCUIT**

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GENENTECH, INC.,

*Appellant,*

v.

ANDREI IANCU, Director, U.S. Patent and Trademark Office,

*Intervenor.*

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Appeal from the United States Patent and Trademark Office,  
Patent Trial and Appeal Board in No. IPR2017-01122

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**NON-CONFIDENTIAL BRIEF FOR APPELLANT GENENTECH, INC.**

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DARALYN J. DURIE  
ADAM R. BRAUSA  
DURIE TANGRI LLP  
217 Leidesdorff Street  
San Francisco, CA 94111  
(415) 362-6666

ANDREW J. DANFORD  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
60 State Street  
Boston, MA 02109  
(617) 526-6000

ROBERT J. GUNTHER, JR.  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
7 World Trade Center  
250 Greenwich Street  
New York, NY 10007  
(212) 230-8800

NORA Q.E. PASSAMANECK  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
1225 Seventeenth Street  
Suite 2600  
Denver, CO 80202  
(720) 274-3135

THOMAS G. SPRANKLING  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
950 Page Mill Road  
Palo Alto, CA 94304  
(650) 858-6000

THOMAS G. SAUNDERS  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
1875 Pennsylvania Avenue NW  
Washington, DC 20006  
(202) 663-6000

July 9, 2019

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## CERTIFICATE OF INTEREST

Counsel for Appellant Genentech, Inc. certifies the following:

1. The full name of every party or *amicus* represented by me is:

Genentech, Inc.

2. The names of the real party in interest represented by me is:

Not applicable.

3. All parent corporations and any publicly held companies that own 10 percent or more of the stock of the party or *amicus curiae* represented by me are:

Genentech, Inc. is a wholly-owned subsidiary of Roche Holdings Inc. Roche Holdings Inc.'s ultimate parent, Roche Holdings Ltd, is a publicly held Swiss corporation traded on the Swiss Stock Exchange. Upon information and belief, more than 10% of Roche Holdings Ltd's voting shares are held either directly or indirectly by Novartis AG, a publicly held Swiss corporation.

4. The names of all law firms and the partners or associates that appeared for the party or *amicus* now represented by me in the trial court or agency or are expected to appear in this Court (and who have not or will not enter an appearance in this case) are:

WILMER CUTLER PICKERING HALE AND DORR LLP: Owen K. Allen (former), Lauren V. Blakely, David L. Cavanaugh, Lisa J. Pirozzolo, Kevin S. Prussia, Rebecca A. Whitfield (former)

5. The title and number of any case known to counsel to be pending in this or any other court or agency that will directly affect or be directly affected by this Court's decision in the pending appeal:

*Genentech, Inc. et al. v. Amgen Inc.*, No. 1:18-cv-00924 (D. Del.)

*Genentech, Inc. v. Iancu*, No. 19-1263 (Fed. Cir.)

*In re Genentech, Inc.*, No. 19-1265 (Fed. Cir.)

*Genentech, Inc. v. Iancu*, No. 19-1267 (Fed. Cir.)

Dated: July 9, 2019

/s/ Robert J. Gunther, Jr.

ROBERT J. GUNTHER, JR.

WILMER CUTLER PICKERING

HALE AND DORR LLP

7 World Trade Center

250 Greenwich Street

New York, NY 10007

(212) 230-8800

## TABLE OF CONTENTS

|   | Page |
|---|------|
| CERTIFICATE OF INTEREST .....               | i    |
| TABLE OF AUTHORITIES .....                  | v    |
| STATEMENT OF RELATED CASES .....            | 1    |
| JURISDICTIONAL STATEMENT .....              | 1    |
| INTRODUCTION .....                          | 1    |
| STATEMENT OF ISSUES ON APPEAL .....         | 3    |
| STATEMENT OF THE CASE.....                  | 4    |
| A.    HER2-Positive Breast Cancer .....     | 4    |
| B.    The Invention of the '549 Patent..... | 4    |
| C.    Prior Art.....                        | 10   |
| 1.    Baselga '96.....                      | 11   |
| 2.    Taxol PDR '95 .....                   | 12   |
| 3.    Seidman '96 .....                     | 13   |
| 4.    Pegram.....                           | 13   |
| D.    Prosecution of the '549 Patent .....  | 14   |
| E.    The Board Proceedings .....           | 16   |
| SUMMARY OF THE ARGUMENT .....               | 17   |
| STANDARD OF REVIEW .....                    | 19   |
| ARGUMENT .....                              | 20   |



|     |  |    |
|-----|--|----|
| I.  | THE BOARD INCORRECTLY CONSTRUED THE TERMS “IN AN AMOUNT EFFECTIVE TO EXTEND THE TIME TO DISEASE PROGRESSION IN THE HUMAN PATIENT” AND “EFFECTIVE AMOUNT” TO REQUIRE COMPARISON TO AN UNTREATED PATIENT ..... | 20 |
| II. | UNDER A PROPER CONSTRUCTION, THE INVENTIONS OF THE CLAIMS ARE NONOBVIOUS .....   | 24 |
| A.  | The Board Erred In Finding That The Claimed Efficacy Was An Inherent Result Of An Otherwise Obvious Combination .....  | 25 |
| B.  | The Board Erred In Finding The Claimed Efficacy Was Obvious.....   | 29 |
|     | CONCLUSION .....   | 33 |
|     | ADDENDUM   |    |
|     | CERTIFICATE OF SERVICE   |    |
|     | CERTIFICATE OF COMPLIANCE  |    |

**CONFIDENTIAL MATERIAL OMITTED**

The material omitted from page 9 contains confidential communication from the FDA concerning Genentech’s submission.

## TABLE OF AUTHORITIES

### CASES

|  | Page(s)    |
|--|------------|
| <i>3M Innovative Properties Co. v. Tredegar Corp.</i> ,<br>725 F.3d 1315 (Fed. Cir. 2013) .....          | 22         |
| <i>Bayer AG v. Elan Pharmaceutical Research Corp.</i> ,<br>212 F.3d 1241 (Fed. Cir. 2000) .....          | 22         |
| <i>Ecolab, Inc. v. FMC Corp.</i> ,<br>569 F.3d 1335 (Fed. Cir. 2009) .....                               | 23         |
| <i>Endo Pharmaceuticals Solutions, Inc. v. Custopharm Inc.</i> ,<br>894 F.3d 1374 (Fed. Cir. 2018) ..... | 26, 27     |
| <i>Hamilton Beach Brands, Inc. v. f'real Foods, LLC</i> ,<br>908 F.3d 1328 (Fed. Cir. 2018) .....        | 19         |
| <i>In re Omeprazole Patent Litigation</i> ,<br>483 F.3d 1364 (Fed. Cir. 2007) .....                      | 27         |
| <i>Leo Pharmaceutical Products, Ltd. v. Rea</i> ,<br>726 F.3d 1346 (Fed. Cir. 2013) .....                | 19         |
| <i>Millennium Pharmaceuticals, Inc. v. Sandoz Inc.</i> ,<br>862 F.3d 1356 (Fed. Cir. 2017) .....         | 26         |
| <i>Omega Engineering, Inc. v. Raytek Corp.</i> ,<br>334 F.3d 1314 (Fed. Cir. 2003) .....                 | 22         |
| <i>PAR Pharmaceutical, Inc. v. TWI Pharmaceuticals, Inc.</i> ,<br>773 F.3d 1186 (Fed. Cir. 2014) .....   | 26, 27, 28 |
| <i>Personal Web Technologies, LLC v. Apple, Inc.</i> ,<br>848 F.3d 987 (Fed. Cir. 2017) .....            | 19         |
| <i>SanDisk Corp. v. Memorex Products, Inc.</i> ,<br>415 F.3d 1278 (Fed. Cir. 2005) .....                 | 22         |
| <i>Santarus, Inc. v. Par Pharmaceutical, Inc.</i> ,<br>694 F.3d 1344 (Fed. Cir. 2012) .....              | 28         |

|   |    |
|---|----|
| <i>Shire LLC v. Amneal Pharmaceuticals, LLC</i> ,<br>802 F.3d 1301 (Fed. Cir. 2015) ..... | 30 |
|---|----|

## STATUTES

|                                 |   |
|---------------------------------|---|
| 28 U.S.C. § 1295(a)(4)(A) ..... | 1 |
| 35 U.S.C.                       |   |
| § 141(c) .....                  | 1 |
| §§ 311-319 .....                | 1 |
| § 319.....                      | 1 |

## **STATEMENT OF RELATED CASES**

No appeal from the same proceeding was previously before this Court or any other appellate court. The following cases will directly affect or be directly affected by this Court's decision in the pending appeal: *Genentech, Inc. et al. v. Amgen Inc.*, No. 1:18-cv-00924 (D. Del.); *Genentech, Inc. v. Iancu*, No. 19-1263 (Fed. Cir.); *In re Genentech, Inc.*, No. 19-1265 (Fed. Cir.); and *Genentech, Inc. v. Iancu*, No. 19-1267 (Fed. Cir.).

## **JURISDICTIONAL STATEMENT**

The Patent Trial and Appeal Board asserted jurisdiction under 35 U.S.C. §§ 311-319. This Court has jurisdiction over the appeal from the Board's final written decisions pursuant to 28 U.S.C. § 1295(a)(4)(A) and 35 U.S.C. §§ 141(c) & 319. Genentech filed a timely notice of appeal on November 30, 2018 in IPR2017-01122. Appx15017-15021.

## **INTRODUCTION**

The invention in this case arises from Genentech's groundbreaking work in the treatment of breast cancer. As of the priority date, the FDA had never approved an antibody therapy for solid tumors, such as breast cancer. But Genentech made a critical discovery: an "anti-ErbB2" antibody could be used in combination with a relatively new type of chemotherapy drug called a "taxoid" to treat cancers that overexpress a protein called HER2. Specifically, Genentech's

priority application disclosed the first results ever reported from human trials of the combination of the anti-ErbB2 antibody “trastuzumab” (also called “rhuMAb HER2”) and the taxoid “paclitaxel.” Those results showed that rhuMAb HER2 and paclitaxel, in the absence of another common chemotherapy drug (an “anthracycline derivative”), could extend the time to disease progression (i.e., the time from diagnosis or treatment until the disease starts to worsen or spread), without increasing overall severe adverse events.

Genentech claimed that invention in U.S. Patent No. 7,846,441 (“the ’441 patent”), and when the FDA approved Genentech’s drug Herceptin®, the combination became the only approved first-line antibody-based therapy for solid tumors. The patent in this appeal, U.S. Patent No. 7,892,549 (“the ’549 patent”), is a continuation of the ’441 patent and shares the same specification. While there are various differences between the ’549 and ’441 claims, the most notable is that all of the ’549 claims recite the third agent (i.e., “further growth inhibitory agent” or “further therapeutic agent”) in addition to the combination of an anti-ErbB2 antibody and a taxoid claimed in the ’441 patent. The ’549 patent has been terminally disclaimed over the ’441 patent.

This appeal arises from a final written decision by the Patent Trial and Appeal Board declaring all claims of the ’549 patent unpatentable. Much of the dispute before the Board turned on the meaning of the claim terms “in an amount

effective to extend the time to disease progression in the human patient” and “effective amount.” Based on a single inartful statement in the prosecution history, the Board construed these terms in a way that did not match what Genentech taught in its specification or the subject matter it wants to protect. Specifically, the Board misconstrued these terms to require comparing the claimed combination with an untreated patient—i.e., a cancer patient receiving no treatment whatsoever. The Board did so even though (1) the specification disclosed comparisons to patients treated with paclitaxel alone, but no comparisons to untreated patients; (2) as a matter of basic medical ethics, a patient cannot be left untreated; and (3) when read in context, the statement from the prosecution history that the Board relied on was not a clear and unmistakable disclaimer. Under the correct construction, which requires comparing the combined treatment to treatment with paclitaxel alone, there was no sound basis to rule that Genentech’s claims are unpatentable.

### **STATEMENT OF ISSUES ON APPEAL**

I. Whether the Board incorrectly construed the terms “in an amount effective to extend the time to disease progression in the human patient” and “effective amount” to require a comparison to a patient who had received no treatment at all.

II. Whether, applying the proper construction of the terms “in an amount effective to extend the time to disease progression in the human patient” and

“effective amount,” the Board’s decision should be reversed because it was not supported by substantial evidence.

## **STATEMENT OF THE CASE**

### **A. HER2-Positive Breast Cancer**

“HER2-positive” cancers have a genetic mutation that causes them to overexpress human epidermal growth factor 2 (“HER2”), also known as human ErbB2. Out of the hundreds of thousands of women each year who are diagnosed with breast cancer, roughly 25-30% are HER2-positive. Appx72(1:26-32); Appx9741. HER2-positive breast cancer is particularly aggressive: In the 1990s, it had the worst prognosis in women with breast cancer. It was associated with a high rate of tumor recurrence and spreading to other areas of the body, shorter time to relapse, and shorter overall survival. Appx8957; Appx8892-8894; Appx8899. While HER2-normal breast cancer patients could expect to live for six to seven years post-diagnosis, the post-diagnosis life expectancy of HER2-positive breast cancer patients receiving standard chemotherapy treatment in 1996 was about 18 months. Appx8943; Appx8945; Appx9741-9743.

### **B. The Invention of the ’549 Patent**

The ’549 patent claims a method for treating HER2-positive breast cancer patients with an anti-ErbB2 antibody such as “trastuzumab” (aka “rhuMAb HER2”) in combination with a type of chemotherapy drug called a “taxoid,” along

with “a further growth inhibitory agent” (claims 1, 16) or “a further therapeutic agent” (claim 5). Specifically, the ’549 claims reflect a novel method of treatment for cancer that overexpresses ErbB2 (e.g., HER2-positive breast cancer), which comprises (i) “administering a combination” of an anti-ErbB2 antibody (such as rhuMAb HER2), a taxoid (a type of chemotherapy drug), and “a further growth inhibitory agent” (claims 1, 16) or “a further therapeutic agent” (claim 5); (ii) “to the human patient”; (iii) “in an amount effective to extend the time to disease progression in the human patient” (claims 1, 16) or in “an effective amount” (claim 5). Claims 16 and 17 further require “the absence of an anthracycline derivative” from the claimed combination therapy.

In the 1990s, engineered antibodies—proteins specially-designed to bind to molecular targets, called “antigens”—were a focus for therapeutic research. Appx75-76(8:45-9:4). However, the body’s immune system also tended to attack these antibodies, preventing them from having a therapeutic effect. Appx9054. Articles from the 1990s described antibody therapy for cancer as “a story of unending failures,” Appx9091, with “significant obstacles,” Appx9084, and “no hint of a consistent therapeutic efficacy,” Appx8979. When the provisional application for the ’441 patent was filed in December 1997, *no antibody* had been approved for the treatment of solid tumors such as breast cancer.



During this time, oncologists were also slow to adopt taxoids for treating breast cancer. The prior art came to conflicting conclusions about HER2 response to taxanes (a type of taxoids), with reports that “HER2 over-expression in [metastatic breast cancer] seems to confer sensitivity rather than resistance to taxanes,” Appx3479, and that “breast cancers that overexpress p185 [*i.e.*, HER2] ***will not respond well to Taxol,***” Appx9016 (emphasis added). As of December 1997, ***no clinical results*** had been reported for the combination of trastuzumab and a taxoid. The only results for the combination were in preclinical mouse models. In these models, mice with suppressed immune systems are injected with human cancer cells and treated with therapies being considered for human testing. Preclinical mouse models were understood at the time to be a useful initial mechanism to screen for drugs that show some activity against particular cancer cells, and to understand their mechanism of function. Appx9569-9572; Appx9727-9729. However, as of 1997, it was also well-recognized that mouse studies failed to reliably predict what therapies would ultimately be successful in humans. *See* Appx9492 (noting “[t]he fundamental problem in drug discovery for cancer is that the model systems are not predictive” and “drugs tested in the xenografts appeared effective but worked poorly in humans”); Appx8961 (“very low” likelihood of mouse studies predicting responses in humans).

There are many reasons for this. Mouse studies are short-term and generally measure only “response rate”—i.e., the ability of a therapy to shrink tumors—not effect on time to disease progression (“TTP”). Response rate and TTP are measuring different endpoints. A therapy may demonstrate a response rate by initially shrinking tumors, but fail to eradicate the most-aggressive cancer cells that cause the cancer to progress quickly. It was established that therapies may improve response rates but not affect TTP. Appx9738-9739; Appx9884. Mice are also often administered a proportionally larger dose than humans can tolerate, which allows for positive outcomes not possible in humans. Appx8946. Therapies also frequently cause toxicity in humans, but not in mice, due to differences in cell and tissue types between mice and humans. Appx9578-9580; Appx9730. Furthermore, mouse models are more likely to show positive outcomes because they use tumor cell lines from tissue culture. These divide more rapidly than human cells, which are heterogenous and therefore can display greater sensitivity to treatment. Appx9562-9563; Appx9730.

In addition, prior to December 1997, no Phase III clinical results existed for the antibody trastuzumab, alone or in any combination. Clinical testing of a drug—that is, testing of a drug in humans—occurs in stages, beginning with initial small-scale studies (i.e., “Phase I” and “Phase II” studies), followed by large-scale “Phase III” controlled trials designed to evaluate specific clinical endpoints.

Appx9734-9736. At each of these stages, a large number of therapies fail. In the 1990s, only 5% of cancer drugs that advanced to clinical trials resulted in an approved product. Appx8952-8953. Even for drugs that advanced to late-stage, Phase III clinical trials, nearly 60% ultimately failed to result in an approved drug. *Id.*

Without running a Phase I or Phase II study, Genentech decided to test the combination of trastuzumab and a taxoid—specifically paclitaxel—in a Phase III trial of HER2-positive metastatic breast cancer patients. Genentech tested this combination, not because of promising results in the prior art, but because Genentech’s ongoing Phase III study involving a combination of trastuzumab and ***a different chemotherapeutic agent***—anthracyclines—was having difficulty enrolling patients. Appx10257. Moreover, the inventor who proposed the combination had just joined Genentech from the company that made Taxol (paclitaxel) and had unique familiarity with the drug well beyond the knowledge of an ordinary artisan. Appx9753.

Running a Phase III study without first testing the drug in Phases I and II is so unusual that, while the proposal to add a trastuzumab and paclitaxel arm to the Phase III study was adopted, it was met with skepticism both at Genentech and at the FDA. *See, e.g.*, Appx8090 (“[T]he expected clinical outcome for the administration of rhuMAb HER2 with taxol is less certain than co-administration

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with cisplatin or doxorubicin.”); Appx10518 (FDA noting that Genentech has

“[REDACTED]  
[REDACTED]”).

Yet when the Phase III study reached its primary endpoint in late 1997, the results were surprising. Appx8613-8620; Appx8666-8671. The study data showed that trastuzumab and paclitaxel in the absence of an anthracycline derivative extended TTP compared to paclitaxel alone, without an increase in overall severe adverse events. Appx8761. In fact, the combination of trastuzumab and paclitaxel was dramatically more effective than paclitaxel alone. *See, e.g.*, Appx2665 (“[T]he combination is surprisingly synergistic with respect to extending TTP.”). In addition, the combination of trastuzumab and paclitaxel unexpectedly avoided the surprising cardiotoxicity that resulted from the combination of trastuzumab and anthracyclines. Appx8760; Appx8012; Appx3664; Appx2664; Appx86. These data are reflected in the provisional patent application filed December 12, 1997. Appx8874-8879, and led to the FDA approval of Herceptin as a first-line treatment.

Herceptin revolutionized the treatment of HER2-positive breast cancer. Appx8945 (“Genentech are now poised for another impressive therapeutic breakthrough for late-stage treatment of breast cancer,” with clinical trials showing

“particularly encouraging [results] in combination with chemotherapy using paclitaxel[.]”).

### C. Prior Art

The Board’s Final Written Decision addressed the validity of the ’549 patent based on Baselga ’96,<sup>1</sup> the Taxol PDR ’95,<sup>2</sup> Seidman ’96,<sup>3</sup> and Pegram,<sup>4</sup> and the knowledge of one of ordinary skill in the art, evidenced, in part, by Baselga Abstract 53,<sup>5</sup> Baselga Abstract 2262,<sup>6</sup> and Seidman ’95.<sup>7</sup> Appx17-22. None of

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<sup>1</sup> Baselga et al., *Phase II Study of Weekly Intravenous Recombinant Humanized Anti-p185<sup>HER2</sup> Monoclonal Antibody in Patients with HER2/neu-Overexpressing Metastatic Breast Cancer*, 14 J. Clin. Oncol. 737-744 (1996). Appx3665-3674.

<sup>2</sup> Taxol® (Paclitaxel) for Injection Concentrate, Physicians’ Desk Reference, 682-685 (49th ed. 1995). Appx3480-3487.

<sup>3</sup> Seidman et al., *Over-Expression and Clinical Taxane Sensitivity: A Multivariate Analysis in Patients with Metastatic Breast Cancer (MBC)*, 15 Proc. Am. Soc’y Clin. Oncology 104 (1996) (Abstract 80). Appx3475-3479.

<sup>4</sup> M. Pegram et al., *Phase II Study of Intravenous Recombinant Humanized Anti-p185 HER-2 Monoclonal Antibody (rhuMAb HER-2) Plus Cisplatin in Patients with HER-2/NEU Overexpressing Metastatic Breast Cancer*, 14 Proc. Am. Soc’y Clinical Oncology 106 (Mar. 1995) (Abstract 124). Appx3678-3680.

<sup>5</sup> J. Baselga et al., *Anti HER2 Humanized Monoclonal Antibody (MAb) Alone and in Combination with Chemotherapy Against Human Breast Xenografts*, 13 Proc. Am. Soc’y Clinical Oncology 63 (1994) (Abstract 53). Appx3661-3664.

<sup>6</sup> Baselga et al., *Antitumor Activity of Paclitaxel in Combination with Anti-growth Factor Receptor Monoclonal Antibodies in Breast Cancer Xenografts*, 35 Proc. Am. Ass’n For Cancer Res. 380 (1994) (Abstract 2262). Appx3675-3677.

<sup>7</sup> Seidman et al., *Memorial Sloan-Kettering Cancer Center Experience with Paclitaxel in the Treatment of Breast Cancer*, 22 Seminars Oncology 108-116 (1995). Appx3466-3474.

these references contain any clinical data showing the effect of trastuzumab plus a taxoid in humans. Indeed, it is undisputed that no such clinical data was reported prior to December 12, 1997. Appx9749-9750.

### **1. Baselga '96**

Baselga '96 is an article published in March 1996. It describes the results of a Phase II clinical study in which 46 patients received rhuMAb HER2 *alone*, not combined with a taxoid (or any other chemotherapy or agent). Appx3668.

The clinical endpoint evaluated in the trial was response rate, which was evaluated at 11 weeks. Appx3668; Appx3670-3671. Although Baselga '96 measured TTP for individual patients, every patient received rhuMAb HER2 and the study included no control. Baselga '96 thus provided no way to measure *extension* of TTP, which requires a comparator.

The vast majority of patients in Baselga '96 did not show a therapeutic response—only 5 out of the 43 assessable patients (11.6%) had complete or partial responses to treatment with rhuMAb HER2. Of the remaining patients, 2 had a minor response, 14 had stable disease, and 22 patients (over 50%) had disease progression at 11 weeks. Appx3671. While Baselga '96 measured a “median time to progression” of 5.1 months, it measured this for only the 16 patients with minor response or stable disease—it did not take into account the 22 patients whose disease progressed at 11 weeks or earlier. Appx3670.

Baselga '96 explained the mechanism of action of rhuMAb HER2 was not understood, offering several possible explanations for the clinical results.

Appx3672-3673. Baselga '96 also cited earlier preclinical mouse studies, which are described in the Baselga Abstract 53, Appx3664, and Baselga Abstract 2262, Appx3677 (collectively, the "Baselga abstracts"). Baselga '96 noted that in the Baselga abstracts, "rhuMAb HER2 markedly potentiated the antitumor effects of several chemotherapeutic agents, including cisplatin, doxorubicin, and paclitaxel, without increasing their toxicity." Appx3673.

## **2. Taxol PDR '95**

Taxol PDR '95 is the entry from the 1995 Physicians' Desk Reference corresponding with paclitaxel. Appx3480-3487. It does not suggest combining paclitaxel with anti-ErbB2 antibodies, or even mention anti-ErbB2 antibodies. Appx9233. Moreover, it does not mention HER2-positive breast cancer or suggest that taxoids would be effective to treat HER2-positive patients.

Taxol PDR '95 further states that paclitaxel was approved only as a second-line therapy for metastatic breast cancer (i.e., after the failure of other treatments), and notes that, in general, paclitaxel should be used only after anthracycline therapy. Appx3485. Taxol PDR '95 additionally includes a black box "WARNING" regarding the possibility of "severe hypersensitivity reactions" and notes that at least one patient died from those side effects. Appx3484-3485; *see*

*also* Appx9231 (this warning is “the FDA’s way of flagging a drug, some things that you need to know about the drug.”); Appx9772-9773.

### **3. Seidman ’96**

Seidman ’96 is an abstract published in March 1996, which describes a retrospective analysis of tumor samples for metastatic breast cancer patients “who were treated on phase II protocols of single-agent paclitaxel (n=106) or docetaxel (n=20).” Appx3479. Seidman ’96 does not mention antibody therapy at all. In addition, Seidman ’96 does not address whether taxoids extend TTP in HER2-positive patients, instead measuring the “response proportion”—a different clinical endpoint. *Id.* With respect to the single-agent chemotherapies studied, Seidman ’96 reports that the response proportion was 58.8% among HER2-positive patients and 38.7% among HER2-negative patients. *Id.* Given the shortcomings of this disclosure, the Board did not rely on or discuss this reference in its analysis on reasonable expectation of success.

### **4. Pegram**

Pegram is another abstract that describes a Phase II study in which 36 HER2-positive patients were administered a combination of rhuMAb-HER2 plus cisplatin. Appx3680. Pegram does not describe any treatment involving paclitaxel, or suggest a combination with it.



#### **D. Prosecution of the '549 Patent**

The '549 patent issued from Application No. 10/356,824 filed on February 3, 2003, which is a continuation of U.S. Patent Application No. 09/208,649, filed on December 10, 1998, which later issued as the '441 patent. In turn, the '649 application claims priority to Provisional Application No. 60/069,346, filed on December 12, 1997. Appx65. As noted, the December 12, 1997 provisional application contained the first disclosure of results from testing the combination of rhuMAb HER2 and paclitaxel in humans, and the first data of any kind regarding the combination's extension of TTP compared to paclitaxel alone.

During prosecution of the '649 application, the Examiner made the following statement while rejecting the claims pending at the time as indefinite:

The term "extend time to disease progression" is not defined by the claim, the specification does not provide a standard for ascertaining the requisite degree, and one of ordinary skill in the art would not be reasonably apprised of the scope of the invention. Specifically, it is never set forth what the extension of time to disease progress is relative to, for example, is the extension of time to disease progress relative to untreated patients? Patients who received antibody or taxoid alone? Patients who received antibody and an anthracycline?

Appx11400-11401. In January 2002, the applicant responded that "the expression[] 'extend the time to disease progression' ... [is] clear from the specification (see, in particular, page 15, lines 15-17; and pages 42-43) and would be readily understood by the skilled oncologist." Appx11416. The portions of the specification cited by the applicant to indicate that the proper comparison was

“clear from the specification” stated that “efficacy can, for example, be measured by assessing the time for disease progression (TTP),” Appx11019, and then disclosed the Phase III data cited above that compared treatment with a combination of rhuMAb HER2 and paclitaxel to treatment *with paclitaxel alone, not to a lack of treatment altogether*, Appx11046-11047. The applicant’s response to the Examiner followed these citations by saying: “Clearly, the combination of anti-ErbB2 antibody and taxoid is administered in an amount effective to extend the time to disease progression relative to an untreated patient.” Appx11416.

In October 2009, Genentech submitted a declaration from Dr. Mark Sliwowski in response to obviousness rejections over, among other things, a combination of Baselga ’96 and Baselga ’94. Appx3369. Dr. Sliwowski explained that a skilled artisan would not have expected rhuMAb HER2 combined with a taxoid to produce a synergistic response, since those drugs were known to exert their effects at different points in the cell cycle. Appx3370. Dr. Sliwowski also explained that preclinical results would not have provided a reasonable expectation of success as to the clinical results for the combination of rhuMAb HER2 and a taxoid; indeed, he expressed that xenograft models at that time were poor predictors of clinical results for breast cancer. Appx3371-3372.

After Genentech provided a terminal disclaimer over the parent application (which issued as the '441 patent), Appx3108-3109, the Examiner allowed the claims on October 8, 2010, Appx3132-3133.

#### **E. The Board Proceedings**

On March 21, 2017, Petitioner Celltrion filed a Petition for *inter partes* review of claims 1-11 and 14-17 of the '549 patent. Appx14005-14079. Petitioner challenged the patentability of these claims based on a combination of Baselga '96, Seidman '96, Pegram, 1995 TAXOL PDR, and the knowledge of one of ordinary skill in the art, evidenced, in part, by Baselga Abstract 53, Baselga Abstract 2262, and Seidman '95. The Board instituted *inter partes* review on October 4, 2017. Appx14243.

The Board's final written decision, issued on October 3, 2018, determined that Petitioner showed by a preponderance of the evidence that claims 1-11 and 14-17 of the '549 patent would have been obvious over a combination of Baselga '96, Seidman '96, Pegram, 1995 TAXOL PDR, and the knowledge of one of ordinary skill in the art. Appx61. In so holding, the Board relied on a claim construction of "in an amount effective to extend the time to disease progression in the human patient" and "effective amount" that compared "the claimed combination treatment to no treatment." Appx41. The Board also found that an ordinary artisan would have been motivated to combine rhuMAb HER2 and paclitaxel to treat patients

with ErbB2 overexpressing metastatic breast cancer, and that “the challenged claims would have been obvious” even applying “Patent Owner’s preferred construction.” Appx47.

### **SUMMARY OF THE ARGUMENT**

I. The Board adopted an incorrect claim construction of the terms “in an amount effective to extend the time to disease progression in the human patient” and “effective amount.” The Board erroneously interpreted those terms to require a comparison to an untreated patient. Instead, the appropriate comparison is to a patient treated with a taxoid alone, which is the only comparison described in the patent specification that is consistent with the language of the claims. The specification reports nothing about untreated patients.

The Board based its construction on a single statement in the file history about comparison to an “untreated patient.” But that statement, which cites the example in the specification that compares patients treated with the combination of anti-ErbB2 antibody (rhuMAb HER2) and a taxoid (paclitaxel) to patients treated with a taxoid (paclitaxel) alone, does not change how a skilled artisan would understand the term and does not meet the demanding standard to establish prosecution disclaimer.

II. The Board erred in finding that even under Genentech’s construction, a skilled artisan would have had a reasonable expectation that the combination of

anti-ErbB2 antibody and a taxoid would extend TTP as compared to treatment with a taxoid alone.

First, the Board improperly held that the claimed extension of TTP is an inherent benefit of an otherwise obvious combination. This conclusion ignored the high standard for inherency, which requires that a missing limitation must be necessarily present or the natural result of the combination of elements explicitly disclosed in the prior art. It is undisputed that administering the claimed combination does not extend TTP each and every time. Indeed, the record establishes that some patients who were administered rhuMAb HER2 and paclitaxel experienced no extension in TTP. Nor is extension of TTP the natural result expected for a claimed combination expressly taught in the prior art. There was no disclosure in the prior art of treating human patients with rhuMAb HER2, a taxoid, and a third agent. Moreover, there was no disclosure that the combination of rhuMAb HER2 and paclitaxel would extend TTP as compared to paclitaxel alone in human patients.

Second, substantial evidence did not support the Board's finding that a skilled artisan would have a reasonable expectation that the combination would extend TTP as compared to treatment of a taxoid alone by improperly relying on Baselga '96's report that the TTP in patients administered rhuMAb HER2 alone was 5.1 months, as compared to the TTP of paclitaxel reported in the Taxol PDR

'95 of 3.0 or 4.2 months. A skilled artisan would not make this comparison, much less draw the same conclusion as the Board. The 5.1 month TTP reported in Baselga '96 was only for those patients that reported a minor response or stable disease, and excluded over half of the patients—those whose cancer progressed.

### **STANDARD OF REVIEW**

The Board's claim construction is subject to *de novo* review where, as here, the Board relied on only intrinsic evidence to construe the claims. *Hamilton Beach Brands, Inc. v. f'real Foods, LLC*, 908 F.3d 1328, 1339 (Fed. Cir. 2018).

The Board's ultimate finding on obviousness is a legal conclusion, which this Court reviews de novo. *Personal Web Techs., LLC v. Apple, Inc.*, 848 F.3d 987, 991 (Fed. Cir. 2017); *Leo Pharm. Prods., Ltd. v. Rea*, 726 F.3d 1346, 1353 (Fed. Cir. 2013) (“[A]t bottom, this court confronts a question of law: whether, in light of the prior art references and objective indicia of nonobviousness, the claimed invention would have been obvious to a person of ordinary skill in the art at a time just before the time of invention.”).

Underlying factual findings are reviewed for substantial evidence. Substantial evidence review asks “whether a reasonable fact finder could have arrived at the agency’s decision, which requires examination of the record as a whole, taking into account evidence that both justifies and detracts from an

agency's decision.” *Personal Web Techs.*, 848 F.3d at 991 (quotation marks omitted).

## **ARGUMENT**

This Court should reverse because the Board erroneously construed the claims and its alternative ruling under the correct construction was not supported by substantial evidence.

### **I. THE BOARD INCORRECTLY CONSTRUED THE TERMS “IN AN AMOUNT EFFECTIVE TO EXTEND THE TIME TO DISEASE PROGRESSION IN THE HUMAN PATIENT” AND “EFFECTIVE AMOUNT” TO REQUIRE COMPARISON TO AN UNTREATED PATIENT**

The claim language and specification make clear that the terms “in an amount effective to extend the time to disease progression in the human patient” and “effective amount” require comparing the claimed combination treatment to treatment with a taxoid alone. All of the data contained in the patent focuses on this comparison. The Board nonetheless construed the claims to require a comparison to a patient who has received no treatment based on an isolated, if inartful, statement in the prosecution history that does not satisfy the demanding standard for establishing a disclaimer. This Court should reverse.

The specification makes clear that the claims require comparing the claimed combination treatment to treatment with a taxoid alone. There is no data in the '549 patent comparing the time to disease progression of patients treated with rhuMAb HER2 and paclitaxel against an untreated patient. Rather, the '549 patent

describes a Phase III clinical trial measuring the efficacy of the combination of an anti-ErbB2 antibody (rhuMAb HER2) with a taxoid (paclitaxel) *against a control arm of paclitaxel alone*. Appx86(29:11-30:25) (comparing T + H (i.e., Taxol and Herceptin) to T (i.e., Taxol)).<sup>8</sup> The specification thus refutes the Board's conclusion that the claims require comparing Genentech's combined treatment to no treatment at all.

Indeed, a comparison to an untreated patient makes no sense in the context of a disease like breast cancer where there were already therapies approved by the FDA. Undisputed expert testimony established that it would be unethical to conduct a study comparing the efficacy of a tested therapy against no therapy where there was already an approved therapy that would provide a clinical benefit to the target patient population for a life-threatening disease like breast cancer. Appx9766-9767 ("It would not be ethical to design a study to compare efficacy against no therapy alone where there was already an approved therapy that would provide a clinical benefit to the target patient population.").

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<sup>8</sup> The '549 patent also describes the efficacy of rhuMAb HER2 combined with chemotherapy (paclitaxel or anthracyclines) versus chemotherapy alone, or rhuMAb HER2 combined with anthracyclines versus anthracycline therapy alone. Appx86(29:11-30:25). However, given that the claims expressly exclude anthracycline therapy, the relevant comparison is the combination of rhuMAb HER2 and paclitaxel versus paclitaxel alone.



The Board did not dispute any of these points. Instead, it based its claim construction exclusively on the prosecution history. Specifically, the Board relied on the applicant's statement in January 2002 that "the expression[] 'extend the time to disease progression' ... [is] clear from the specification (see, in particular, page 15, lines 15-17; and pages 42-43) and would be readily understood by the skilled oncologist. Clearly, the combination of anti-ErbB2 antibody and taxoid is administered in an amount effective to extend the time to disease progression relative to an untreated patient." Appx11416.

The Board's use of this prosecution history to override the meaning evident from the specification was error. The standard for establishing prosecution disclaimer is high: The statement must "show 'a clear and unmistakable surrender of subject matter.'" *Bayer AG v. Elan Pharm. Research Corp.*, 212 F.3d 1241, 1251 (Fed. Cir. 2000). In other words, the statement must "unequivocally disavow[] a certain meaning." *Omega Eng'g, Inc. v. Raytek Corp.*, 334 F.3d 1314, 1324 (Fed. Cir. 2003).

"There is no 'clear and unmistakable' disclaimer if a prosecution argument is subject to more than one reasonable interpretation, one of which is consistent with a proffered meaning of the disputed term." *SanDisk Corp. v. Memorex Prods., Inc.*, 415 F.3d 1278, 1287 (Fed. Cir. 2005); *see also 3M Innovative Props. Co. v. Tredegar Corp.*, 725 F.3d 1315, 1326 (Fed. Cir. 2013) ("Where an

applicant's statements are amenable to multiple reasonable interpretations, they cannot be deemed clear and unmistakable.”). Further, the clarity of a statement cannot be determined in isolation but must be considered in the context of the entire record. “Even if an isolated statement appears to disclaim subject matter, the prosecution history as a whole may demonstrate that the patentee committed no clear and unmistakable disclaimer.” *Ecolab, Inc. v. FMC Corp.*, 569 F.3d 1335, 1342 (Fed. Cir. 2009).

The applicant's statement regarding an “untreated patient” was admittedly inartful when juxtaposed against the Examiner's questions. Read in context, however, it does not rise to the level of a “clear and unmistakable disclaimer.” The Board failed to give any weight to the applicant's immediately preceding statement that the meaning of the limitation was “clear from the specification (see, in particular, page 15, lines 15-17; and pages 42-43).” The highlighted portions of the specification introduced the concept of measuring TTP and disclosed the Phase III data cited above that compared treatment with a combination of rhuMAb HER2 and paclitaxel to treatment *with paclitaxel alone*, not a lack of treatment altogether, Appx11019; Appx11046-11047. From the outset, the applicant's reference to an “untreated patient” was thus made based on data showing a comparison to patients *untreated with the combination of rhuMAb HER2 and*

*paclitaxel because they were treated with paclitaxel alone.* The applicant’s statement thereby undermines, rather than supports, the Board’s construction.

Consistent with the plain meaning of the claim and specification, this Court should construe the terms “in an amount effective to extend the time to disease progression in the human patient” and “effective amount” as requiring a measurement against a patient treated with a taxoid alone.

## **II. UNDER A PROPER CONSTRUCTION, THE INVENTIONS OF THE CLAIMS ARE NONOBVIOUS**

The Board held that even under Genentech’s proposed claim construction, claims 1-17 of the ’549 patent are obvious because (1) the claimed extension of TTP is an inherent benefit of an otherwise obvious combination; and (2) an ordinary artisan would have expected the claimed extension of TTP based on the reported TTP of rhuMAb HER2 alone in Baselga ’96. Appx41-47. The Board’s conclusions cannot stand because they are based on a misunderstanding of the legal framework of inherency and not supported by substantial evidence.

It is undisputed that, as of December 1997, Genentech was at the leading edge of a fundamentally new approach to treating breast cancer. Instead of traditional chemotherapy, it was using rhuMAb HER2, a human-engineered antibody, to treat solid tumors—an approach that had never received approval from the FDA. Adding to the unpredictability, it was combining the use of a therapeutic antibody with a relatively new compound, paclitaxel, to achieve synergistic

improvement to TTP without increasing adverse events compared to treatment with paclitaxel alone.

The prior art that the Board relied on to hold that Genentech's breakthrough would have been obvious left significant gaps that the Board never overcame. For example, it is undisputed that no testing of the combination of rhuMAb HER2 and paclitaxel in humans had ever been reported before Genentech's patent application. Further, it is undisputed that no TTP results for the combination had ever been reported—even in a preclinical model. In an unpredictable art like breast cancer therapy, these holes in the prior art left the Board without a legally or scientifically sound basis for finding a reasonable expectation of success. The Board's failure to recognize this unpredictability and willingness to stretch the references was error.

**A. The Board Erred In Finding That The Claimed Efficacy Was An Inherent Result Of An Otherwise Obvious Combination**

The Board's inherency ruling was legally flawed. Petitioner did not argue inherency in its petition or present any expert testimony on the subject, but the Board held the claimed extension of TTP "is an inherent benefit of an otherwise obvious combination, and ... such an inherent result cannot establish patentability." Appx42. The Board reasoned that "when rhuMoAb is administered with a chemotherapy in the taxoid family, this claimed combination therapy significantly extends the time to disease progression (TTP) as compared with patients receiving taxoid therapy alone." Appx42 (internal quotation marks

omitted). In making this findings, the Board did not honor the legal framework governing inherency and ignored the substantial shortcomings of the prior art.

A challenger must “meet a high standard in order to rely on inherency to establish the existence of a claim limitation in the prior art in an obviousness analysis.” *PAR Pharm., Inc. v. TWI Pharms., Inc.*, 773 F.3d 1186, 1195-1196 (Fed. Cir. 2014); *see also id.* at 1195 (“inherency, a doctrine originally rooted in anticipation, must be carefully circumscribed in the context of obviousness”). “The limitation at issue ***necessarily*** must be present, or the ***natural result*** of the combination of elements ***explicitly disclosed*** by the prior art.” *Id.* (emphasis added); *see also Endo Pharms. Sols., Inc. v. Custopharm Inc.*, 894 F.3d 1374, 1381-1382 (Fed. Cir. 2018) (outlining elements and describing them as a “rigorous” standard). The record evidence does not support a finding of inherency under either of these prongs.

First, extension of TTP does not “necessarily” result from the claimed combination. “The mere fact that a certain thing ***may result*** from a given set of circumstances is not sufficient to render the result inherent.” *Millennium Pharms., Inc. v. Sandoz Inc.*, 862 F.3d 1356, 1367 (Fed. Cir. 2017) (internal quotations omitted) (emphasis added). “Inherency ... may not be established by probabilities or possibilities.” *PAR Pharm.*, 773 F.3d at 1195. Rather, inherency requires that the claimed property result “***each and every time***” an ordinary artisan combines the

other recited elements. *Endo Pharms. Sols., Inc.*, 894 F.3d at 1382 (emphasis added); *see, e.g., In re Omeprazole Patent Litig.*, 483 F.3d 1364, 1372-1373 (Fed. Cir. 2007).

Extension of time to disease progression does not result “each and every time” a patient is administered the claimed combination. Indeed, the record clearly shows that some patients who were administered rhuMAb HER2 and paclitaxel did not experience extension in time to disease progression. Appx8435 (showing that certain patients treated with rhuMAb HER2 and paclitaxel experienced a shorter TTP than certain patients treated with paclitaxel alone). Phase II studies for rhuMAb HER2 alone, which were discussed in Baselga ’96, likewise reported that the majority of patients did not even have a minor response to treatment. Appx41-47; Appx3670.<sup>9</sup>

Second, extension of TTP is not the “natural result” of elements “explicitly disclosed by the prior art.” *PAR Pharm.*, 773 F.3d at 1196. There was no disclosure in the prior art of treating human patients with rhuMAb HER2, a taxoid, and a third agent.

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<sup>9</sup> As discussed below, Baselga ’96 had many shortcomings that would have prevented an ordinary artisan from drawing reliable conclusions regarding *extension* of TTP. *See infra* pp. 30-31. But the lack of effectiveness in some patients is relevant to the issue of inherency, since failure in even a single patient would defeat a finding of inherency.

Further, even focusing solely on the combination of rhuMAb HER2 and paclitaxel without a third agent, there was no disclosure inherently linking extension of TTP to that combination. The only prior art describing any results of the combination of rhuMAb HER2 and paclitaxel was in preclinical xenograft models, and thus did not involve administration of the treatment to human patients. Accordingly, there was no basis for concluding that extending TTP is the “natural result” of such a combination.

The Board’s reliance on *Santarus, Inc. v. Par Pharm., Inc.*, 694 F.3d 1344 (Fed. Cir. 2012) in these circumstances was misplaced. The Board cited *Santarus* for the proposition that “an obvious formulation cannot become nonobvious simply by administering it to a patient and claiming the result[.]” Appx42 (citing *Santarus*, 694 F.3d at 1354) (alterations in Board’s decision). But *Santarus* involved a single prior art reference that disclosed all elements of the claimed formulation except the resulting serum concentrations listed in the claims. 694 F.3d at 1348. And, critically, “neither party disputed that the blood serum concentrations claimed in *Santarus* were expected in light of the dosages disclosed in the prior art.” *PAR*, 773 F.3d at 1195. In contrast, the claimed extension of TTP does not necessarily or naturally result each time the combination of rhuMAb HER2 and paclitaxel is administered.

Finally, in the absence of any actual evidence of inherency presented by Petitioner, the Board relied on a purported concession from Genentech—namely, that Genentech “admits” that the combination of rhuMAb HER2 and paclitaxel “extends the time to disease progression (‘TTP’) as compared with patients receiving taxoid therapy alone.” Appx41-42. But saying that the combination can achieve that result—as, indeed, Genentech proved in the data reported in its patents—does not mean that it necessarily will or that an ordinary artisan would expect such a result to naturally flow from the combination. An ordinary statement from a patent owner regarding the enablement of its invention does not substitute for the evidence required to meet the demanding standard of proving inherency. That evidence was missing, and the Board’s inherency decision cannot stand.

**B. The Board Erred In Finding The Claimed Efficacy Was Obvious**

The Board’s conclusion that an ordinary artisan would reasonably expect that rhuMAb HER2 in combination with a taxoid would extend TTP in a human patient as compared to a taxoid alone was based on Baselga ’96 and the Taxol PDR ’95. Baselga ’96 reported results from a Phase II clinical trial of rhuMAb HER2 alone, while the Taxol PDR ’95 reported a TTP of Taxol of 3.0 or 4.2 months reported for Taxol in the Physicians’ Desk Reference. The Board reasoned that “because effective amounts of rhuMoAb and paclitaxel were known, one of ordinary skill in the art would have had a reasonable expectation that a



combination of these agents would extend the time to disease progression relative to treatment with paclitaxel alone.” Appx43. This reliance on Baselga ’96’s reported TTP of rhuMAb HER2 was error because it ignored critical information and omissions.

Statements in the prior art must be “read in context.” *Shire LLC v. Amneal Pharms., LLC*, 802 F.3d 1301, 1308 (Fed. Cir. 2015). The Board violated this principle when it read Baselga ’96’s reported TTP of 5.1 months in isolation. First, the Board failed to grapple with the fact that Baselga ’96 measured response and TTP, ***not extension of TTP*** as required by Genentech’s claims. As to TTP, Baselga ’96 included no control arm, and therefore provided no way to draw any conclusions regarding improvement in TTP compared to other patients in the same study. Appx9796-9797; Appx9320 (agreeing that Baselga ’96 included no control). The Board brushed over this difference, stating that Baselga 96 also reported response rate, which is a surrogate for extension of TTP. Appx43. But the record evidence established that an ordinary artisan would have understood that a therapy could reduce tumor size without improving TTP because tumors can shrink and then grow back. Appx9582; Appx9796; Appx10211 (“The proportion of patients whose tumors shrink by at least 50% is the primary endpoint of most phase II trials although the durability of such responses is also of interest ... such trials ... do not determine the ‘effectiveness’ of the treatment.”).

The Board also overlooked the fact that Baselga '96 included in its calculation of TTP only a limited subset of patients: those patients with either a minor response or stable disease, which included *only 16 of the 43 assessable patients*. Appx3670. Baselga '96 *excluded* from the calculation over half of the patients in the study, 22 of the 43 total, who showed progression of disease. In other words, Baselga '96 did not calculate TTP for the entire patient population. Rather, Baselga '96 calculated TTP for only the patients most likely to respond favorably to the treatment, skewing the result upward by excluding from its calculation the patients who showed faster disease progression. Accounting for the patients Baselga '96 excluded, who all had TTP shorter than the median 5.1 months, the TTP is necessarily shorter than the 5.1 months on which the Board relied. Thus, an ordinary artisan could not draw any comparison between the rhuMAb HER2 TTP reported in Baselga '96 and the paclitaxel TTP in the Taxol PDR. Appx10661-10665.

The Board's recitation of principles of combination therapy does not save its analysis. The Board cited a textbook that addresses combinations of two *chemotherapies*, but does not address whether and how to combine therapies involving a novel biologic such as rhuMAb HER2. Appx4803 (discussing "studies of combination chemotherapy"). Rather, the prior art cautioned that "[t]he incorporation of biological agents ... into combination regimens with standard

chemotherapeutic agents offers an important challenge to the medical oncologist since the assumptions for their use likely differ from those for chemotherapeutic agents.” Appx10464. The properties of rhuMAb HER2 were not well-understood: No antibodies had been approved for treatment of solid tumors, no Phase III trial using rhuMAb HER2 had been conducted, there was no known and approved dose for rhuMAb HER2 as a single agent, and the mechanism of action of rhuMAb HER2 was uncertain. Appx10699; Appx3672-3673. Even Petitioner’s expert, Dr. Earhart, was unaware of any publication as of December 1996 applying these principles to combine a chemotherapeutic agent and an antibody. Appx9310. As a result, a skilled artisan would not have simply applied a formula intended for two chemotherapies to rhuMAb HER2. Appx10702.

Finally, the Board improperly dismissed Genentech’s evidence showing that the high failure rate of Phase III clinical trials of 60% supports that an ordinary artisan would not have a reasonable expectation of success. Appx9739-9740. The Board improperly dismissed this evidence, reasoning that the failure rate of clinical trials addresses single compounds, not combination therapies. Appx45. What the Board failed to recognize, however, is that the combination of rhuMAb HER2 and paclitaxel created more, not less, uncertainty, than a single drug trial. RhuMAb HER2 was not yet approved by the FDA, and, in sharp contrast to most Phase III studies which follow Phase II studies of the same treatment, there were no previous

clinical trials testing rhuMAb HER2 and paclitaxel. Appx9749-9750. Indeed, even Petitioner's expert Dr. Earhart testified that although monoclonal antibodies were "exciting" during this time, "it wasn't like it looked like it was going to be smooth sailing [for their] development," Appx9302, and that during this time paclitaxel was still considered a "novel agent." Appx9272.

In sum, the prior art establishes that significant uncertainties existed as to whether the combination of rhuMAb HER2, a taxoid, and a further agent would extend TTP as compared to a taxoid alone. Baselga '96 did not provide reliable information on the TTP of rhuMAb HER2 where it excluded more than half of the patients in its calculation, and the Board's vague references to principles of combination therapy cannot save the analysis. Therefore, the Board's finding that ordinary artisans would reasonably expect the claimed efficacy in the '549 patent claims is not supported by substantial evidence.

### **CONCLUSION**

For the foregoing reasons, the decision of the Board should be vacated and the case should be remanded for further proceedings on Genentech's non-contingent motion to amend. In the alternative, the Board's decision on the original claims should be reversed.

Respectfully submitted,

/s/ Robert J. Gunther, Jr.

ROBERT J. GUNTHER, JR.  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
7 World Trade Center  
250 Greenwich Street  
New York, NY 10007  
(212) 230-8800

THOMAS G. SAUNDERS  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
1875 Pennsylvania Avenue NW  
Washington, DC 20006  
(202) 663-6000

ANDREW J. DANFORD  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
60 State Street  
Boston, MA 02109  
(617) 526-6000

DARALYN J. DURIE  
ADAM R. BRAUSA  
DURIE TANGRI LLP  
217 Leidesdorff Street  
San Francisco, CA 94111  
(415) 362-6666

THOMAS G. SPRANKLING  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
950 Page Mill Road  
Palo Alto, CA 94304  
(650) 858-6000

NORA Q.E. PASSAMANECK  
WILMER CUTLER PICKERING  
HALE AND DORR LLP  
1225 Seventeenth Street  
Suite 2600  
Denver, CO 80202  
(720) 274-3135

July 9, 2019

# **ADDENDUM**

## **TABLE OF CONTENTS**

|   | <b>Page(s)</b> |
|---|----------------|
| Final Written Decision, Paper No. 88 (Oct. 3, 2018) ..... | Appx1-64       |
| U.S. Patent No. 7,892,549 (Exhibit 1001) .....            | Appx65-88      |

[Trials@uspto.gov](mailto:Trials@uspto.gov)  
Tel: 571-272-7822

Paper 88  
Entered: October 3, 2018

UNITED STATES PATENT AND TRADEMARK OFFICE

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BEFORE THE PATENT TRIAL AND APPEAL BOARD

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Celltrion, Inc.  
Petitioner,

v.

GENENTECH, INC.,  
Patent Owner.

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Case IPR2017-01122  
Patent 7,892,549 B2

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Before ZHENYU YANG, CHRISTOPHER G. PAULRAJ, and  
ROBERT A. POLLOCK, *Administrative Patent Judges*.

POLLOCK, *Administrative Patent Judge*.

FINAL WRITTEN DECISION AND RELATED ORDERS

Claims 1–11 and 14–17 Shown to Be Unpatentable  
*35 U.S.C. § 318(a); 37 C.F.R. § 42.73*

Denying Patent Owner's Motion to Amend  
*35 U.S.C. § 316(d); 37 C.F.R. § 42.121*

Denying Patent Owner's Motion to Exclude Evidence  
Denying Petitioner's First and Second Motions to Exclude Evidence  
*37 C.F.R. § 42.64*

Granting-In-Part Parties' Motions to Seal  
*37 C.F.R. § 42.55*



IPR2017-01122  
Patent 7,892,549 B2

## I. INTRODUCTION

This is a Final Written Decision in an *inter partes* review challenging the patentability of claims 1–11 and 14–17 of U.S. Patent No. 7,892,549 B2 (Ex. 1001, “the ’549 patent”). We have jurisdiction under 35 U.S.C. § 6.

Having reviewed the arguments of the parties and the supporting evidence, we find that Petitioner has demonstrated by a preponderance of the evidence that each of the challenged claims is unpatentable.

### A. Procedural History

Petitioner Celltrion, Inc. (“Celltrion”)<sup>1</sup> filed a Petition requesting *inter partes* review of claims 1–11 and 14–17 of the ’549 patent. Paper 2 (“Pet.”). Patent Owner, Genentech, Inc., filed a Preliminary Response to the Petition. Paper 6 (“Prelim. Resp.”). Based on the record then before us, we instituted trial with respect to all challenged claims. Paper 9, 27–28 (“Dec.”).

After institution of trial, Patent Owner filed a Patent Owner Response (Paper 28, “PO Resp.”) and Petitioner filed a Reply to the Patent Owner Response (Paper 45, “Pet. Reply”).

Patent Owner also filed a Contingent Motion to Amend. Paper 26. Petitioner opposed. Paper 42. Patent Owner responded with a Reply in support of its motion (Paper 53); Petitioner further submitted an authorized Sur-Reply (Paper 64).

With respect to technical experts, Petitioner relies on the declarations of Robert Earhart, MD., Ph.D. (Exs. 1002, 1054, 1105); Patent Owner relies on the

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<sup>1</sup> Petitioner further identifies Celltrion Healthcare Co., Ltd. and Teva Pharmaceuticals International GmbH as real parties-in-interest. Paper 10, 2.

IPR2017-01122

Patent 7,892,549 B2

declarations of Robert S. Kerbel, Ph.D. (Exs. 2061, 2143), Dr. Susan Tannenbaum (Exs. 2062, 2144).

Patent Owner filed motions for observations on the depositions of Dr. Earhart (Papers 69, 72), to which Petitioner provides responses (Papers 76, 80).

We heard oral argument on May 18, 2018. A transcript of that proceeding is entered as Paper 85 (“Tr.”).

The parties filed the following motions to exclude evidence. Patent Owner filed one motion to exclude evidence. Paper 59. Petitioner opposed (Paper 72) and Patent Owner submitted a reply in support of its motion (Paper 75). Petitioner filed a first motion to exclude evidence. Paper 61. Patent Owner opposed (Paper 71) and Petitioner submitted a reply in support of its first motion (Paper 80). Petitioner filed a second motion to exclude evidence. Paper 81. Patent Owner opposed (Paper 83) and Petitioner submitted a reply in support of its second motion (Paper 84). Also before us are five unopposed motions to seal pursuant to the Modified Default Standing Protective Order governing this case: Papers 27 and 52 (by Patent Owner) and Papers 44, 47, and 62 (by Petitioner); *see also* Paper 24 (entering Modified Default Standing Protective Order (Exhibit 2036) and granting Patent Owner’s motion to seal Exhibits 2001–2005, 2007, and 2008).

#### B. Related Applications and Proceedings

The ’549 Patent issued from Application No. 10/356,824, filed February 3, 2003, which is a continuation of Application No. 09/208,649, filed Dec. 10, 1998 (the “’649 Application”). U.S. Patent No. 7,846,441 B2 (“the ’441 Patent”) issued from the ’649 Application on December 7, 2010. The ’549 and ’441 Patents claim benefit of priority to Provisional Application No. 60/069,346, filed Dec. 12, 1997 (“the ’346 application”). *See e.g.*, Ex. 1001, (21), (63) (60), 1:4–9.

IPR2017-01122  
Patent 7,892,549 B2

In addition to this proceeding, Petitioner has challenged claims 1–14 of the related '441 Patent in copending IPR2017-01121. Petitioner has also filed IPR2017-01139 and IPR2017-01140 involving claims of U.S. Patent Nos. 6,627,196 and 7,371,379, respectively. These two patents are not in the chain of priority of the '549 and '441 Patents but involve subject matter similar to that at issue here.

The '549, '441, '196, and '379 Patents are also the subject of pending *inter partes* reviews, IPR2017-00737, IPR2017-00731, IPR2017-00804, and IPR2017-00805, respectively, brought by Hospira, Inc. (“Hospira”).<sup>2</sup> With respect to the '549 Patent, we refer herein to our Decision to institute trial in IPR2017-00737 as the “Hospira Decision.” *See Hospira, Inc. v. Genentech, Inc.*, Case IPR2017-00737 (PTAB July 27, 2017) (Paper 19).

We issue concurrently our Decisions in IPR2017-00731, IPR2017-00737, IPR2017-01139, IPR2017-01140, IPR2017-01121, IPR2017-00804, and IPR2017-00805.

Patent Owner identifies the following District Court actions, “that relate or may relate to U.S. Patent Application No. 10/356,824, which issued as U.S. Patent No. 7,892,549:” *Celltrion, Inc. v. Genentech, Inc.*, No. 18-cv-00274 (N.D. Cal.) and *Celltrion, Inc. v. Genentech, Inc.*, No. 18-cv-00095 (D. Del.). Paper 33, 2.

### C. The '549 Patent and Relevant Background

According to the Specification, 25% to 30% of human breast cancers overexpress a 185-kD transmembrane glycoprotein receptor (p185<sup>HER2</sup>), also known as HER2 (human epidermal growth factor receptor-2) or ErbB2. Ex. 1001,

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<sup>2</sup> Hospira also challenged claims of the '549 and '441 Patents in IPR2017-00739 and IPR2018-00016, respectively, which we denied. *See* IPR2017-00739, Paper 16; IPR2018-00016, Paper 25.

IPR2017-01122

Patent 7,892,549 B2

1:21–32, 5:16–21. These HER2-positive cancers are associated with poor prognoses and resistance to many chemotherapeutic regimens including anthracyclines (e.g., doxorubicin or epirubicin). *Id.* at 3:43–52; 4:11–12, and 11:41–45. Conversely, patients with HER2-positive cancers are three times more likely to respond to treatment with taxanes than those with HER2 negative tumors. *Id.* at 3:52–56 (citing Baselga '97 (Ex. 1007)).

Although “ErbB2 overexpression is commonly regarded as a predictor of poor prognosis,” “a humanized version of the murine anti-ErbB2 antibody 4D5, referred to as rhuMAb HER2 or HERCEPTIN®<sup>3</sup> has been clinically active in patients with ErbB2-overexpressing metastatic breast cancers that had received extensive prior anti-cancer therapy.” Ex. 1001, 3:35–61 (citing Baselga '96 (Ex. 1020)).<sup>4</sup> Anti-ErbB2 4D5 antibodies also “enhance the activity of paclitaxel (TAXOL®) and doxorubicin against breast cancer xenographs in nude mice injected with BT-474 human breast adenocarcinoma cells, which express high levels of HER2.” *Id.* at 3:56–61 (citing Baselga Abstract 53 (Ex. 1019)).<sup>5</sup>

According to the Specification,

The present invention concerns the treatment of disorders characterized by overexpression of ErbB2, and is based on the recognition that while treatment with anti-ErbB2 antibodies markedly enhances the clinical benefit of the use of chemotherapeutic agents in

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<sup>3</sup> As Patent Owner notes, “HERCEPTIN® is the tradename for the commercial product of the humanized antibody, trastuzumab.” Paper 26, 3 fn.2.

<sup>4</sup> Baselga et al., *Phase II Study of Weekly Intravenous Recombinant Humanized Anti-p195<sup>HER2</sup> Monoclonal Antibody in Patients with HER2/neu-Overexpressing Metastatic Breast*, Cancer, 14(3) J. Clin. Oncol. 737–44 (1996). Ex. 1020.

<sup>5</sup> Baselga et al., *Anti Her2 Humanized Monoclonal Antibody (Mab) Alone And In Combination With Chemotherapy Against Human Breastcarcinoma Xenografts*, 15 PROC. AM. SOC'Y. CLIN. ONCOL. 63, Abstract 53 (1994) (designated “Baslega '94” in IPR2017-00737). Ex. 1019.

IPR2017-01122

Patent 7,892,549 B2

general, a syndrome of myocardial dysfunction that has been observed as a side-effect of anthracycline derivatives is increased by the administration of anti-ErbB2 antibodies.

*Id.* at 3:65–4:5.

The '549 Patent, thus, relates to the treatment of breast cancers that overexpress HER2/ErbB2 “comprising administering a therapeutically effective amount<sup>[6]</sup> of a combination of an anti-ErbB2 antibody and a chemotherapeutic agent other than an anthracycline derivative, e.g. doxorubicin or epirubicin, in the absence of an anthracycline derivative to the human patient.” *Id.* at 4:6–13. In some embodiments, the anti-ErbB2 antibody of the combination is Herceptin® and the chemotherapeutic agent “is a taxoid, such as TAXOL® (paclitaxel) or a TAXOL® derivative.” *Id.* at 4:23–25. The combination may further include one or more additional anti-ErbB2 antibodies, “antibodies which bind to the EGFR . . . ErbB3, ErbB4, or vascular endothelial factor (VEGF),” “one or more cytokines,” or “a growth inhibitory agent.” *Id.* at 11:4–40 (defining “chemotherapeutic agent” and “growth inhibitory agent”), 23:60–24:5, and 25:20–34.

The '549 Patent also provides an Example disclosing the conduct and results of a clinical trial involving 469 women with metastatic HER2-positive breast cancer. *Id.* at 26:34–30:25. All patients were treated with one of two chemotherapy regimens (CRx) designated either “AC” for anthracycline (doxorubicin or epirubicin) and cyclophosphamide, or “T” for Taxol (paclitaxel). *See id.* at 28:5–47; 29:13–30:12. Half of the patients were also treated with the anti-ERbB2 antibody Herceptin, designated “H.” *Id.* The Specification discloses

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<sup>6</sup> The Specification defines a “therapeutically effective amount” of the combination as “an amount having an antiproliferative effect,” which can be “measured by assessing the time to disease progression (TTP) or determining the response rates (RR).” *Id.* at 10:41–50.

IPR2017-01122

Patent 7,892,549 B2

that “[a]t a median follow-up of 10.5 months, assessments of time to disease progression (TTP in months) and response rates (RR) showed a significant augmentation of the chemotherapeutic effect by HERCEPTIN®, without increase in overall severe adverse events (AE).” *Id.* at 29:13–18. In addition, “[a] syndrome of myocardial dysfunction similar to that observed with anthracyclines was reported more commonly with a combined treatment of AC-H (18% Grade  $\frac{3}{4}$ ) than with AC alone (3%), T (0%), or T+H (2%).” *Id.* at 30:13–16. According to the inventors:

These data indicate that the combination of anti-ErbB2 antibody treatment with chemotherapy markedly increases the clinical benefit, as assessed by response rates and the evaluation of disease progression. However, due to the increased cardiac side-effects of doxorubicin or epirubicin, the combined use of anthracyclines with anti-ErbB2 antibody therapy is contraindicated. The results, taking into account risk and benefit, favor the combined treatment with HERCEPTIN® and paclitaxel (TAXOL®).

*Id.* at 30:17–25.

#### D. Challenged Claims and Reviewed Ground of Unpatentability

We instituted trial on the sole Ground set forth in the Petition, that claims 1–11 and 14–17 are unpatentable under 35 U.S.C. § 103 based on the combination of Baselga 1996, Seidman 1996,<sup>7</sup> Pegram,<sup>8</sup> 1995 TAXOL PDR,<sup>9</sup> and the

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<sup>7</sup> Seidman et al., *Her-2/neu Over-Expression and Clinical Taxane Sensitivity: A Multivariate Analysis in Patients with Metastatic Breast Cancer (MBC)*, 15 PROC. AM. SOC’Y. CLIN. ONCOL. 104, Abstract 80 (1996). Ex. 1011.

<sup>8</sup> Pegram et al., *Phase II Study of Intravenous Recombinant Humanized Anti-p185 HER-2 Monoclonal Antibody (rhuMAB HER-2) Plus Cisplatin in Patients with HER-2/NEU Overexpressing Metastatic Breast Cancer*, 14 PROC. AM. SOC’Y. CLIN. ONCOL 106, Abstract 124. Ex. 1022.

<sup>9</sup> TAXOL (paclitaxel) for Injection Concentrate, in PHYSICIAN’S DESK REFERENCE, 682–85 (49<sup>th</sup> ed. 1995). Ex. 1012.

IPR2017-01122

Patent 7,892,549 B2

knowledge of one of ordinary skill in the art. Dec. 27–28; *see* Pet. 24.

Claims 1, 5, and 16 are independent. Claim 1, reproduced below, requires “administering a combination” of three agents—an anti-ErbB2 antibody, a taxoid, and “a further growth inhibitory agent”—“in an amount effective to extend the time to disease progression:”

1. A method for the treatment of a human patient with breast cancer that overexpresses ErbB2 receptor, comprising administering a combination of an antibody that binds ErbB2, a taxoid, and a further growth inhibitory agent to the human patient in an amount effective to extend the time to disease progression in the human patient, wherein the antibody binds to epitope 4D5 within the ErbB2 extracellular domain sequence.

Independent claim 16 is similar to claim 1, but further includes a negative limitation requiring the administration of an anti-ErbB2 antibody, a taxoid, and a further growth inhibitory agent “in the absence of an anthracycline derivative.” Independent claim 5 recites “administering an effective amount of a combination” of three agents similar to those of claims 1 and 16, wherein the antibody binds to the 4D5 epitope of ErbB2, the taxoid is paclitaxel, and the third element is broadly described as a “therapeutic agent.”

Patent Owner does not separately argue the patentability of claims 2–4, 6–11, 14, 15, or 17.

## II. ANALYSIS

### A. Principles of Law

A claim is unpatentable under 35 U.S.C. § 103(a) if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a

IPR2017-01122

Patent 7,892,549 B2

person having ordinary skill in the art to which that subject matter pertains.<sup>10</sup> *KSR Int’l Co. v. Teleflex Inc.*, 550 U.S. 398, 406 (2007). The question of obviousness is resolved based on underlying factual determinations including: (1) the scope and content of the prior art; (2) any differences between the claimed subject matter and the prior art; (3) the level of ordinary skill in the art; and (4) objective evidence of nonobviousness, if present. *Graham v. John Deere Co.*, 383 U.S. 1, 17–18 (1966).

“[T]he [obviousness] analysis need not seek out precise teachings directed to the specific subject matter of the challenged claim, for a court can take account of the inferences and creative steps that a person of ordinary skill in the art would employ.” *KSR*, 550 U.S. at 418. Moreover, “any need or problem known in the field of endeavor at the time of invention and addressed by the patent can provide a reason for combining the elements in the manner claimed.” *Id.* at 420.

Accordingly, a party that petitions the Board for a determination of unpatentability based on obviousness must show that “a skilled artisan would have been motivated to combine the teachings of the prior art references to achieve the claimed invention, and that the skilled artisan would have had a reasonable expectation of success in doing so.” *In re Magnum Oil Tools Int’l, Ltd.*, 829 F.3d 1364, 1381 (Fed. Cir. 2016) (citations omitted).

We analyze the instituted ground of unpatentability in accordance with these principles.

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<sup>10</sup> The Leahy-Smith America Invents Act, Pub. L. No. 112-29, 125 Stat. 284 (2011) (“AIA”), amended 35 U.S.C. §§ 102 and 103. Because the challenged claims of the ’405 patent have an effective filing date before the effective date of the applicable AIA amendments, throughout this Final Written Decision we refer to the pre-AIA versions of 35 U.S.C. §§ 102 and 103.



IPR2017-01122

Patent 7,892,549 B2

**B. Person of Ordinary Skill in the Art**

Patent Owner argues that we should apply the same definition of a person of ordinary skill as set forth in the Hospira Petition, which also involves the '549 Patent. Prelim. Resp. 37; PO Resp. 33. In that case, we adopted Petitioner Hospira's definition of one of ordinary skill as "a clinical or medical oncologist specializing in breast cancer with several years of experience with breast cancer research or clinical trials." Hospira Decision at 8–9 (quoting IPR2017-00737 Pet. 6). In the present Petition, however, Celltrion argues that a person of ordinary skill in the art as of the effective filing date of the '549 patent "would have been an M.D. with subspecialty training in oncology and substantial experience treating breast cancer patients and/or a Ph.D. with substantial experience in researching and developing oncologic therapies." Pet. 43 (citing Ex. 1002, ¶ 29). According to Petitioner, "[s]uch an individual would also have had substantial experience in the design and/or implementation of clinical trials for breast cancer treatments, and/or an active research role relating to breast cancer treatments." *Id.*

For the reasons set forth in our institution Decision, we agree with Patent Owner. Dec. 8–9. Petitioner has not explained why its proposed definition better defines the level of ordinary skill in the art, nor why its alternative definition would have any bearing on the outcome of the present case. We do not discern an appreciable difference in the parties' respective definitions of the level of ordinary skill in the art. Indeed, both parties contend that a person of ordinary skill in the art would have had experience with breast-cancer research and treatment. Accordingly, we adopt Patent Owner's definition of the level of ordinary skill in the art as "a clinical or medical oncologist specializing in breast cancer with several years of experience with breast cancer research or clinical trials." *See also* Hospira Decision, 8–9 (defining the skill level the same way); Ex. 2020 ¶ 78

IPR2017-01122

Patent 7,892,549 B2

(implicitly adopting same definition). In any event, as Petitioner does not explain why its alternative definition would have any bearing on the outcome of the present case, and as we discern no appreciable difference in the parties' definitions, we note our findings and conclusions would be the same regardless of which definition were adopted. *See* PO Resp. 33 (arguing that the challenged claims would not have been obvious under either parties proposed definition).

We further note that the prior art itself demonstrates the level of skill in the art at the time of the invention. *See Okajima v. Bourdeau*, 261 F.3d 1350, 1355 (Fed. Cir. 2001) (explaining that specific findings regarding ordinary skill level are not required “where the prior art itself reflects an appropriate level and a need for testimony is not shown”) (quoting *Litton Indus. Prods., Inc. v. Solid State Sys. Corp.*, 755 F.2d 158, 163 (Fed. Cir. 1985)).

### C. Claim Construction

In an *inter partes* review, claim terms in an unexpired patent are interpreted according to their broadest reasonable construction in light of the specification of the patent in which they appear. 37 C.F.R. § 42.100(b); *Cuozzo Speed Techs., LLC v. Lee*, 136 S. Ct. 2131, 2144–46 (2016) (upholding the use of the broadest reasonable interpretation standard). “Under a broadest reasonable interpretation, words of the claim must be given their plain meaning, unless such meaning is inconsistent with the specification and prosecution history.” *Trivascular, Inc. v. Samuels*, 812 F.3d 1056, 1062 (Fed. Cir. 2016). Any special definitions for claim terms must be set forth with reasonable clarity, deliberateness, and precision. *In re Paulsen*, 30 F.3d 1475, 1480 (Fed. Cir. 1994).

#### 1. “administering a combination”

In IPR2017-00737 (involving claims 1–17 of the same patent), we initially adopted Patent Owner’s unopposed definition of “administering a combination” as

IPR2017-01122

Patent 7,892,549 B2

requiring “a single treatment regimen in which the patient receives all drugs that are part of the claimed combination.” Hospira Decision, 10. Patent Owner subsequently recast its proposed definition “to mean that the drugs are administered as part of the same treatment regimen,” which we adopted.

IPR2017-00737, PO Resp. 37, IPR2017-00737 Final Decision, 11–12. Also in that proceeding, we noted that Patent Owner’s two definitions were interchangeable, as they would be here. *See* IPR2017-00737 Final Decision, 12. In the interests of clarity and consistency, we again define “administering a combination” to mean that the drugs are administered as part of the same treatment regimen.

2. “*an amount effective to extend the time of disease progression*” and “*an effective amount*”

Independent claims 1 and 16 require administering a combination of an anti-ErbB2 antibody, a taxoid, and a further agent, “in an amount effective to extend the time to disease progression [TTP] in the human patient.” Claim 5, the remaining independent claim before us, similarly recites administering the three-part combination to a human patient in “an effective amount.” To the extent that these terms may differ in scope, neither party contends that any difference affects the patentability analysis and we consider them together.

In our Decision to Institute, we construed “an amount effective to extend the time to disease progression in the human patient” in independent claims 1 and 16 as an amount sufficient to extend the time to disease progression in a human patient having breast cancer that overexpresses ErbB2 receptor *as compared to one receiving no treatment*. Dec. 11–13. We also construed the language “an effective amount” of independent claim 5 as encompassing “an amount effective to extend the time to disease progression in the human patient” and, thus, similarly indicating a comparison to an untreated patient. *See id.*

IPR2017-01122

Patent 7,892,549 B2

Patent Owner disagrees with our construction, contending that the proper comparator in both claim terms is not an untreated patient, but to a patient treated with taxoid alone. PO Resp. 34–37. In particular, Patent Owner argues that comparison to an untreated patient “is not consistent with the specification as understood by a POSA,” and “makes no sense in the context of a disease like breast cancer.” *Id.* at 34–35. Yet this is precisely the comparison Applicants made to obtain allowance of the challenged claims.

“A patent’s specification, together with its prosecution history, constitutes intrinsic evidence to which the [the Board] gives priority when it construes claims.” *Knowles Elecs. LLC v. Cirrus Logic, Inc.*, 883 F.3d 1358, 1361 (Fed. Cir. 2018). “The purpose of consulting the prosecution history in construing a claim is to exclude any interpretation that was disclaimed during prosecution.” *Chimie v. PPG Indus., Inc.*, 402 F.3d 1371, 1384 (Fed. Cir. 2005) (internal quotation marks omitted). Prosecution disclaimer

requires that the alleged disavowing actions or statements made during prosecution be both clear and unmistakable. Thus, when the patentee unequivocally and unambiguously disavows a certain meaning to obtain a patent, the doctrine of prosecution history disclaimer narrows the meaning of the claim consistent with the scope of the claim surrendered. Such disclaimer can occur through amendment or argument. . . . [and] includes all express representations made by or on behalf of the applicant to the examiner to induce a patent grant . . . includ[ing] amendments to the claims and arguments made to convince the examiner.

*Aylus Networks, Inc. v. Apple Inc.*, 856 F.3d 1353, 1359 (Fed. Cir. 2017) (internal citations and quotations omitted); *see Arendi S.A.R.L. v. Google LLC*, 882 F.3d 1132, 1135–36 (Fed. Cir. 2018). Those conditions are satisfied here.

The claim language “an amount effective to extend the time to disease progression” implies that time to disease progression is extended in relation to

IPR2017-01122

Patent 7,892,549 B2

some metric, but none of the challenged claims expressly identifies the intended comparator. The Examiner addressed this facial ambiguity during the prosecution leading to the issuance of the '549 Patent. In particular, during the prosecution of the '649 Application (the direct predecessor to the '842 Application, from which the '549 Patent issued), the Examiner rejected then-pending claims under 35 U.S.C. § 112, second paragraph because:

The phrase “extend the time to disease progression” . . . is a relative term which renders the claim[s] indefinite. The term “extend time to disease progression” is not defined by the claim, the specification does not provide a standard for ascertaining the requisite degree, and one of ordinary skill in the art would not be reasonably apprised of the scope of the invention. Specifically, it is never set forth what the extension of time to disease progress is relative to, for example, is the extension of time to disease progress relative to untreated patients? Patients who received antibody or taxoid alone? Patients who received antibody and an anthracycline?

Ex. 3001, 400-402 (OA dated 7/17/01).<sup>11</sup> In response, Applicants asserted that:

the expression[] “extend the time to disease progression”. . . [is] clear from the specification (see, in particular, page 15, lines 15-17; and pages 42-43) and would be readily understood by the skilled oncologist. Clearly, the combination of anti-ErbB2 antibody and taxoid is administered in an amount effective to extend the time to disease progression relative to an untreated patient.

*Id.* at 416 (Response dated 1/17/2001); *see also* Ex. 3001-1, 19, (15:12–17), 46–47 (42–43). The Examiner withdrew the rejection in the next office action, stating that “[a]ll claims are allowable.” *Id.* at 624 (OA dated 3/27/2002) (suspending prosecution due to potential interference); *see also id.* at 634–39 (OA dated

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<sup>11</sup> Excerpts of prosecution history of US Application No. 09/208,649. Citations refer to pages of the exhibit overall rather than to the native pagination.

IPR2017-01122

Patent 7,892,549 B2

8/12/2003) (new grounds of rejection not relating to the phrase “extend the time to disease progression”).

Accordingly, Applicants overcame the § 112 rejection by providing an express definition of the term “extend the time to disease progression” as meaning “relative to an untreated patient.” Our construction reflects Applicants’ choice. *See In re Paulsen*, 30 F.3d at 1480 (holding an applicant may choose to be his own lexicographer).

Patent Owner contends that “the clinical trial results reported in the ’441 specification measure efficacy of the combination of an anti-ErbB2 antibody (rhuMAb HER2) with a taxoid (paclitaxel) against a control arm of paclitaxel alone,” whereas “[t]here is no data in the patent comparing the TTP of patients treated with an anti-ErbB2 antibody and a taxoid against an untreated patient.” PO Resp. 34–35. That may well be the case; yet, it does not render our construction inconsistent with the Specification of the ’441 patent. As Dr. Tannenbaum, an expert for Patent Owner, explains, “cancer generally continues to progress without treatment.” Ex. 2062 ¶ 133. As a result, an ordinary artisan would have understood that, even without any explicit disclosure in the ’549 Patent, administering the claimed combinations would extend the TTP as compared to untreated patients. *See e.g.*, Ex. 1002 ¶ 111 (Dr. Earhart indicating that the choice of claim construction does not impact the obviousness analysis); Ex. 1054 (Dr. Earhart testifying that “a person of ordinary skill would have had a reasonable expectation that a combination treatment with paclitaxel and trastuzumab would extend the time to disease progression relative to treatment with paclitaxel and relative to no treatment”); *id.* ¶ 24 (same analysis with respect to proposed amended claims).

IPR2017-01122

Patent 7,892,549 B2

With respect to the prosecution history, Dr. Tannenbaum testifies that, “in context,” Applicants used the term “untreated patient” to refer to “a patient that had not received the combination therapy, but instead received paclitaxel alone.” Ex. 2062 ¶ 138. We do not find Dr. Tannenbaum’s argument persuasive.

The Examiner asked Applicants to choose from various potential meanings for the claim language: “is the extension of time to disease progress[ion] relative to untreated patients? Patients who received antibody or taxoid alone? Patients who received antibody and an anthracycline?” Ex. 3001, 401–402. Despite being presented with the option of selecting “taxoid alone” as the comparator, Applicant did not do choose that option. Applicant instead specifically excluded that possibility. *Id.* at 416 (stating “[c]learly, the combination of anti-ErbB2 antibody and taxoid is administered in an amount effective to extend the time to disease progression **relative to an untreated patient**”) (emphases added). Indeed, Dr. Tannenbaum admitted that much at her deposition in the related Hospira case, agreeing that “there can be no confusion” that Applicants were “choosing the comparator untreated patients rather than taxoid alone.” *See* IPR20117-00737 Ex. 1087, 225:15–226:13.

For the reasons set forth above, we maintain that the proper analysis of the claim language “in an amount effective to extend the time to disease progression [TTP] in the human patient” and administering the three-part combination to a human patient in “an effective amount” involves comparing the claimed combination treatments to no treatment. To the extent Patent Owner is correct that our construction “makes no sense in the context of a disease like breast cancer” (PO Resp, 35), Applicants chose this definition “with reasonable clarity, deliberateness, and precision,” and obtained the ’549 Patent only after doing so. *See In re Paulsen*, 30 F.3d at 1480. Under such circumstances, we must give the

IPR2017-01122

Patent 7,892,549 B2

term the construction the applicant set out, even if such construction would lead to a “nonsensical result.” *Source Vagabond Sys. Ltd. v. Hydrapak, Inc.*, 753 F.3d 1291, 1301 (Fed. Cir. 2014).

#### D. Asserted Ground of Unpatentability

Petitioner challenges claims 1–11 and 14–17 as unpatentable under 35 U.S.C. § 103 based on the combination of Baselga 1996, Seidman 1996, Pegram, 1995 TAXOL PDR, and the knowledge of one of ordinary skill in the art, evidenced, in part, by Baselga Abstract 53, Baselga Abstract 2262,<sup>12</sup> and Seidman 1995.<sup>13</sup> *See* Pet. 43–53; Pet Reply 4–22. Patent Owner opposes.<sup>14</sup> PO Resp. 37–54.

We begin with an overview of the above-recited references.

##### 1. Overview of Baselga 1996 (Ex. 1020)

Baselga 1996 teaches that “[i]n preclinical studies . . . rhuMAb HER2 markedly potentiated the antitumor effects of several chemotherapeutic agents, including cisplatin, doxorubicin, and paclitaxel, without increasing their toxicity.” Ex. 1020 at 9 (citing Baselga Abstract 53). As a result, “[l]aboratory studies of the

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<sup>12</sup> Baselga *et al.*, *Antitumor Activity of Paclitaxel in Combination with Anti-growth Factor Receptor Monoclonal Antibodies in Breast Cancer Xenografts*, 35 PROC. AM. ASS’N FOR CANCER RES. 380, Abstract 2262. Ex. 1021.

<sup>13</sup> Seidman *et al.*, *Memorial Sloan-Kettering Cancer Center Experience with Paclitaxel in the Treatment of Breast Cancer*, 22(5) Suppl. 12 SEMINARS ONCOLOGY 108–16. Ex. 1010.

<sup>14</sup> Although Patent Owner objects to Petitioner’s reliance of references other than Baselga 1996, Seidman 1996, Pegram, 1995 TAXOL PDR (PO Resp. 37, n.12.) to establish the knowledge of one of ordinary skill in the art, “it is permissible, and sometimes even necessary, to establish such background knowledge by pointing to other prior art.” *Rovalma, S.A. v. Bohler-Edelstahl GmbH & Co. KG*, 856 F.3d 1019, 1027 n.1 (Fed. Cir. 2017) (citations omitted).



IPR2017-01122

Patent 7,892,549 B2

mechanism of this effect and clinical trials of such combination therapy are currently in progress.” *Id.*

Baselga 1996 further teaches that after successful experiments in mouse models, a humanized version of the 4D5 anti-ErbB2 antibody, rhuMAb HER2, was used in a phase II clinical trial for patients with metastatic breast cancer that overexpressed HER2. *Id.* at 3–4. “[P]atients were selected to have many sites of metastatic involvement, one of the most dire prognostic characteristics regarding response to therapy.” *Id.* at 7. Of the 46 patients enrolled, 82.6% had received at least one regimen for metastatic disease, and 63% had received two or more regimens. *Id.* at 5.

Patients were administered 10 weekly doses of rhuMAb HER beginning with a 250 mg loading dose, and 100 mg doses thereafter. *Id.* at 4. “Adequate pharmacokinetic levels of rhuMAb HER2 were obtained in 90% of the patients.” *Id.* at 3. “Treatment with rhuMAb HER2 was remarkably well tolerated.” *Id.* at 5. “Toxicity was minimal and no antibodies against rhuMAb HER2 were detected in any patients.” *Id.* at 3.

“37% of patients achieved minimal responses or stable disease.” *Id.* at 7. “Objective responses were seen in five of 43 assessable patients, and included one complete remission and four partial remissions” for an overall response rate of 11.6%. *Id.* at Abstract; *see id.* at 3. Baselga 1996 predicts “that the percentage of patients who show objective tumor regression to rhuMAb HER2 will be higher when patients with less extensive breast cancer are treated, since laboratory studies have shown that the response to antireceptor antibodies is greater with lower tumor burden.” *Id.* at 7.

“Time to tumor progression was calculated from the beginning of therapy to progression,” and “[t]he median time to progression for the patients with either

IPR2017-01122

Patent 7,892,549 B2

minor or stable disease was 5.1 months.” *Id.* at 4, 6. Baselga 1996 notes that, in contrast to many anticancer drugs, rhuMAB HER2 elicits cytostatic growth arrest rather than cell death in laboratory studies. *See id.* at 7. Accordingly, the authors posit that “stable disease may be an authentic reflection of the biologic action of [rhuMAB HER2]” and “[t]he unusually long durations of minimal responses and stable disease seen in [the] trial” may be indicative of the cytostatic effects of the antibody. *Id.*

## 2. Overview of Seidman 1996 (Ex. 1011)

Seidman 1996 analyzes tissue samples from 126 patients with metastatic breast cancer (MBC) who received single-agent taxane treatment (paclitaxel or docetaxel). Ex. 1011. Of the 51 of these patients determined to be HER2 positive, 58.8% responded to taxane treatment, as compared to only 38.7% of the 75 patients that did not overexpress HER2. *Id.* Seidman concludes that “HER2 overexpression [sic] in MBC seems to confer sensitivity rather than resistance to taxanes,” and although HER2 overexpression generally correlates with a poor prognosis, “stratified analysis controlling for confounding variables demonstrated the value of HER2 status in predicting good taxane response.” *Id.*

## 3. Overview of Pegram 1995 (Ex. 1022)

Pegram 1995 reports on a phase II clinical trial of patients with HER2 positive metastatic breast cancer treated with a combination of cisplatin and rhuMAB HER2 (250 mg loading dose followed by 100 mg weekly doses for 8 weeks). Ex. 1022; *see* Ex. 1002 ¶¶ 62–65. Of the 36 patients evaluated, one had a complete response and 7 had partial responses. *Id.* According to the authors:

The toxicity profile was that expected from [cisplatin], and there were no acute serious adverse events recorded following treatment with rhuMAB HER-2. The use of rhuMAb HER-2 plus [cisplatin] in patients with HER2/*neu* overexpressing MBC resulted in response rates

IPR2017-01122

Patent 7,892,549 B2

above that expected from [cisplatin] alone, and the combination showed no apparent increase in toxicity.

*Id.*

Pegram 1995 also notes by way of background that, in Phase I studies, “rhMAB HER-2 has no substantial toxicity at any dose level and localizes to malignant cells overexpressing the HER-2 receptor protein. In preclinical studies, therapy with this antibody plus cisplatin (CDDP) elicits a synergistic and cytocidal effect on tumor cells which express p185HER-2/*neu*.” *Id.*

4. *Overview of 1995 Taxol PDR (Ex. 1012)*

According to 1995 TAXOL PDR, paclitaxel “is indicated for the treatment of breast cancer after failure of combination chemotherapy for metastatic disease or relapse within 6 months of adjuvant chemotherapy. Prior therapy should have included an anthracycline unless clinically contraindicated.” Ex. 1012, 6. “For patients with carcinoma of the breast, TAXOL at a dose of 175 mg/m<sup>2</sup> administered intravenously over 3 hours every three weeks has been shown to be effective after failure of chemotherapy for metastatic disease or relapse within 6 months of adjuvant chemotherapy.” *Id.* at 8. The 1995 TAXOL PDR further discloses that when used in combination with cisplatin, “myelosuppression was more profound when TAXOL was given after cisplatin than with the alternate sequence.” *Id.* at 6.

5. *Overview of Baselga Abstract 53 (Ex. 1019)*

Baselga Abstract 53 (cited in Baselga 1996) describes xenograft studies in which BT-474 HER2 overexpressing human breast cancer cells were injected into nude mice followed by treatment with humanized 4D5-antibody alone, or in combination with various chemotherapeutic agents. Ex. 1019, 4. Whereas either the antibody or paclitaxel alone produced 35% tumor growth inhibition, the

IPR2017-01122

Patent 7,892,549 B2

combination treatment resulted in “major antitumor activity with 93% inhibition of growth” without increasing toxicity. *Id.* In addition, whereas doxorubicin alone resulted in 27% growth inhibition in this model, the combination of doxorubicin and antibody resulted in 70% growth inhibition. *Id.*

According to Baselga Abstract 53, [t]hese observations suggest that dual insults to cell cycle transversal through checkpoints (Mab-mediated growth factor deprivation, and drug mediated damage to DNA or tubulin) may activate cell death in tumor cells which can survive either treatment given singly. *Id.* The authors conclude “anti-HER2 MAbs can eradicate well established tumors and enhance the activity of paclitaxel and doxorubicin against human breast cancer xenografts. *Id.*

6. *Overview of Baselga Abstract 2262 (Ex. 1021)*

Baselga Abstract 2262 provides additional details regarding the work reported in Baselga Abstract 53. *See* Ex. 1002 ¶ 53 & n.16. According to Baselga Abstract 2262:

The combined treatment with paclitaxel plus 4D5 resulted in a major antitumor activity with 93% inhibition of growth. This result was markedly better than doxorubicin plus 4D5 (70% inhibition). Thus, equipotent doses of paclitaxel and doxorubicin differed in their combined effect with ARMAs, which suggests synergy between paclitaxel and 4D5. ARMAs did not increase the toxicity of paclitaxel in animals as determined by animal survival and weight loss. The antitumor effects of paclitaxel can be markedly enhanced by the addition of ARMAs.

Ex. 1021.

7. *Overview of Seidman 1995 (Ex. 1010)*

Siedman 1995 is a review article regarding the clinical use and laboratory investigations of paclitaxel, “the most important new cytotoxic agent to be introduced for the management of breast cancer in many years.” Ex. 1010, 1.

IPR2017-01122

Patent 7,892,549 B2

Siedman 1995 reports that in a phase II trial for metastatic breast cancer, paclitaxel monotherapy showed “significant antitumor activity in patients with minimal prior treatment.” Ex. 1010, 2. Subsequent investigation of paclitaxel in patients who had previously been treated with anthracyclines also showed anti-tumor activity and a “lack of significant cross-resistance between paclitaxel and doxorubicin.” *Id.* at 2–3, Fig. 1. Seidman 1995 further discusses the development of optimal dosing schedules for paclitaxel therapy (*id.* at 3–4) and the development of combination therapies of paclitaxel, with doxorubicin, cisplatin, and trastuzumab (*id.* at 4–5).

Referencing Baselga Abstract 2262, among others, Seidman 1995 states that “[s]triking antitumor effects are observed when paclitaxel is given in human breast cancer xenografts in combination with . . . anti-HER-2 MoAbs. This strong synergy is achieved with no increased toxicity in the animal model.” *Id.* at 5. “[t]hese data provide a lead for translation into the clinic. Indeed, future clinical trials combining paclitaxel with anti-growth factor receptor MoAbs [e.g., rhuMAB HER2] are being planned.” *Id.*

#### E. Analysis of Asserted Ground

Petitioner has provided a reasoned, claim-by-claim explanation for the basis of its contention that claims 1–11 and 14–17 would have been obvious over the combination of Baselga 1996, Seidman 1996, Pegram 1995, and 1995 TAXOL PDR, in view of the knowledge of a person of ordinary skill in the art. Pet. 24–75. Petitioner asserts that one of ordinary skill would have been “motivated to combine trastuzumab, cisplatin, and paclitaxel based on the dire need for treatments of HER2-positive breast cancer,” which was “notoriously difficult to treat because HER2-positive breast cancer frequently did not respond to traditional anti-cancer

IPR2017-01122

Patent 7,892,549 B2

treatments.” *Id.* at 45 (citing Ex. 1002 ¶¶ 119–122, Ex. 1020, 837; Ex. 1001, 3:41–50). As articulated by Petitioner’s expert, Dr. Earhart:

Particularly for the population of metastatic HER2+ breast cancer patients, which typically had a worse prognosis than other cancer patients . . . a person of ordinary skill in the art would have been interested in testing combinations with any drug that had proven efficacy for metastatic HER2+ breast cancer. Baselga 1996, Pegram 1995, and Seidman 1996 respectively report the clinical efficacy of trastuzumab, trastuzumab/cisplatin, and paclitaxel in the metastatic HER2+ breast cancer population, and therefore provided a strong motivation to test those drugs in combination in human metastatic HER2+ breast cancer patients.

Ex. 1002 ¶ 119.

Petitioner, thus, points to Baselga 1996 as teaching that the rhuMAb HER2 antibody “was clinically effective in patients with advanced metastatic HER2-positive breast carcinoma, was ‘remarkably well tolerated,’ and lacked ‘significant toxicity,’ even though the patients had ‘dire prognostic characteristics’ based on the extensive metastasis of their cancers and prior failures with other treatments.” Pet. 43–44 (citing Ex. 1020, 7). Petitioner argues that before the priority date of the challenged claims, an ordinary artisan “would have been motivated to pursue combination therapies that incorporate trastuzumab . . . in combination with drugs that had shown broad efficacy against all types of metastatic cancer.” *Id.* at 44 (citing Ex. 1002 ¶¶ 119–121). As such, Petitioner notes that Baselga 1996 discloses ongoing clinical trials of trastuzumab in combination with each of paclitaxel, doxorubicin, and cisplatin (*id.* (citing Ex. 1020, 9, Ex. 1002 ¶¶ 58, 123)); Pegram 1995 discloses that “the combination of trastuzumab/cisplatin was clinically effective in patients with metastatic HER2-positive breast cancer, with greater response rates and no apparent increase in toxicity relative to cisplatin

IPR2017-01122

Patent 7,892,549 B2

alone”; and “Seidman 1996 reports that paclitaxel is clinically effective against metastatic HER2-positive breast cancer.” *Id.* at 44–45.

Petitioner further argues that

as of December 1996, paclitaxel was one of the “most promising” chemotherapeutic drugs with efficacy against metastatic breast cancer. (Ex. 1007 (Abrams), 1164.) As such, a POSA would have been motivated to treat HER2-positive breast cancer patients with paclitaxel and to incorporate paclitaxel into the known, effective trastuzumab/cisplatin combination. (Ex. 1002, ¶ 119.) A POSA would have been particularly encouraged to combine paclitaxel with trastuzumab/cisplatin because Seidman 1996 reports that paclitaxel is clinically effective against metastatic HER2-positive breast cancer. (*Id.*, ¶ 119; Seidman 1996 (Ex. 1011).) The combination of trastuzumab and paclitaxel was already undergoing clinical trials for metastatic HER2+ breast cancer (Baselga 1996 (Ex. 1020), 743), and, indeed, paclitaxel and cisplatin were already being used in combination with one another to treat cancers, including metastatic breast cancer. (Ex. 1002, ¶ 119; Ex. 1012 (1995 TAXOL PDR), 683; *see also* Ex. 1013 (Tolcher), 37;<sup>[15]</sup> Ex. 1014 (Gelmon 1996), 1185.)<sup>[16]</sup>

Pet. 45.

In addition to clinical data, Petitioner also argues that “preclinical data reporting synergy between trastuzumab and paclitaxel in mouse xenografts would have provided even more motivation to a POSA to treat HER2-positive breast

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<sup>15</sup> Tolcher, *Paclitaxel Couplets with Cyclophosphamide or Cisplatin in Metastatic Breast Cancer*, 23(1) Supp. 1 SEMINARS ONCOLOGY 37–43 (1996) (discussing “potential advantages” of paclitaxel/cisplatin therapy and concluding that “[t]he paclitaxel/cisplatin combination has demonstrated an encouraging level of antitumor activity in women with metastatic breast cancer and has an acceptable level of toxicity”). Ex. 1013.

<sup>16</sup> Gelmon et al., *Phase I/II Trial of Biweekly Paclitaxel and Cisplatin in the Treatment of Metastatic Breast Cancer*, 14(4) J. CLINICAL ONCOLOGY 1185-91 (1996) (concluding that “[b]iweekly paclitaxel and cisplatin is an active combination for the treatment of metastatic breast cancer, including for patients with previous exposure to anthracyclines”). Ex. 1014.

IPR2017-01122

Patent 7,892,549 B2

cancer patients with this combination” as shown in Baselga 1996, Baselga Abstract 53 (cited in Baselga 1996), and Baselga Abstract 2262. Pet. at 46 (citing Exs. 1019, 1021); *see* sections II(D) (1),(5), and (6), *supra*.

Further with respect to motivation to combine, Petitioner contends that “[c]ombining trastuzumab, cisplatin, and paclitaxel for metastatic HER2-positive breast cancer particularly made sense because the combination satisfied the four principles of combination therapy.” *Id.* at 45–49 (citing Ex. 1002 ¶¶ 125–130); *see also id.* at 38–39 (stating the principles include “non-cross resistant drugs with single-agent activity, differing mechanisms of action, and nonoverlapping toxicity”) (quoting Ex. 1016, 204); Pet. Reply 15.

In sum, and relying on the clinical efficacy and toxicity profiles of trastuzumab, trastuzumab with paclitaxel, paclitaxel with cisplatin, as well as the preclinical data showing a synergistic effect of trastuzumab with paclitaxel, Petitioner contends that there would have been reasonable expectation of success that the three-drug combination would have been safe and effective. Pet. 52–53 (citing, Ex. 1002 ¶¶ 117–35); *see* Pet. Reply 1.

With respect to the limitation of claims 16 and 17, requiring administration of the claimed 3-part combination “in the absence of an anthracycline derivative,” Petitioner asserts that an ordinary artisan would have had multiple reasons to administer the claimed combination without an anthracycline derivative. Pet. 51–53. Petitioner first argues that an ordinary artisan “would have limited use of anthracycline derivatives in treatment whenever possible” due to the cardiotoxicity issues with anthracycline derivatives. *Id.* at 51. Moreover:

[B]ecause anthracycline derivatives were a first-choice therapy for metastatic breast cancer, many patient candidates for treatment with the trastuzumab and paclitaxel combination would have already been treated with anthracycline-based therapy. (Ex. 1002, ¶ 138; Ex. 1016



IPR2017-01122

Patent 7,892,549 B2

(Abeloff), 810.)<sup>[17]</sup> This means that many patients with metastatic disease who were prescribed a paclitaxel-containing regimen would have already endured extensive anthracycline-based therapy and would risk significant cardiotoxic effects with continued anthracycline-based therapy. (Ex. 1002, ¶ 138.)

*Id.* at 51–52. As a result, Petitioner contends that an ordinary artisan “would have avoided administering further anthracycline derivatives to the many patients who had already been treated with this class of drug or to the many patients who are resistant to treatment with anthracyclines.” *Id.*

With respect to the claim language “an amount effective to extend the time to disease progression in the human” (claims 1 and 16) and “effective amount” (claim 5), we credit Dr. Earhart’s testimony that “a person of ordinary skill in the art would have known that treatment with paclitaxel extends the time to disease progression relative to no treatment.” Ex. 1002 ¶ 157, n.28. We also find persuasive Petitioner’s argument that an ordinary artisan would have started with “the known amounts that were effective to extend the time to disease progression” in amounts previously shown to effectively treat metastatic breast cancer. Pet. at 49 (citing Ex. 1002 ¶ 132; Ex. 1020, 4–5 (effective doses of trastuzumab); Ex. 1012 (effective doses of paclitaxel)). “To the extent any modification to the amounts of the combination was necessary,” Petitioner continues, an ordinary artisan “would have readily optimized the combination treatment to arrive at an amount that results in the claimed efficacy and safety parameters.” *Id.* (citing Ex. 1002 ¶¶ 133–34; *see id.* at 50, n.16. Petitioner contends that “[s]uch optimization was routine in the art.” *Id.* at 49–50 (citing Ex. 1002 ¶ 134; Ex. 1016, 11, 13–14; Ex. 1001, 25:1–19, 43–54).

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<sup>17</sup> Excerpts from CLINICAL ONCOLOGY (Martin D. Abeloff et al., eds., Churchill Livingstone 1995). (“Abeloff”). Ex. 1016.

IPR2017-01122

Patent 7,892,549 B2

Patent Owner counters that Petitioner has not demonstrated that a person of ordinary skill in the art would have been motivated to combine rhuMAb HER2 with a taxoid; that the Board applied an incorrect claim construction, wherein under its preferred claim construction, Petitioner has not established a reasonable expectation of achieving the claimed clinical efficacy; and that the Sliwkowski Declaration, submitted during prosecution, confirms the patentability of the challenged claims. Patent Owner does not rely on evidence of secondary considerations. We address the relevant issues below.

a) Motivation to Combine rhuMAb HER2 with a Taxoid

On pages 37–41 of its Response, Patent Owner argues that the clinical and preclinical results discussed in Seidman 1996 and Baselga 1996 would not have motivated one of ordinary skill in the art to administer a combination of rhuMAb HER2 and a taxoid for the treatment of breast cancer.

(1) Seidman 1996

Relying on the testimony of Dr. Tannenbaum, Patent Owner contends that one of ordinary skill in the art would not have read the clinical data in Seidman 1996 as demonstrating that paclitaxel is clinically effective against metastatic HER2-positive breast cancer because “Seidman 1996 is an abstract, which a POSA would understand as reflecting a preliminary hypothesis, not proven efficacy; and a POSA would await an expanded analysis in a peer-reviewed journal before drawing any conclusions.” PO Resp. 39 (citing Ex. 2062 ¶¶ 184–185).

For the following reasons, we do not find this argument persuasive. First, as Petitioner points out, Patent Owner’s own experts rely on abstracts when favorable to its position. *See* Pet. Reply 5–6 (citing Ex. 1004, 321; Ex. 1056, ¶ 22); *see also* IPR2017-00737, Paper 102, Tr. 64:14–67:10 (Patent Owner admitting at oral argument that it relied on preclinical data from the Baselga Abstract 53 (“Baselga

IPR2017-01122

Patent 7,892,549 B2

'94") to justify to the FDA conducting phase III trials in the absence of phase II trials); *see also*, Ex. 2007, 63–64; Ex. 2001, 6–7, 39 (Patent Owner's reliance on abstracts in FDA submissions). Second, the inventors of the '549 patent do not appear to have considered abstracts unreliable as the patent cites numerous abstracts and posters on its face. *See* Ex. 1001, (56) References Cited. Indeed, in a declaration submitted during prosecution, Applicants expressly relied on an abstract to overcome prior-art rejections. *See* Ex. 1004-8, 1552; *see also* Ex. 1054 ¶ 16 ("Absent any allegation of misconduct on the part of the authors, a person of ordinary skill in the art would have had no reason to doubt their reported data.").

Under such circumstances, we are not persuaded that an ordinary artisan would have ignored or discounted the teachings of Seidman 1996 simply because it is an abstract.<sup>18</sup>

Patent Owner further appears to argue that a person of ordinary skill in the art would not have interpreted Seidman 1996 as showing the proven efficacy of taxoids in HER2-positive patients because "[t]he Seidman authors themselves continued to research the issue and ultimately found no 'statistically significant association with clinical response to taxane therapy' for patients who are HER2-positive." PO Resp. 39–40 (citing Ex. 2024, 2322). Patent Owner's argument,

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<sup>18</sup> With respect to the reliability of the Seidman 1996 authors, we note that they hale from the highly-respected Memorial Sloan-Kettering Cancer Center, and include two recipients of awards from the American Society of Clinical Oncology (Ex. 1011) and at least one—in Patent Owner's own words— a "leading practitioner" in the field (PO Resp. 62; *see* Reply 5). These authors also appear to have been collaborating with scientists of Patent Owner in rhuMAb HER2 research and clinical trials. *See, e.g.*, Ex. 1020, 3 (showing some of the same authors in Baselga 1996 as in Seidman 1996 and attributing the work on rhuMAb HER2 to both Memorial Sloan-Kettering Cancer Center and Genentech); *see also* Ex. 1019, 4 (Baselga Abstract 53 showing the same).

IPR2017-01122

Patent 7,892,549 B2

however, relies on Exhibit 2024, a 2002 article by van Poznak. As with Patent Owner’s unpublished internal documents evidencing the history of the invention and the development of its clinical trials, van Poznak was not available to one of ordinary skill in the art as of the date of the invention. *See* Ex. 1054 ¶ 14.

We are not persuaded that the van Poznak article, which reports on further research, fairly evidences what would have been understood by one of ordinary skill in the art at the time of the invention with respect to efficacy. *In re Kotzab*, 217 F.3d 1365, 1369 (Fed. Cir. 2000) (“A critical step in analyzing the patentability of claims pursuant to section 103(a) is casting the mind back to the time of invention, to consider the thinking of one of ordinary skill in the art, guided only by the prior art references and the then-accepted wisdom in the field.”); *see also Millennium Pharms., Inc. v. Sandoz Inc.*, 862 F.3d 1356, 1367 (Fed. Cir. 2017) (“obviousness is measured objectively in light of the prior art, as viewed by a person of ordinary skill in the field of the invention.”). Nor are we persuaded that the substance of van Poznak supports Patent Owner’s position. As Petitioner points out, van Poznak states, “[o]ur prior assessment of tumor HER2 expression through monoclonal antibody (45D5) and the polyclonal antibody (pAb-1) demonstrated that 4D5 positivity was predictive of positive response to taxane monotherapy.” Pet. Reply 6 (quoting Ex. 2024, 2320); *see* Ex. 1054 ¶ 15 (explaining that a closer reading of van Poznak shows that it “did not negate the finding that HER2+ patients are sensitive to paclitaxel”).<sup>19</sup>

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<sup>19</sup> We further note that, as the basis for its “prior assessment,” van Poznak references “Baselga J, Seidman AD, Rosen PP, et al: HER2 overexpression and paclitaxel sensitivity in Breast Cancer: Therapeutic Implications, *Oncology*, 2:43–48, 1997,” which appears to be the Baselga ’97 reference cited as prior art in IPR2017-00737 involving the same patent at issue here. *See* IPR2017-00737, Exhibit 1007. As noted in our Final Decision in that case, “Baselga ’97 teaches

IPR2017-01122

Patent 7,892,549 B2

We also find unpersuasive Patent Owner's citation to Yu et al.'s statement that "breast cancers that overexpress p185 [*i.e.*, HER2] will not respond well to Taxol" as evidence that one of ordinary skill would have been discouraged from using taxoids to treat HER2-positive breast cancer patients. PO Resp. 40 (citing Ex. 2029, 1362).<sup>20</sup> Taken in context, the cited statement in Yu et al., refers to the use of standalone paclitaxel, whereas the claimed invention relates to a taxoid *in combination* with rhuMAB HER2. Moreover, we find persuasive Petitioner's explanation that because the work of Yu et al. was done in tissue culture on cells engineered to overexpress HER2, one of ordinary skill would have regarded those findings as less predictive than the *in vivo* preclinical and clinical teachings of Baselga 1996 (Ex. 1011) and Seidman 1996 (Ex. 1010). *See*, Pet. Reply 7 (citing e.g., Ex. 1054 ¶ 17; Ex. 1002 ¶¶ 60, 124); *see also* Ex. 1040, 55:10–56:20 (Dr. Kerbel admitting that one study does not give rise to a widespread assumption that HER2-positive cells are less responsive to paclitaxel); Paper 64 at 4–5 (noting Exhibit 1043<sup>21</sup> a review paper regarding paclitaxel sensitivity in breast cancer fails to cite Yu, but "cites Seidman '96, Baselga '96 and the Baselga xenograft studies as suggesting that HER2+ tumors are sensitive to paclitaxel, and that combining trastuzumab with paclitaxel increased its antitumor activity").

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that rhuMoAb HER2, alone, 'is clinically active in patients who have metastatic breast cancers that overexpress HER2 and have received extensive prior therapy.'" *Id.* Paper 106, 19.

<sup>20</sup>Yu et al., *Overexpression of c-erbB-2/neu in breast cancer cells confers increased resistance to Taxol via mdr-1-independent mechanisms*, 13 ONCOGENE 1359–654 (1996).

<sup>21</sup> Baselga et al., *HER2 Overexpression and Paclitaxel Sensitivity in Breast Cancer: Therapeutic Implications*, Update on the Taxanes in Breast Cancer, *Oncology*, Vol. 11, No. 3 (Suppl. 2), 43–48 (1997) (cited as Baselga '97 in IPR2017-00737).

IPR2017-01122

Patent 7,892,549 B2

We therefore conclude that one of ordinary skill in the art would have understood that taxoids were 1) used in combination therapy for the treatment of metastatic breast cancer (Ex. 1012, 6; Ex. 1014; Ex. 1013), 2) were suggested to be particularly useful for HER 2 positive breast cancer (Ex. 1011), and 3) demonstrated synergy in combination with anti-HER-2 monoclonal antibodies in animal models of HER2 breast cancer (*see* Ex. 1020, 9 (“In preclinical studies . . . rhuMAB HER2 markedly potentiated the antitumor effects of several chemotherapeutic agents, including . . . paclitaxel, without increasing their toxicity.”); Ex. 1010, 5; Ex. 1021). We find no merit in Patent Owner’s argument that safety concerns would have “dissuaded POSAs from using combination therapy involving taxoids.” *See* PO Resp. 41 (citing Ex. 2062 ¶¶ 59–61, 194–198); *see also, e.g.*, Ex. 1010 (referencing paclitaxel as “the most important new cytotoxic agent to be introduced for the management of breast cancer in many years”); Ex. 1010, 5 (stating that “clinical trials combining paclitaxel with anti-growth factor receptor MoAbs [e.g., rhuMAB HER2] are being planned”); Ex. 1020, 9; Ex. 2111, 4 (“Paclitaxel was selected [to combine with rhuMAB HER2] because of its activity in metastatic breast cancer and preclinical studies that supported its use.”).<sup>22</sup>

(2) Baselga 1996

With respect to Baselga 1996, Patent Owner argues that the reference merely discloses the administration of rhuMAB HER2 alone and “discusses preclinical combinations with ‘several chemotherapeutic agents, including cisplatin, doxorubicin, and paclitaxel.’” PO Resp. 37–38. And although Patent Owner

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<sup>22</sup> S. Shak, *Overview of the Trastuzumab (Herceptin) Anti-HER2 Monoclonal Antibody Clinical Program in HER2-Overexpressing Metastatic Breast Cancer*, *Sem. Oncol.* 26(4), Supp. 12 (1999).

IPR2017-01122

Patent 7,892,549 B2

admits that Baselga 1996 discloses that “clinical trials of such combination therapy are currently in progress,” it argues that “it could not have been referring to rhuMAb HER2 plus paclitaxel because there was no clinical study involving that combination at the time Baselga-1996 was submitted.” *Id.* at 38–39; *see also id.* at 20–23 (relying on non-prior art documents to establish the history of the invention and development of related clinical trials).

We do not find Patent Owner’s argument persuasive for the reasons set forth on pages 16–18 of Petitioner’s Reply.<sup>23</sup> Baselga 1996 states that “[i]n preclinical studies . . . rhuMAb HER2 markedly potentiated the antitumor effects of several chemotherapeutic agents, including cisplatin, doxorubicin, and paclitaxel, without increasing their toxicity” and, as a result, “clinical trials of such combination therapy are currently in progress.” Ex. 1020, 9. Based on our reading of Baselga 1996 as a whole, we agree with Petitioner that one of ordinary skill in the art would have understood from this passage that clinical trials of rhuMAb HER2 in combination with each of cisplatin, doxorubicin, and paclitaxel were currently in progress for the treatment of breast cancer. *See* Pet. Reply 16 (citations omitted); *see also* Ex. 1010, 5 (stating that “clinical trials combining paclitaxel with anti-growth factor receptor MoAbs [e.g., rhuMAB HER2] are being planned”).

That a clinical study involving rhuMAb HER2 in combination with paclitaxel may not have yet commenced when Baselga 1996 was published does not, as Petitioner points out, diminish its teachings because the record fails show

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<sup>23</sup> We note that the relevant time for our obviousness analysis is not the submission date of the prior art, as Patent Owner appears to suggest, but the date of the alleged invention, which in this case, is later than the publication date of Baselga 1996. It is undisputed that at the time Baselga 1996 was published, a clinical study involving the claimed combination was indeed in progress.

IPR2017-01122

Patent 7,892,549 B2

that one of ordinary skill in the art would have been aware of this fact. In this respect, Patent Owner's citation to Shak<sup>24</sup> is unavailing as Shak merely indicates that a paclitaxel arm was added sometime after the trial began in June 1995. *See* Ex. 2111, 73. Patent Owner's reliance on non-public documents to establish when it added a paclitaxel arm is similarly insufficient because there is no evidence one of ordinary skill in the art would "have been privy to [Patent Owner's] internal, non-public development history." Pet. Reply 16.

Further, and though we do not find relevant Patent Owner's non-public documents evidencing the history of the invention and the development of clinical trials involving rhuMAb HER2 in combination with a taxoid, we agree with Petitioner that these documents do not evidence any uniform opposition or skepticism but "show[] that the suggestion to add the paclitaxel/trastuzumab arm was quickly accepted both internally and at FDA." Pet. Reply 17–18 & n.11; *see e.g.*, Ex. 1035 (reporting that FDA "thought our plan [regarding HER2 protocol changes] was reasonable" and that "[t]heir preliminary review of our plan seemed to be reasonable since we are having difficulties recruiting patients.") Ex. 2004, 4 (noting that "[i]nitial FDA feedback on the Taxol modification is positive.") 10, (quoting internal reviewers as stating: "I support the Taxol amendment"; "The parallel strategy is important and I support it"; suggested changes "are appropriate"; and "a good gamble"), (comments of non-supporting reviewer directed to statistical power rather than use of taxol, *per se*).

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<sup>24</sup> Shak et al., *Overview of the Trastuzumab (Herceptin) Anti-HER2 Monoclonal Antibody Clinical Program in HER2-Overexpressing Metastatic Breast Cancer*, 26(4), Suppl. 12 SEMINARS ONCOLOGY 71-77 (1999). Ex. 2111.



IPR2017-01122  
 Patent 7,892,549 B2

### (3) Reliability of Baselga Xenograft Data

Patent Owner also argues that the preclinical results referenced in Baselga 1996 (as further discussed in Baselga Abstract 53 (Ex. 1019) and Baselga Abstract 2262 (Ex. 1021)) fail to provide motivation to combine rhuMAb HER2 and a taxoid. PO Resp. 41–42. We do not find Patent Owner’s argument persuasive for the reasons set forth on pages 7–12 of Petitioner’s Reply. We find particularly compelling Petitioner’s evidence that Patent Owner itself relied on the Baselga xenograft results to obtain FDA approval to test the rhuMAb HER2/paclitaxel combination in Phase III clinical trials. *See, e.g.*, Ex. 2007, 27, 64; Ex. 2001, 6–7, 39; Ex. 1052, 144:17–150:16; *see also* IPR2017-00737, Paper 102, Tr. 64:14–67:18 (Patent Owner’s admission at oral argument that Baselga xenograft data was used, at least “[i]n part,” to justify to the FDA conducting phase III trials in the absence of phase II trials). In this regard, the Federal Circuit has recognized that “FDA approval may be relevant to the obviousness inquiry.” *Allergan, Inc. v. Sandoz Inc.*, 726 F.3d 1286, 1291 (Fed. Cir. 2013) (citing *Knoll Pharm. Co., Inc. v. Teva Pharms. USA, Inc.*, 367 F.3d 1381, 1385 (Fed. Cir. 2004)).

Despite relying on the Baselga xenograft data in its FDA submissions, Patent Owner now argues that the design of the preclinical study renders that data unreliable. *See* PO Resp. 41–43. We do not, however, find persuasive Patent Owner’s implication that one of ordinary skill in the art would have discounted Baselga’s results because the authors used a single cell line (BT-474) with a high level of HER2 expression. *See* PO Resp. 42 (citing Ex. 2061 ¶ 62; Ex. 2062 ¶ 168).<sup>25</sup> We credit, instead, the testimony of Dr. Earhart that one of ordinary

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<sup>25</sup> In Paper 26, Patent Owner further contends that model cell lines having 11 (MDA-435), 31 (SK-BR3), and 52 (BT-474) copies of ErbB2 per cell reflects “the heterogeneity of human chromosomes.” Paper 26, 14–15 (citations omitted). To

IPR2017-01122

Patent 7,892,549 B2

skill in that art would consider this high level of HER2 gene expression “advantageous, rather than detrimental” because high levels of HER2 expression was known to be correlated with poor treatment outcomes. Ex. 1054 ¶ 10. Accordingly, “[a] person of ordinary skill in the art would consider positive results using the BT-474 cell line as a motivation to pursue the tested agent.” *Id.*

With respect to the site of tumor implantation, we also credit Dr. Earhart’s opinion that the subcutaneous implantation technique used by Baselga was reliable, routinely used, and still common today. *Id.*; *see also* Ex. 1105 ¶ 9 (explaining why Baselga’s reporting of only a single time point was not evidence of unreliability). Accordingly, we find reasonable Dr. Earhart’s opinion that “no person of ordinary skill in the art would question the validity of [Baselga’s] subcutaneous xenograft studies in comparing proposed combination treatment regimens.” *Id.* at ¶ 9.

Patent Owner also appears to argue that one of ordinary skill in the art would not have risked treating a patient with a combination of rhuMAb HER2 and a taxoid because the Baselga data lacked, “e.g., testing [of] multiple cell lines, creation of orthotopic xenograft models, and analysis of dosing amounts.” PO Resp. 44. We do not find this argument persuasive in light of Patent Owner’s reliance on the Baselga data in its FDA submissions, the known use of rhuMAb HER2 (Ex. 1020, Ex. 1022) and paclitaxel (Ex. 1010; Ex. 1011; Ex. 1012) in treating breast cancer, and Dr. Earhart’s explanation that, “when each element of a combination therapy had previously been shown to be safe and effective on its own

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the extent Patent Owner intends to convey that the variation in ErbB2 copy number in the referenced cell lines reflects the heterogeneity of HER2 expression within or between HER2-positive tumors in human patients, this would appear to support Dr. Earhart’s position that it was reasonable to rely on cell line BT-474 in preclinical trials, as it would be expected to have the highest, yet still physiologically relevant, expression level among the referenced cell lines.

IPR2017-01122

Patent 7,892,549 B2

in clinical studies [as is the case here], it would not be necessary to run preclinical studies on the combination” (Ex. 1054 ¶ 8).

Patent Owner also raises Hsu in response to Petitioner’s reliance on the Baselga xenograft data. Patent Owner introduced Exhibit 2135 (“Hsu”)<sup>26</sup> at Dr. Earhart’s April 17, 2018 deposition (*see* Paper 83, 1), and submitted arguments with respect to Hsu in connection with its motions on observation (Paper 68, ¶ 8; Paper 76 ¶ 8), to which Petitioner replied (Paper 74, ¶¶ 3–4; Paper 80 ¶¶ 3–4). Hsu is also subject to Petitioner’s motion to exclude, discussed below, in section III(C)(2).

Hsu is an abstract appearing in the Proceedings of a March 7–12, 1997 conference on Basic & Clinical Aspects of Breast Cancer. Ex. 2135. According to Hsu, in vitro cytotoxicity assays on HER2-expressing human breast cancer cells showed that rhuMAb HER2 in combination with taxol had additive cytotoxic effects, whereas in a mouse model involving these “HER-2/*neu*-transfected MCF-7 human breast cancer” cells, “[x]enografts treated with rhuMAb HER-2 plus taxol . . . were not significantly different from drug alone controls with the doses and dose schedules tested in this model.” *Id.*

As we understand Patent Owner’s position, one of ordinary skill in the art would have discounted Baselga’s xenograft results in light of Hsu’s (allegedly) contradictory teachings demonstrating a lack of synergy between rhuMAb HER2 and a taxoid. *See* Paper 68, ¶ 8; Paper 74, ¶¶ 3–4. We are not persuaded by the

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<sup>26</sup> Hsu, et al., *Therapeutic Advantage of Chemotherapy Drugs in Combination with Recombinant, Humanized, Anti-HER-2/*neu* Monoclonal Antibody (rhuMAb HER-2) Against Human Breast Cancer Cells and Xenografts with HER-2/*neu* Overexpression*, Proc. Basic & Clin. Aspects of Breast Cancer, A-39 (March 7-12, 1997). Ex. 2135.

IPR2017-01122

Patent 7,892,549 B2

merits of Patent Owner's argument. Relying on the testimony of Dr. Earhart, Petitioner reasonably argues that Hsu fails to describe the doses and schedules tested such that one of ordinary skill in the art would not have known whether they were comparable to Baselga's. Paper 64, 8–9 (citing Ex. 1105 ¶ 13). Petitioner further distinguishes Hsu as using HER2-transfected cells, rather than naturally-HER2 overexpressing human tumor cells such as the BT-474 cell line used in Baselga. *Id.* Based on the evidence of record in this case, we agree with Petitioner that one of ordinary skill in the art would not conclude that Hsu's teachings were inconsistent with those of Baselga. *See also* IPR2017-00737, Paper 86 (Final Written Decision), section II(E)(1).

b) “In the Absence of an Anthracycline Derivative”

With respect to the limitation of independent claim 16, requiring the administration of an anti-ErbB2 antibody, a taxoid, and a further growth inhibitory agent “in the absence of an anthracycline derivative,” Patent Owner argues that one of ordinary skill in the art would have been motivated to combine rhuMAb HER2 with an anthracycline rather than with a taxoid in light of safety and efficacy concerns associated with taxoids. PO Resp. 45–46 (citations omitted). For the reasons set forth at pages 12–15 of Petitioner's Reply Brief, we do not find Patent Owner's arguments persuasive.

As an initial matter, we credit Patent Owner's argument that one of ordinary skill in the art seeking to combine rhuMAb HER2 with an existing anti-cancer drug would have reasonably looked to anthracyclines because they were a common first-line chemotherapy agent with known, but manageable, side effects. PO Resp. 45–46. This, however, is insufficient to establish the non-obviousness of the rhuMAb HER2/taxoid combination. *See Bayer Pharma AG v. Watson Labs., Inc.*, 874 F.3d 1316, 1329 (Fed. Cir. 2017) (quoting *In re Fulton*, 391 F.3d 1195, 1200

IPR2017-01122

Patent 7,892,549 B2

(Fed. Cir. 2004)) (“While a skilled artisan may have preferred a delayed-release formulation over the claimed immediate-release formulation, ‘that the prior art as a whole suggests the desirability of a particular combination need not be supported by a finding that the prior art suggests that the combination claimed . . . is the preferred, or most desirable, combination.’”).

The evidence of record shows that while anthracyclines were widely employed, one of ordinary skill in the art would also have been motivated to combine rhuMAb HER2 with a taxoid such as paclitaxel rather than with an anthracycline. Paclitaxel was approved for the treatment of metastatic breast cancer, recommended as a “highly active . . . initial chemotherapy for metastatic breast cancer,” and shown to be clinically effective against HER2-positive breast cancers. Ex. 1012, 6; Ex. 1011; Ex. 1019; Ex. 1021; Ex. 1039, 1943; *see also* Ex. 1014 (disclosing that paclitaxel is active as a single agent in metastatic breast cancer, and exhibits advantageous, if not synergistic, effects in combination therapy); Ex. 1054 ¶13 (noting that paclitaxel side effects were controllable and generally not dose limiting). Moreover, in light of preclinical studies demonstrating that paclitaxel was synergistic with anti-HER2 antibodies, Baselga 1996 states that “clinical trials [including rhuMAb HER2/taxoid] combination therapy are currently in progress.” *See* Ex. 1020, 9. Consistent with this considerable interest in taxoids for the treatment of breast cancer, a contemporary review of a wide variety of chemotherapeutic agents for breast cancer including anthracyclines, touts taxanes (i.e., taxoids, including paclitaxel and docetaxel), as “foremost among these new agents” and “one of the most exciting new classes of chemotherapeutic agents to be developed.” Ex. 1007, 6.

The evidence of record also shows that one of ordinary skill in the art would have been motivated to administer the claimed combination “in the absence of an

IPR2017-01122

Patent 7,892,549 B2

anthracycline derivative,” where prior treatment with anthracyclines was discontinued due to drug resistance or cumulative cardiotoxicity. *See* Pet. 51–52; Ex. 1002 ¶ 106, 138–139, 161; Ex. 1016, 26–30. The FDA-approved labeling for Taxol, for example, states that it “is indicated, after failure of first-line or subsequent chemotherapy” where “[p]rior therapy should have included an anthracycline.” Ex. 2112, 6. The prior art of record confirms that many patients with metastatic breast cancer will have previously been treated with, and become resistant to, first-line anthracycline chemotherapeutics. Gelmon 1996, for example, discloses that “[a]ll but two of the women in our trial had been treated with previous adjuvant chemotherapy, and 23 of 29 patients had previous exposure to anthracyclines.” Ex. 1014, 5. Thus, on the present record, we find persuasive Dr. Earhart’s testimony that

A person of ordinary skill in the art would have expected that many patients had previous anthracycline treatment, given that anthracyclines were a first-line therapy for breast cancer. (Ex. 1016 at 1693.) Therefore, particularly for patients who had already been treated with an anthracycline, it would have been obvious not to include the drug in the combination of trastuzumab and paclitaxel.

Ex. 1002 ¶ 138.

c) The Sliwkowski Declaration<sup>27</sup>

During the prosecution leading to the issuance of the ’549 Patent, the Examiner withdrew an obviousness rejection involving Baselga 1996 “in view of the declaration of Mark X. Sliwkowski, PhD.” Ex. 1019-7, 47–48. Although none of its experts address the Sliwkowski Declaration, Patent Owner states “if the Board considers Dr. Sliwkowski’s declaration, it only confirms the patentability of

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<sup>27</sup> Declaration of Mark X. Sliwkowski, Ph.D., executed October 15, 2009. Ex. 1009.

IPR2017-01122

Patent 7,892,549 B2

the challenged claims.” PO Resp. 52; *see also id* at 27, 52–54 (discussing aspects of the Sliwowski Declaration). Thus, although Patent Owner does not appear to rely on the Sliwowski Declaration, in the interest of completeness, we accept Patent Owner’s invitation to consider it.

The Sliwowski Declaration asserted, *inter alia*, that “a skilled scientist would have anticipated that paclitaxel would provide little or no additional benefit to treatment with trastuzumab alone since trastuzumab would arrest the cell cycle before paclitaxel would be able to act,” and that one of ordinary skill in the art would recognize that “anti-HER2 antibodies acting by inducing cell cycle arrest in the G1 phase, would antagonize the effect of taxoids, such as paclitaxel, since they arrest cell cycle before it reaches the G2/M phase, where taxoids exert their apoptotic antitumor activity.” Ex. 1009, 341–345 ¶¶ 3, 4. Patent Owner’s experts nowhere address this concept and we accept Dr. Earhart’s well-reasoned conclusion that “Dr. Sliwowski’s theory and reasoning . . . are based on several false assumptions about how these agents work to treat cancer, and are contradicted by the data available in the prior art, which predicted a favorable interaction between trastuzumab and paclitaxel.” Ex. 1002 ¶¶ 140–150.

According to Patent Owner, the Sliwowski Declaration “also explained that preclinical results would not have provided a reasonable expectation of success as to the clinical results for the combination of rhuMAb HER2 and a taxoid; indeed, xenograft models at that time were poor predictors of clinical results for breast cancer.” PO Resp. 27. With respect to these issues, the Sliwowski Declaration adds nothing more to Patent Owner’s position, and we agree with Petitioner that the Sliwowski Declaration does not negate the motivation to combine or reasonable expectation of success demonstrated in the prior art. *See* Pet. 53–62.

IPR2017-01122

Patent 7,892,549 B2

d) Reasonable Expectation of Success

Patent Owner also contends that Petitioner has not established a reasonable expectation of success in achieving either the claimed clinical efficacy or the claimed clinical safety. PO Resp. 49–54. We do not find Patent Owner’s argument persuasive.

As set forth in section II(C)(2), above, the proper interpretation of “extend the time to disease progression” requires a comparison of the claimed combination treatment to no treatment. Petitioner asserts that combining trastuzumab with paclitaxel satisfies the limitation of clinical efficacy because each of trastuzumab and paclitaxel extends time to disease progression relative to no treatment, and an ordinary artisan “would not have expected the combination to change this.” Pet. 50 n.16 (citing Ex. 1002 ¶¶ 137, 157 n.28; Ex. 1010); *see* Ex. 1020, 6–7 (describing time to tumor progression for the patients with either minor or stable disease as having “unusually long,” with a median duration of 5.1 months). We find Petitioner’s argument persuasive. Indeed, Patent Owner does not argue, and we do not find, that combining a taxoid with rhuMAb HER2 would abrogate the effect of either therapeutics. *See* Dec. 23–24. Thus, an ordinary artisan would have had a reasonable expectation of success in achieving the claimed clinical efficacy.

e) Patentability under Patent Owner’s Claim Construction

We also address patentability under Patent Owner’s proposed construction of “an amount effective to extend the time to disease progression in the human patient” and “an effective amount” as comparing the three-part treatment to treatment with taxoid alone. As an initial matter, Patent Owner argues that “no reference disclosed that the claimed combination extended TTP in human patients compared to patients treated with paclitaxel alone.” *See* Paper 53, 11–12. Patent



IPR2017-01122

Patent 7,892,549 B2

Owner also admits, however, that when rhuMoAb is “administered with a chemotherapy in the ‘taxoid’ family, this claimed combination therapy significantly extends the time to disease progression (‘TTP’) as compared with patients receiving taxoid therapy alone.” PO Resp. 2. The claimed extension of time to disease progression is, thus, an inherent benefit of an otherwise obvious combination, and such an inherent result cannot establish patentability. “[A]n obvious formulation cannot become nonobvious simply by administering it to a patient and claiming the result[.]” *Santarus, Inc. v. Par Pharm., Inc.*, 694 F.3d 1344, 1354 (Fed. Cir. 2012). “To hold otherwise would allow any formulation—no matter how obvious—to become patentable merely by testing and claiming an inherent property.” *Id.*

With respect to the parties’ arguments, Patent Owner contends that under its preferred construction, Petitioner has not established that one of ordinary skill in the art would have had a reasonable expectation of success in achieving the claimed efficacy—i.e., administration of the claimed composition “in an amount effective to extend the time to disease progression” as compared to a patient treated with a taxoid alone. PO Resp. 47–52. In particular, Patent Owner argues that neither Seidman 1996 nor Pegram 1995 address TTP, and although the 1995 Taxol PDR and Baselga 1996, respectively, provide TTP data for patients treated with Taxol and rhuMoAb monotherapy, neither provides a basis to determine whether the claimed combination extends TTP compared to treatment with taxoid alone. *Id.* at 48–49. According to Patent Owner, these failings cannot be overcome by reference to patient response rates in Baselga 1996. *Id.* at 49–50.<sup>28</sup>

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<sup>28</sup> Although we agree with Patent Owner that Petitioner has not shown that one of ordinary skill in the art would read the preclinical studies described in the Baselga

IPR2017-01122  
Patent 7,892,549 B2

Upon careful consideration of all the evidence, we find that Petitioner has the better argument. *See* Pet. Reply 19–22. In particular, we credit the testimony of Dr. Earhart that because effective amounts of rhuMoAb and paclitaxel were known, one of ordinary skill in the art would have had a reasonable expectation that a combination of these agents would extend the time to disease progression relative to treatment with paclitaxel alone. *See, e.g.*, Ex. 1002 ¶¶ 119–120, 132, 132, 157; Ex. 1054 ¶¶ 20–23.

Although Patent Owner points out that the cited references do not expressly state that monotherapy with rhuMoAb or paclitaxel extends the time to disease progression, we credit Dr. Earhart’s testimony that response rates and TTP are clinical surrogate endpoints used to estimate the likelihood of overall survival, and that a person of ordinary skill in the art would understand that a positive response rate would likely correlate with an increased TTP. Ex. 1054 ¶ 22; *see* Pet. Reply 20–22; Ex. 1002 ¶¶ 92–94, 136–137, 157, 166; Paper 64, 6. Consistent with this testimony, the ’549 Specification also suggests time to disease progression and response rates as alternative measurements of efficacy. *See* Ex. 1057-1, 19 (15:12–17) (’649 priority application defining therapeutically effective amount; noting that “efficacy can . . . be measured by assessing the time for disease progression (TTP), or determining the response rates (RR)” 46–47 (42–43) (noting that clinical benefit is “assessed by response rates and the evaluation of disease progression”).

Accordingly, we are persuaded that “a person of ordinary skill in the art would have understood that the response rate results reported in Baselga were

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references as supporting an increase in TTP (*see* PO Resp. 50), this does not affect our ultimate determination as to the obviousness of the challenged claims.

IPR2017-01122

Patent 7,892,549 B2

likely to correlate with an extension of time to disease progression and an increase in overall survival.” Ex. 1054 ¶ 23; *see* Ex. 1020, 6, 7 & Table 4 (reporting “37% of patients achieved minimal responses or stable disease,” and “an overall response rate of 11.6%”). As Dr. Earhart explains, “[a] person of ordinary skill in the art would have been motivated to combine trastuzumab with paclitaxel, with a reasonable expectation of success that the combination would perform better than no treatment and better than paclitaxel alone . . . [and] achieve an extension of TTP over paclitaxel alone based on the superior TTP of trastuzumab.” Ex. 1054 ¶¶ 19–20.

Patent Owner also emphasizes the high failure rate of clinical trials, in general, as evidence for the unpredictability of treating cancer. PO Resp. 11–12. Patent Owner relies on Exhibit 2021,<sup>29</sup> a review article on the pharmaceutical industry by Kola and Landis. PO Resp. 11–12. According to Patent Owner’s expert, Kola and Landis “showed that approximately only five percent of oncology drugs were successful,” and “that in oncology, the rate of failure in Phase III trials ‘is as high as 59%,’” Ex. 2062 ¶¶ 91–92, 218.<sup>30</sup> Kola and Landis, however, focuses on clinical trials of individual compounds (i.e., new chemical entities (NCEs) and biologics) rather than combinations of known or promising therapies. *See e.g.*, Ex. 2021, 711 (discussing the “[d]epressing approval rates of NCEs and biologics”); *id.* at 712 (Table entitled, “NCEs required to achieve specific real growth targets as a function of 2002 revenues”); (addressing “the root causes of

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<sup>29</sup> Kola and John Landis, *Can the Pharmaceutical Industry Reduce Attrition Rates?* 3 NATURE REV. 711-715 (2004) (“Ex. 2021”).

<sup>30</sup> We further note that Dr. Tannenbaum appears to base “success” on FDA approval, which is a higher standard than required for patentability. *See* Ex. 2062 ¶ 214.

IPR2017-01122

Patent 7,892,549 B2

why **compounds** undergo attrition in the clinic,” and stating that “more than 70% of oncology **compounds** fail [in Phase II trials]” and “approximately 45% of all **compounds** that enter [Phase III trials] undergo attrition and in some therapeutic areas, such as oncology, it is as high as 59%”) (emphasis added).

Kola and Landis does not discuss the likelihood of failure of combination therapies like those at issue here—wherein paclitaxel was already FDA approved for treatment of breast cancer, rhuMoAb HER2 showed promise in Phase II trials, and both paclitaxel and rhuMoAb HER2 had been used successfully in combination therapy with a third compound, cisplatin. Moreover, despite Dr. Tannenbaum’s assertion that the increased cardiotoxicity of anthracyclines in combination with rhuMAB HER2 shows the lack of predictability of new combinations of existing therapies, such information was not in the prior art at the time of the invention. Ex. 2062 ¶ 207. Accordingly, we do not give substantial weight to Dr. Tannenbaum’s opinions on this topic.

Also relying on Dr. Tannenbaum’s testimony, Patent Owner argues that the four principles of combination therapy discussed by Dr. Earhart (*see* Ex. 1002 ¶¶ 125–130; Ex. 1024 ¶¶ 130–131)<sup>31</sup> only apply to small molecule chemotherapeutics and are inapplicable to combinations involving antibodies such as rhuMoAb HER2. *See* PO Resp 11–12, 46–47, 51 (citations omitted). We do not find Patent Owner’s arguments persuasive.

At its core, Patent Owner’s assertion is based on the fact that the “four principles” concept was established before the use of therapeutic antibodies such that there is no record evidence of researchers expressly applying these principles

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<sup>31</sup> Although not necessary to our Decision, we find that Dr. Earhart’s discussion of these principles underscores and further supports our patentability analysis.

IPR2017-01122

Patent 7,892,549 B2

to combinations involving antibodies. *See id.* But this merely reflects the historical use of small molecule chemotherapeutic combinations before the development of more complex therapeutic antibodies. *See* Ex. 2072, 365 (noting the introduction of chemotherapeutic combination therapy for advanced breast cancer in 1963).

Patent Owner further bases its assertion on evidence that combining chemotherapy with chemoendocrine (hormone) therapy “did not increase the response rate, TTP, or survival as compared to either treatment alone.” PO Resp. 51–52. Patent Owner does not, however, suggest that such therapy involved therapeutic antibodies, nor persuade us that the failure of the chemotherapy/hormone therapy combination would dissuade one of ordinary skill in the art from combining chemotherapeutic treatments with other therapies. Moreover, Patent Owner’s expert, Dr. Tannenbaum admitted that she was not aware of any prior art suggesting that the four principles would not apply to chemotherapy/antibody combinations such as rhuMoAb HER2/paclitaxel. Ex. 1052, 71:25–72:6, 90:9–91:6; *see also id.* at 99:11–18, 102:17–106:20, 108:24–109:12 (admitting that the prior art suggested the use of antibodies with chemotherapies, including the rhuMoAb/paclitaxel combination).

Patent Owner also references Exhibit 2136<sup>32</sup> (Wadler) as indicating that incorporating various biological agents in combination regimens with chemotherapeutic “offers an important challenge to the medical oncologist.” Paper 53, 7–8. While we do not completely discount the teachings of this reference, we note Petitioner’s argument that Wadler is primarily focused on cytokines and

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<sup>32</sup> Wadler & Schwartz, *Antineoplastic Activity of the Combination of Interferon and Cytotoxic Agents Against Experimental and Human Malignancies: A Review*, *Cancer Res.* 50:3473-3486 (1990) (Exhibit 2136).

IPR2017-01122

Patent 7,892,549 B2

growth factors, rather than antibodies, and does not take into account the body of knowledge in the art regarding the use of rhuMoAb HER2. *See* Paper 64; Ex. 1105 ¶ 15 (noting that Wadler “recommends further study of a combination of interferon alpha [sic] with 5-fluorouracil”). On balance, the record does not suggest that one of ordinary skill in the art would reject the four principles of combination therapy when considering rhuMoAb HER2 therapy. The record as a whole supports a finding that an ordinary artisan would have had a reason to combine trastuzumab and paclitaxel for the treatment of metastatic HER2-positive breast cancer.

a) Conclusion

Considering the evidence as a whole, we agree with Petitioner that one of ordinary skill in the art would have been motivated to combine the teachings of Baselga 1996, Seidman 1996, Pegram 1995, and 1995 TAXOL PDR with a reasonable expectation of success in achieving the invention of claims 1–11 and 14–17 of the ’549 Patent. Accordingly, and applying either the construction set forth in section II(C)(2), above, or Patent Owner’s preferred construction, we conclude that Petitioner has demonstrated by a preponderance of evidence that the challenged claims would have been obvious.

III. Motions

A. Patent Owner’s Motion to Amend

Having concluded that claims 1–11 and 14–17 are unpatentable, we address Patent Owner’s contingent Motion to Amend.

1. *Threshold Requirements*

In an *inter partes* review, amended claims are not added to a patent as of right, but rather must be proposed as a part of a motion to amend. 35 U.S.C. § 316(d). The Board must assess the patentability of the proposed substitute claims

IPR2017-01122

Patent 7,892,549 B2

“without placing the burden of persuasion on the patent owner.” *Aqua Prods., Inc. v. Matal*, 872 F.3d 1290, 1328 (Fed. Cir. 2017). Patent Owner’s proposed substitute claims, however, must still meet the statutory requirements of 35 U.S.C. § 316(d) and the procedural requirements of 37 C.F.R. § 42.121. *See* “Guidance on Motions to Amend in view of *Aqua Products*” (Nov. 21, 2017), available at [https://www.uspto.gov/sites/default/files/documents/guidance\\_on\\_motions\\_to\\_amend\\_11\\_2017.pdf](https://www.uspto.gov/sites/default/files/documents/guidance_on_motions_to_amend_11_2017.pdf). Accordingly, Patent Owner must demonstrate (1) the amendment proposes a reasonable number of substitute claims; (2) the amendment does not seek to enlarge the scope of the claims of the patent or introduce new subject matter; (3) the amendment responds to a ground of unpatentability involved in the trial; and (4) the original disclosure sets forth written description support for each proposed claim. *See* 35 U.S.C. § 316(d); 37 C.F.R. § 42.121.

In its Motion to Amend, “Petitioner conditionally seeks to amend the claims to make explicit that the claimed comparison is against a patient treated with paclitaxel alone.” PO Resp. 48, n.14; *see* Paper 26, 4. Accordingly, Patent Owner proposes to replace all existing claims (claims 1–17) with substitute claims 18–20, of which claims 18 and 19 are independent. Paper 26, 2 and Appendix A. Under the circumstances, we agree with Patent Owner that it proposes a reasonable number of substitute claims. *See Id.* at Abstract.

With respect to the substance of the proposed claims, Claim 18, submitted as a replacement for claim 1, recites:

18. A method of treatment of a human patient with breast cancer that overexpresses ErbB2 receptor, comprising administering a combination of rhuMAb HER2, paclitaxel, and a further growth inhibitory agent to a human patient in an amount effective to extend the time to disease progression in the human patient, as compared to paclitaxel alone, wherein the antibody binds to epitope 4D5 within the ErbB2 extracellular domain sequence.

IPR2017-01122

Patent 7,892,549 B2

*Id.* Claim 19, submitted as a replacement for claim 16 is similar, but further recites the administration of rhuMAb HER2, paclitaxel, and a further growth inhibitory agent “in the absence of an anthracycline derivative.” *Id.* Depending from claim 19, claim 20 specifies that the ErbB2 overexpressing breast cancer is metastatic breast carcinoma and is identical to original claim 17 but for its dependency.

Patent Owner contends that the substitute claims do not enlarge but, instead, narrow the scope of the original claims. *Id.* at 2–5. According to Patent Owner, the proposed substitute claims narrow the scope of the claimed antibody by replacing the genus of “an antibody that binds ErbB2” of claim 1 or “an intact antibody which binds to epitope 4D5 with the ErbB2 extracellular domain sequence” of claim 16, with the “specific antibody species, ‘rhuMAb HER2,’ a recombinant humanized 4D5 anti-ErbB2 antibody also known as HERCEPTIN®.” Paper 26, 2–3. Patent Owner similarly argues that the substitute claims narrow the genus encompassing “a taxoid” by reciting “‘paclitaxel,’ which is a specific species of taxoid.” *Id.* at 3.

With respect to the claim language, “an amount effective to extend the time to disease progression in the human patient,” Patent Owner contends that “the Challenged Claims do not expressly identify a comparator for the claimed ‘time to disease progression’; therefore, by further limiting the claims with a specific comparator (patients treated with paclitaxel alone), the Substitute Claims do not enlarge the scope of the claims.” *Id.* at 4. Alternatively, Patent Owner argues that the additional limitation merely makes explicit that, under Patent Owner’s preferred construction of the original claims, “the proper comparator by which to measure the claimed efficacy is to a patient treated with paclitaxel alone.” *Id.* With respect to the original claims, we apply our construction for the term “extend the time to disease progression” as indicating that the results of the claimed



IPR2017-01122

Patent 7,892,549 B2

combination therapy is compared to patients receiving no treatment. Because we do not discern, and Petitioner does not contend, that the comparator of patients receiving no treatment is broader than those receiving paclitaxel alone in the proposed amended claims, we agree with Patent Owner that the amendment does not seek to enlarge the scope of the claims as required under 35 U.S.C. § 316(d) and 37 C.F.R. § 42.121.

Petitioner argues that we should deny Patent Owner's Motion to Amend under 37 C.F.R. § 42.121(a)(2)(i) because the amendments narrowing the claims to specifically recite "rhuMAb HER2" and "paclitaxel" do not respond to the instituted grounds of unpatentability. Paper 43, 2–6; Paper 64, 1–2. According to Patent Owner, "[i]t is not required that *every* amended limitation be solely for the purpose of overcoming an instituted ground" such it is sufficient that the proposed claims have been amended to specify that the comparator for an amount effective to extend the time to disease progression is paclitaxel alone. *See* Paper 26, 9 & fn.3. (citing *Veeam Software Corp. v. Veritas Techs., LLC*, IPR2014-00090, Paper 48 at 28-29 (PTAB July 17, 2017)). We agree with Patent Owner. "[37 C.F.R. § 42.121(a)(2)(i)] does not require, however, that every word added to or removed from a claim in a motion to amend be solely for the purpose of overcoming an instituted ground. Additional modifications that address potential 35 U.S.C. § 101 or § 112 issues, *for example*, are not precluded by rule or statute." *Western Digital Corp. v. SPEX Techs., Inc.*, Case IPR2018-00082 (PTAB Apr. 25, 2018) (Paper 13) (informative), slip op. at 6 (emphasis added). Although Patent Owner does not indicate whether the disputed limitations are intended to address 35 U.S.C. §§ 101 or 103 issues, this is not expressly required under our rules. Moreover, in indicating that addressing potential § 101 or § 112 issues are merely exemplary, *Western Digital* suggests that Patent Owner may have other reasons for entering

IPR2017-01122

Patent 7,892,549 B2

such amendments. As the disputed limitations are peripheral to our patentability analysis (*see* section III(A)(2), below) and do not otherwise unduly burden the just and speedy resolution of this matter, we do not reject Patent Owner’s Motion to Amend under 37 C.F.R. § 42.121(a)(2)(i).

Petitioner also argues that the substitute claims add new subject matter in contravention of Section 316(d) and Rule 42.121(a)(2)(ii). *See* Paper 43, 6–7; Paper 80, 3. Although Patent Owner asserts that each of the proposed substitute claims find support in the original disclosure (Paper 26, 5–9; Paper 53, 3–4), Petitioner argues that “a person of ordinary skill in the art would not have recognized from the specification [of the asserted priority documents] that the inventor had possession of a triple combination treatment that extends time to disease progression compared to paclitaxel alone,” (Paper 43, 6–7), i.e., that the priority documents that Patent Owner relies on lack sufficient written descriptive support for the full scope of the proposed claims.

“In determining whether claims introduce new matter, we look to whether the original application provides adequate written description support for the claims.” *Kapsch TrafficCom IVHS Inc. v. Neology, Inc.*, Case IPR2016-01763, slip op. at 47 (PTAB Mar. 20, 2018) (Paper 60). The written description requirement is met when the specification “conveys to those skilled in the art that the inventor had possession of” and “actually invented” the claimed subject matter. *Ariad Pharm., Inc. v. Eli Lilly & Co.*, 598 F.3d 1336, 1351 (Fed. Cir. 2010) (*en banc*). “And while the description requirement does not demand any particular form of disclosure, or that the specification recite the claimed invention *in haec verba*, a description that merely renders the invention obvious does not satisfy the requirement.” *Id.* at 1352 (citations omitted); *See also In re Wertheim*, 541 F.2d 257, 262 (CCPA 1976) (“It is not necessary that the application describe the claim

IPR2017-01122

Patent 7,892,549 B2

limitations exactly, . . . but only so clearly that persons of ordinary skill in the art will recognize from the disclosure that appellants invented processes including those limitations.”).

Patent Owner’s proposed substitute claims require the administration of a three-drug combination —rhuMAb HER2, paclitaxel, and a further growth inhibitory agent— “in an amount effective to extend the time to disease progression in the human patient, as compared to paclitaxel alone.” Patent Owner’s support for the clinical effects of this three-drug combination, however, relates to the administration of a two-drug combination. *See* Paper 26, 5–8; Paper 53, 3. In particular, Patent Owner relies on “a clinical study in which patients with metastatic [HER2-positive] breast cancer or overexpression of the ErbB2 oncogene were treated with a combination of a humanized version of the murine 4D5 antibody (HERCEPTIN®) (also known as rhuMAb HER2) and Taxol® (also known as paclitaxel) in the absence of anthracycline derivative.” Paper 26, 7. Patent Owner asserts that “[t]he results state that ‘assessments of time to disease progression (TTP) in months) and response rates (RR) showed a significant augmentation of the chemotherapeutic effect by HERCEPTIN®, without increase in overall severe adverse events (AE).’” *Id.* (citing Ex. 1004-1 49 (43:19–21) and Ex. 1009, 43–44 (42:29–43:2)).

The written description requirement demands that inventors “do more than merely disclose that which would render the claimed invention obvious.” *ICU Medical, Inc. v. Alaris Medical Systems, Inc.* 558 F.3d 1368, 1377 (Fed. Cir. 2009). Considering the evidence of record, we agree with Petitioner that “a person of ordinary skill in the art would not have recognized from the specifications that the inventor had possession of a triple combination treatment that extends time to disease progression compared to paclitaxel alone.” Paper 43, 7. “Showing

IPR2017-01122

Patent 7,892,549 B2

possession of a different, unclaimed combination is insufficient.” *See Ariad*, 598 F.3d at 1352. Because Patent Owner has not shown, and we do not find adequate written description supporting the proposed substitute claims, they likewise fail to satisfy the no new matter requirement of 35 U.S.C. § 316(d) and 37 C.F.R. § 42.121(a)(2)(ii). Accordingly, we deny Patent Owner’s Motion to Amend.

2. *Unpatentability of the Amended Claims*

In addition to its failure to meet the “no new matter” requirement for a motion to amend, we deny Patent Owner’s Motion to Amend because Petitioner has shown by a preponderance of the evidence that claims 18–20 are obvious in view of the art of record. *See* Paper 43, 7–20. Paper 64, 3–10. In short, Patent Owner does not contend, nor do we discern, that narrowing the proposed claims to specifically recite “rhuMAb HER2” and “paclitaxel” bears on patentability, but relies on the addition of the words “as compared to paclitaxel alone” to make explicit the claim construction it argued with respect to the originally-challenged claims. PO Resp. 48, n.14; *see* Paper 26, 4. Patent Owner then recites substantially the same arguments it put forth with respect to claims 1–11 and 14–17 under its preferred construction. *Cf.* Paper, 26, 9–24; Paper 53, 4–12 with PO Resp. 37–54. Having found those arguments unavailing (*see* section II(E), above), we decline to revisit them here.

B. Patent Owner’s Motion to Exclude Evidence

Patent Owner filed one motion to exclude evidence. Paper 59. Petitioners opposed (Paper 72) and Patent Owner submitted a reply in support of its motion (Paper 75).

1. *Evidence Relating to Secondary Considerations*

Patent Owner requests that we exclude Exhibits 1033, 1034, 1038, 1059, and 1060 as irrelevant. Paper 59, 1–2. According to Patent Owner, these exhibits

IPR2017-01122

Patent 7,892,549 B2

relate to secondary considerations, which it does not assert in this proceeding. *Id.*; Paper 75, 1. Petitioner concurs, noting that it has not cited these documents in this *inter partes* review. Paper 72, 1. Accordingly, we dismiss Patent Owner’s request as moot.

## 2. *Evidence Concerning Surrogate Endpoints*

Patent Owner moves to exclude Exhibit 1055, as well as select paragraphs of Dr. Earhart’s reply declaration (Ex. 1054 ¶¶ 22–23), which relate to Petitioner’s argument that one of ordinary skill in the art would understand that response rates and time to disease progression are surrogates for time to disease progression, and that one of ordinary skill in the art would expect some measure of correlation between these values. *See* Paper 59, 2–4; Paper 72, 1–2. Patent Owner argues that we should exclude this evidence as untimely because Petitioners raised it for the first time in their reply, “after which PO had no opportunity to respond.” Paper 59, 2.

We do not find Patent Owner’s arguments persuasive for the reasons set forth in Petitioners’ opposition (Paper 72, 1–3), which we adopt. In particular, we agree with Petitioner that Exhibit 1055 and paragraphs 22–23 of Exhibit 1054 are proper rebuttal to Patent Owner’s contention that “the ‘response rates disclosed in the instituted references . . . do not suggest an extension of TTP when using the claimed combination.’” *Id.* at 3 (citing PO Resp. 49). *See Ericsson Inc. v. Intellectual Ventures I LLC*, No. 2017-1521, 2018 WL 4055815, at \*6 (Fed. Cir. Aug. 27, 2018) (Board improperly refused to consider Reply testimony that “merely expands on a previously argued rationale”); *Belden Inc. v. Berk-Tek LLC*, 805 F.3d 1064, 1077–78 (Fed. Cir. 2015) (holding that a petitioner may not submit new evidence or argument in reply that it could have presented earlier, e.g. to make

IPR2017-01122

Patent 7,892,549 B2

out a prima facie case of unpatentability, but may submit directly responsive rebuttal evidence in support of its reply).

Accordingly, we deny Patent Owner's motion to exclude Exhibit 1055 and paragraphs 22–23 of Exhibit 1054.

### 3. *Gelmon Declaration*

Patent Owner requests that we exclude Exhibit 1056, which is a declaration submitted by Dr. Karen Gelmon on behalf of Patent Owner in IPR2017-01139. Paper 59, 4–5; Paper 75, 2. As set forth in its opposition, Petitioner relies on Exhibit 1056 to rebut Patent Owner's argument that one of ordinary skill in the art would not rely on Seidman 1996 because it was “merely an abstract.” *See* Paper 72, 3–4. Insofar as Dr. Gelmon relies on an abstract in arguments on behalf of Patent Owner, we find Petitioner's citation to Exhibit 1056 relevant. Although, as Patent Owner points out, Dr. Gelmon relies on additional information, this goes to the weight we accord Petitioner's evidence, not its admissibility. *See* Paper 59, 4–5. Patent Owner has not explained, nor do we discern, how this might “mislead or confuse” the Board. *See id.* at 5. Accordingly we deny Patent Owner's motion to exclude Exhibit 1056.

### 4. *Gottlieb Article*

Patent Owner requests that we “exclude Exhibit 1036, a 1980 article published in *Chest for Pulmonologists, Cardiologists, Cardiothoracic Surgeons and Related Specialists*, entitled, *Late, Late Doxorubicin Cardiotoxicity* (“Gottlieb”), and paragraph 38 of Dr. Earhart's reply declaration relying on Gottlieb (Exhibit 1054)” as untimely because Petitioner raised it for the first time in its reply. Paper 59, 5–6. According to Patent Owner, “to the extent Petitioner wished to present evidence that POSAs would have been motivated to avoid anthracyclines, it was obligated to do so in the Petition as part of its prima facie

IPR2017-01122

Patent 7,892,549 B2

case, rather than wait until its Reply, after which PO had no opportunity to respond.” Paper 75, 3. We do not find Patent Owner’s argument persuasive.

The Petition itself sets forth a reasoned explanation of why one of ordinary skill in the art would have been motivated to avoid anthracyclines, stating, for example, that one of ordinary skill in the art:

would have been well-aware of the cardiotoxicity issues with anthracycline derivatives. (Ex. 1002, ¶ 138; Ex. 1016 (Abeloff), 813.) Anthracyclines were known to cause irreversible cardiotoxicity thereby limiting the total lifetime dose a patient can receive. (Ex. 1002, ¶ 138; Ex. 1016 (Abeloff), 813.) Accordingly, a POSA would have limited use of anthracycline derivatives in treatment whenever possible. (Ex. 1002, ¶ 139; Ex. 1016 (Abeloff), 813.) Further, because anthracycline derivatives were a first-choice therapy for metastatic breast cancer, many candidates for treatment with the trastuzumab and paclitaxel combination would have already been treated with anthracycline-based therapy. (Ex. 1002, ¶ 138; Ex. 1016 (Abeloff), 810.) This means that many patients with metastatic disease who were prescribed a paclitaxel-containing regimen would have already endured extensive anthracycline-based therapy and would risk significant cardiotoxic effects with continued anthracycline-based therapy. (Ex. 1002, ¶ 138.) POSAs would have avoided administering further anthracycline derivatives to the many patients who had already been treated with this class of drug or to the many patients who are resistant to treatment with anthracyclines, rendering the limitation “in the absence of an anthracycline derivative” obvious. (Ex. 1002, ¶ 138; *see also* Ex. 1020 (Baselga 1996), at 740 (reporting that a patient died during treatment with trastuzumab due to congestive heart failure associated with prior anthracycline use); Ex. 1024 (Arbuck), at 128-29 (reporting that many anthracycline-resistant patients responded to paclitaxel).)

Pet. 51–52. In addition, we agree with Petitioner that its introduction of Gottlieb was a reasonable rebuttal to Patent Owner’s argument that one of ordinary skill in the art “would have not have been motivated to avoid anthracycline due to the cardiotoxicity caused by anthracyclines because the cardiotoxicity caused by anthracyclines was ‘manageable.’” Paper 72, 5; *see* Pet. Reply 13 (citing Gottlieb

IPR2017-01122  
Patent 7,892,549 B2

(among others) as teaching that “[t]he cardiotoxicity of anthracyclines was the major factor limiting their use”). Accordingly, we deny Patent Owner’s motion to exclude Exhibit 1036 and Exhibit 1054 ¶38.

#### 5. *Dr. Kerbel’s Patent Application*

Patent Owner requests that we exclude Exhibit 1100, an international patent application naming Dr. Kerbel as an inventor as irrelevant under FRE 402 and as “tend[ing] to mislead and confuse the issues” in contravention of FRE 403. Paper 59, 7–8; Paper 75, 3. Patent Owner has not explained, nor do we discern, how the Board might be misled or confused by Exhibit 1100. Moreover, Petitioners have adequately explained the relevance of these exhibits to the present case. *See* Paper 72, 5–7. Accordingly, we deny Patent Owner’s motion to exclude Exhibit 1100.

#### C. Petitioners’ First and Second Motions to Exclude Evidence

In its first motion (Paper 61), Petitioner moves to exclude Exhibits 2052, 2055, 2070, 2075, 2106, 2133, 2135 and 2139, and portions of expert declarations submitted on behalf of Patent Owner that rely on them (Ex. 2061 ¶ 56; Ex. 2143 ¶¶ 11, 15; Ex. 2144 ¶¶ 27–28). Patent Owner opposed (Paper 70) and Petitioner submitted a reply in support of its first motion (Paper 77). In its second motion, Petitioner moves to exclude Exhibit 2146. Paper 81. Patent Owner opposed (Paper 83) and Petitioner submitted a reply in support of its first motion (Paper 84).

##### 1. Exhibits 2052, 2055, 2070, 2075, 2106, 2133, and 2139

Petitioner contends that Exhibits 2075, 2133, and 2139 are dated after December 12, 1997, the priority date of the ’441 patent, and that Patent Owner has not established that Exhibits 2052, 2055, 2070, and 2106, were published before this date, such that each of these exhibits are “irrelevant for the purpose of establishing the teachings of the prior art, and Patent Owner is relying on them for improper purposes.” Paper 61, 1, 3. We do not, however, expressly rely on



IPR2017-01122

Patent 7,892,549 B2

Exhibits 2052, 2055, 2070, 2075, 2106, 2133, or 2139 in our Decision. Moreover, having considered the merits of Patent Owner's arguments in light of these teachings, our decision as to the patentability of the challenged claims would not change if they were excluded from evidence. Accordingly, we need not decide the merits of Petitioner's motion with respect to these documents and dismiss Petitioner's request as moot.

2. Exhibits 2135 and 2146 (Hsu)

In its first and second motions, Petitioner also requests that we exclude the Hsu Abstract (Exhibit 2135), a related document encompassing Hsu (Exhibit 2146), and certain expert testimony relying on those exhibits. Among other things, Petitioner contends Patent Owner has not established the authenticity or prior art status of Exhibits 2135 and 2146, and that they are hearsay under FRE 802. *See* Paper 61, 7–9; Paper 81, 4–7. As set forth in section II(E)(a)(3), above, we do not find persuasive Patent Owner's evidence regarding the substance of Hsu. Accordingly, and taking no position as to the merits of the parties' arguments relating to the admissibility of the Hsu references, we deny this remaining portion of Petitioner's request as moot.

D. Motions to Seal

We also address the five unopposed motions to seal pursuant to the Modified Default Standing Protective Order set forth in Exhibit 2036 (*see* Paper 24, 3): Papers 27 and 52 (by Patent Owner) and Papers 44, 47, and 62 (by Petitioner).

The Board's standards for granting motions to seal are discussed in *Garmin International v. Cuozzo Speed Technologies, LLC*, IPR2012-00001 (PTAB Mar. 14, 2013) (Paper 34). In summary, there is a strong public policy for making all information filed in *inter partes* review proceedings open to the public, especially because the proceeding determines the patentability of claims in an issued patent

IPR2017-01122

Patent 7,892,549 B2

and, therefore, affects the rights of the public. *Id.* at slip op. 1–2. Under 35 U.S.C. § 316(a)(1) and 37 C.F.R. § 42.14, the default rule is that all papers filed in an inter partes review are open and available for access by the public; a party, however, may file a concurrent motion to seal and the information at issue is sealed pending the outcome of the motion. It is only “confidential information” that is protected from disclosure. 35 U.S.C. § 316(a)(7); *see* Office Patent Trial Practice Guide, 77 Fed. Reg. 48,756, 48,760 (Aug. 14, 2012). The standard for granting a motion to seal is “for good cause.” 37 C.F.R. § 42.54(a). The party moving to seal bears the burden of proof in showing entitlement to the requested relief, and must explain why the information sought to be sealed constitutes confidential information. 37 C.F.R. § 42.20(c).

We remind the parties of the expectation that confidential information relied upon or identified in a final written decision will be made public. *See* Office Trial Practice Guide, 77 Fed. Reg. 48756, 48761 (Aug. 14, 2012). Confidential information that is subject to a protective order ordinarily becomes public 45 days after final judgment in a trial. A party seeking to maintain the confidentiality of the information may file a motion to expunge the information from the record prior to the information becoming public. 37 C.F.R. § 42.56.

#### 1. Patent Owner’s Motions to Seal

In Paper 27, Patent Owner seeks to seal the unredacted version of Exhibit 2050 (the Deposition Transcript of Robert Howard Earhart, Jr., M.D., Ph.D.), and the unredacted version of Exhibit 2069 (the Declaration of Stephanie Mendelsohn, which purports to authenticate previously sealed Exhibits 2001–2005, 2007, and 2008). Patent Owner has shown good cause supporting the motion. Insofar as, none of the material in Exhibits 2050 or 2069 is relied on in our final Decision, Patent Owner’s request is granted. Because we rely herein on Exhibits 2001, 2004,

IPR2017-01122

Patent 7,892,549 B2

and 2008, we rescind our grant of Patent Owner's motion to seal with respect to those documents. *See* Paper 24, 2. Within 14 days of this Decision, Patent Owner may submit redacted versions of Exhibits 2001, 2004, and/or 2008 that fairly disclose the material relied on in this Decision along with a renewed motion to seal, filed jointly.

In Paper 52, Patent Owner seeks to seal Exhibit 2142 (Genentech, Inc. Document GENENTECH\_0000034-GENENTECH-0000139) and the unredacted version of Exhibit 2144 (Supplemental Expert Declaration of Dr. Susan Tannenbaum). Patent Owner has shown good cause supporting the motion. Insofar as, we do not rely on material in Exhibits 2142 or Exhibit 2144 in our final Decision, Patent Owner's request is granted with respect to those documents.

Also in Paper 52, Patent Owner seeks to seal the unredacted version of Paper 53 (Patent Owner's Reply in Support of Contingent Motion to Amend Under 37 C.F.R. § 42.121). Patent Owner's request is denied without prejudice, subject to the conditions set forth in the Order, below.

## 2. Petitioner's Motions to Seal

Petitioner seeks to seal the confidential versions of its Reply (Paper 45), and its Opposition and Sur-Reply to Patent Owner's Motion to Amend (Papers 42, 64, and respectively), because they "refer to materials that Patent Owner Genentech has designated as Confidential pursuant to the Modified Default Standing Protective Order." Paper 44, 1; Paper 62, 1. Petitioner seeks to seal Exhibits 1035, 1046, 1049, and 1058 for the same reason. Paper 47, 1.

Petitioners provide no other justification for why the redacted portions of the cited documents should be kept confidential and, thus, fail to satisfy the good cause requirement. Accordingly, Petitioners' motions are denied. Petitioner's request is further denied with respect to Exhibit 1035, which we rely on in our Decision.

IPR2017-01122

Patent 7,892,549 B2

Patent Owner is invited to file, within 14 days of this Decision, a motion to seal any presently redacted portion(s) of Papers 42, 45, 53, and 64 or Exhibits 1046, 1049, and 1058. The motion must explain why the information sought to be protected is truly confidential and attest that such information is not directly or indirectly relied on in our Final Written Decision. Petitioner may respond within one week of Patent Owner's motion, if desired. These Papers and Exhibits will remain designated Board and Parties Only for 21 days from this Decision or until consideration of any such motion and reply.

#### IV. CONCLUSION

After considering Petitioners' and Patent Owner's arguments and evidence, we conclude that Petitioners have shown, by a preponderance of the evidence, that claims 1–11 and 14–17 of the '549 patent would have been obvious over the combination Baselga 1996, Seidman 1996, Pegram, and the 1995 TAXOL PDR as set forth in the Petition.

Based on the evidence of record, we conclude that proposed amended claims 18–20 introduce new matter in contravention of Section 316(d) and Rule 42.121(a)(2)(ii) and, moreover, would not be patentable over the art of record. The parties' motions to exclude evidence and to seal are addressed in the following Order.

#### V. ORDER

In consideration of the foregoing, it is:

ORDERED that claims 1–11 and 14–17 of the '549 patent are unpatentable;

FURTHER ORDERED that Patent Owners' motion to amend is denied;

FURTHER ORDERED that Patent Owner's motion to exclude Exhibits 1033, 1034, 1038, 1059, and 1060 is denied as moot;

IPR2017-01122

Patent 7,892,549 B2

FURTHER ORDERED that Patent Owner's motion to exclude Exhibits 1100, 1036, 1055, 1056, and paragraphs 22–23, and 38 of Exhibit 1054 is denied.

FURTHER ORDERED that Petitioners' motion to exclude Exhibits 2052, 2055, 2070, 2075, 2106, 2133, 2135, 2139, and 2146 is denied as moot.

FURTHER ORDERED that Patent Owner's motions to seal Exhibits 2069 and 2142, and the confidential versions of Exhibits 2050 and 2144 is granted.

FURTHER ORDERED that, notwithstanding our prior Order in Paper 24, we rescind our Order to seal Exhibits 2001, 2004, and 2008. Within 14 days of this Decision, Patent Owner may submit redacted versions of Exhibits 2001, 2004, and/or 2008 that fairly disclose the material relied on in this Decision along with a renewed motion to seal, filed jointly.

FURTHER ORDERED that Petitioner's motion to seal Exhibit 1035 is denied.

FURTHER ORDERED that Petitioner's motion to seal Exhibits 1046, 1049, and 1058, and the confidential versions of Papers 42, 45, and 64 is denied without prejudice to Patent Owner.

FURTHER ORDERED that Patent Owner may file, within 14 days of this Decision, a motion to seal any of Exhibits 1046, 1049, and 1058 or the presently redacted portion(s) of Papers 42, 45, 52, and 64. The motion must explain why the information sought to be protected is truly confidential and attest that such information is not directly or indirectly relied on in our final Decision. Petitioner may respond within one week of Patent Owner's motion, if desired. These Papers and Exhibits will remain designated Board and Parties Only for 21 days from this Decision or until consideration of any such motion and reply.

IPR2017-01122

Patent 7,892,549 B2

FURTHER ORDERED that, because this is a final written decision, parties to the proceeding seeking judicial review of the decision must comply with the notice and service requirements of 37 C.F.R. § 90.2.

IPR2017-01122  
Patent 7,892,549 B2

PETITIONER:

Cynthia Hardman  
Elizabeth J. Holland  
Robert Cerwinski  
GOODWIN PROCTER LLP  
chardman@goodwinlaw.com  
eholland@goodwinlaw.com  
rcerwinski@goodwinlaw.com

PATENT OWNER:

David L. Cavanaugh  
Lauren V. Blakely  
Robert J. Gunther, Jr.  
Lisa J. Pirozzolo  
Kevin S. Prussia  
Andrew J. Danford  
WILMER CUTLER PICKERING HALE AND DORR LLP  
david.cavanaugh@wilmerhale.com  
lauren.blakely@wilmerhale.com  
robert.gunther@wilmerhale.com  
lisa.pirozzolo@wilmerhale.com  
kevin.prussia@wilmerhale.com  
andrew.danford@wilmerhale.com

Adam R. Brausa  
Daralyn J. Durie  
DURIE TANGRI LLP  
abrausa@durietangri.com  
ddurie@durietangri.com



US007892549B2

(12) **United States Patent**  
**Paton et al.**

(10) **Patent No.:** **US 7,892,549 B2**  
(45) **Date of Patent:** **\*Feb. 22, 2011**

(54) **TREATMENT WITH ANTI-ERBB2 ANTIBODIES**

(75) Inventors: **Virginia E. Paton**, Oakland, CA (US);  
**Steven Shak**, Burlingame, CA (US);  
**Susan D. Hellmann**, San Carlos, CA (US)

(73) Assignee: **Genentech, Inc.**, South San Francisco, CA (US)

(\*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 1827 days.

This patent is subject to a terminal disclaimer.

(21) Appl. No.: **10/356,824**

(22) Filed: **Feb. 3, 2003**

(65) **Prior Publication Data**

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**Related U.S. Application Data**

(63) Continuation of application No. 09/208,649, filed on Dec. 10, 1998.

(60) Provisional application No. 60/069,346, filed on Dec. 12, 1997.

(51) **Int. Cl.**

**A61K 39/395** (2006.01)

**C07K 16/28** (2006.01)

**C07K 16/30** (2006.01)

(52) **U.S. Cl.** ..... **424/143.1**; 424/130.1; 424/133.1; 424/134.1; 424/135.1; 424/136.1; 424/138.1; 424/141.1; 424/152.1; 424/155.1; 424/156.1; 424/172.1; 424/174.1

(58) **Field of Classification Search** ..... 424/130.1, 424/133.1, 138.1, 141.1, 143.1, 155.1, 174.1, 424/134.1, 135.1, 136.1, 152.1, 156.1, 172.1

See application file for complete search history.

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*Primary Examiner*—Alana M. Harris

*Assistant Examiner*—Anne L. Holleran

(74) *Attorney, Agent, or Firm*—Arnold & Porter LLP; Diane Marschang; Ginger R. Dreger

(57) **ABSTRACT**

The present invention concerns the treatment of disorders characterized by the overexpression of ErbB2. More specifically, the invention concerns the treatment of human patients susceptible to or diagnosed with cancer overexpressing ErbB2 with a combination of an anti-ErbB2 antibody and a chemotherapeutic agent other than an anthracycline, e.g. doxorubicin or epirubicin.

17 Claims, 1 Drawing Sheet



## US 7,892,549 B2

Page 2

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US 7,892,549 B2

Page 6

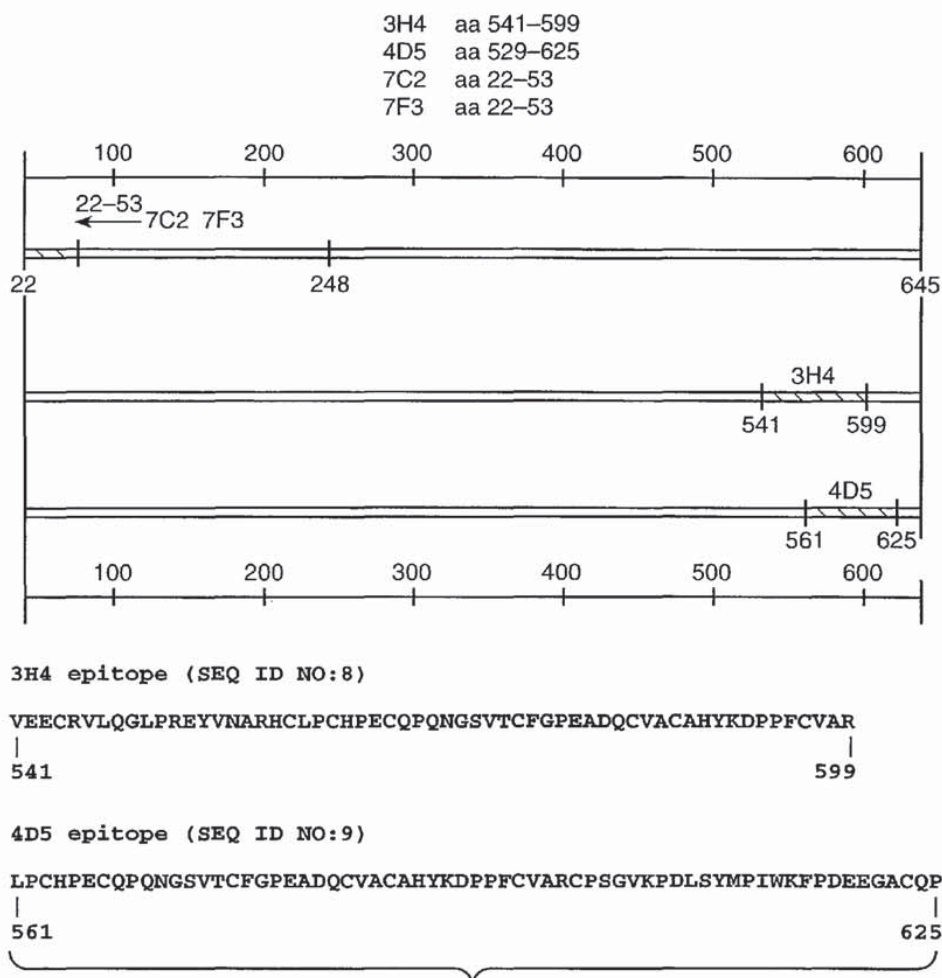
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U.S. Patent

Feb. 22, 2011

US 7,892,549 B2

**FIG. 1**

1 MELAALCRWGLLLALLPPGAASTQVCTGTDMKRLRLPA  
38 SPETHLDMLRHLYQGCQVVQGNLELTYLPTNASLSFL  
75 QDIQEVQGYVLIAHNQVRQVPLQRLRIVRGTLFEDN  
112 YALAVLDNGDPLNNTTPVTGASPGGLRELQLRSLTEI  
149 LKGGVLIQRNPQLCYQDTILWKDIFHKNNQLALTLLID  
186 TNRSRA

**FIG. 2**



US 7,892,549 B2

1

## TREATMENT WITH ANTI-ERBB2 ANTIBODIES

This is a continuation of non-provisional application Ser. No. 09/208,649, filed Dec. 10, 1998, which claims priority under 35 USC §119 to provisional application No. 60/069,346, filed Dec. 12, 1997, the entire disclosures of which are hereby incorporated by reference.

### FIELD OF THE INVENTION

The present invention concerns the treatment of disorders characterized by the overexpression of ErbB2. More specifically, the invention concerns the treatment of human patients susceptible to or diagnosed with cancer overexpressing ErbB2 with a combination of an anti-ErbB2 antibody and a chemotherapeutic agent other than an anthracycline, e.g. doxorubicin or epirubicin.

### BACKGROUND OF THE INVENTION

Proto-oncogenes that encode growth factors and growth factor receptors have been identified to play important roles in the pathogenesis of various human malignancies, including breast cancer. It has been found that the human ErbB2 gene (erbB2, also known as her2, or c-erbB-2), which encodes a 185-kd transmembrane glycoprotein receptor (p185<sup>HER2</sup>) related to the epidermal growth factor receptor (EGFR), is overexpressed in about 25% to 30% of human breast cancer (Slamon et al., *Science* 235:177-182 [1987]; Slamon et al., *Science* 244:707-712[1989]).

Several lines of evidence support a direct role for ErbB2 in the pathogenesis and clinical aggressiveness of ErbB2-overexpressing tumors. The introduction of ErbB2 into non-neoplastic cells has been shown to cause their malignant transformation (Hudziak et al., *Proc. Natl. Acad. Sci. USA* 84:7159-7163 [1987]; DiFiore et al., *Science* 237:178-182 [1987]). Transgenic mice that express HER2 were found to develop mammary tumors (Guy et al., *Proc. Natl. Acad. Sci. USA* 89:10578-10582 [1992]).

Antibodies directed against human erbB2 protein products and proteins encoded by the rat equivalent of the erbB2 gene (neu) have been described. Drebin et al., *Cell* 41:695-706 (1985) refer to an IgG2a monoclonal antibody which is directed against the rat neu gene product. This antibody called 7.16.4 causes down-modulation of cell surface p185 expression on B 104-1-1 cells (NIH-3T3 cells transfected with the neu proto-oncogene) and inhibits colony formation of these cells. In Drebin et al. *PNAS* (USA) 83:9129-9133 (1986), the 7.16.4 antibody was shown to inhibit the tumorigenic growth of neu-transformed NIH-3T3 cells as well as rat neuroblastoma cells (from which the neu oncogene was initially isolated) implanted into nude mice. Drebin et al. in *Oncogene* 2:387-394 (1988) discuss the production of a panel of antibodies against the rat neu gene product. All of the antibodies were found to exert a cytostatic effect on the growth of neu-transformed cells suspended in soft agar. Antibodies of the IgM, IgG2a and IgG2b isotypes were able to mediate significant in vitro lysis of neu-transformed cells in the presence of complement, whereas none of the antibodies were able to mediate high levels of antibody-dependent cellular cytotoxicity (ADCC) of the neu-transformed cells. Drebin et al. *Oncogene* 2:273-277 (1988) report that mixtures of antibodies reactive with two distinct regions on the p185 molecule result in synergistic anti-tumor effects on neu-transformed NIH-3T3 cells implanted into nude mice. Biological effects

2

of anti-neu antibodies are reviewed in Myers et al., *Meth. Enzym.* 198:277-290 (1991). See also WO94/22478 published Oct. 13, 1994.

Hudziak et al., *Mol. Cell. Biol.* 9(3):1165-1172 (1989) describe the generation of a panel of anti-ErbB2 antibodies which were characterized using the human breast tumor cell line SKBR3. Relative cell proliferation of the SKBR3 cells following exposure to the antibodies was determined by crystal violet staining of the monolayers after 72 hours. Using this assay, maximum inhibition was obtained with the antibody called 4D5 which inhibited cellular proliferation by 56%. Other antibodies in the panel, including 7C2 and 7F3, reduced cellular proliferation to a lesser extent in this assay. Hudziak et al. conclude that the effect of the 4D5 antibody on SKBR3 cells was cytostatic rather than cytotoxic, since SKBR3 cells resumed growth at a nearly normal rate following removal of the antibody from the medium. The antibody 4D5 was further found to sensitize p185<sup>erbB2</sup>-overexpressing breast tumor cell lines to the cytotoxic effects of TNF- $\alpha$ . See also WO89/06692 published Jul. 27, 1989. The anti-ErbB2 antibodies discussed in Hudziak et al. are further characterized in Fendly et al. *Cancer Research* 50:1550-1558 (1990); Kotts et al. *In Vitro* 26(3):59A (1990); Sarup et al. *Growth Regulation* 1:72-82 (1991); Shepard et al. *J. Clin. Immunol.* 11(3):117-127 (1991); Kumar et al. *Mol. Cell. Biol.* 11(2):979-986 (1991); Lewis et al. *Cancer Immunol. Immunother.* 37:255-263 (1993); Pietras et al. *Oncogene* 9:1829-1838 (1994); Vitetta et al. *Cancer Research* 54:5301-5309 (1994); Sliwkowski et al. *J. Biol. Chem.* 269(20):14661-14665 (1994); Scott et al. *J. Biol. Chem.* 266:14300-5 (1991); and D'souza et al. *Proc. Natl. Acad. Sci.* 91:7202-7206 (1994).

Tagliabue et al. *Int. J. Cancer* 47:933-937 (1991) describe two antibodies which were selected for their reactivity on the lung adenocarcinoma cell line (Calu-3) which overexpresses ErbB2. One of the antibodies, called MGR3, was found to internalize, induce phosphorylation of ErbB2, and inhibit tumor cell growth in vitro.

McKenzie et al. *Oncogene* 4:543-548 (1989) generated a panel of anti-ErbB2 antibodies with varying epitope specificities, including the antibody designated TA1. This TA1 antibody was found to induce accelerated endocytosis of ErbB2 (see Maier et al. *Cancer Res.* 51:5361-5369[1991]). Bacus et al. *Molecular Carcinogenesis* 3:350-362 (1990) reported that the TA1 antibody induced maturation of the breast cancer cell lines AU-565 (which overexpresses the erbB2 gene) and MCF-7 (which does not). Inhibition of growth and acquisition of a mature phenotype in these cells was found to be associated with reduced levels of ErbB2 receptor at the cell surface and transient increased levels in the cytoplasm.

Stancovski et al. *PNAS* (USA) 88:8691-8695 (1991) generated a panel of anti-ErbB2 antibodies, injected them i.p. into nude mice and evaluated their effect on tumor growth of murine fibroblasts transformed by overexpression of the erbB2 gene. Various levels of tumor inhibition were detected for four of the antibodies, but one of the antibodies (N28) consistently stimulated tumor growth. Monoclonal antibody N28 induced significant phosphorylation of the ErbB2 receptor, whereas the other four antibodies generally displayed low or no phosphorylation-inducing activity. The effect of the anti-ErbB2 antibodies on proliferation of SKBR3 cells was also assessed. In this SKBR3 cell proliferation assay, two of the antibodies (N12 and N29) caused a reduction in cell proliferation relative to control. The ability of the various antibodies to induce cell lysis in vitro via complement-dependent cytotoxicity (CDC) and antibody-mediated cell-dependent cytotoxicity (ADCC) was assessed, with the authors of



US 7,892,549 B2

3

this paper concluding that the inhibitory function of the antibodies was not attributed significantly to CDC or ADCC.

Bacus et al. *Cancer Research* 52:2580-2589 (1992) further characterized the antibodies described in Bacus et al. (1990) and Stancovski et al. of the preceding paragraphs. Extending the i.p. studies of Stancovski et al., the effect of the antibodies after i.v. injection into nude mice harboring mouse fibroblasts overexpressing human ErbB2 was assessed. As observed in their earlier work, N28 accelerated tumor growth whereas N12 and N29 significantly inhibited growth of the ErbB2-expressing cells. Partial tumor inhibition was also observed with the N24 antibody. Bacus et al. also tested the ability of the antibodies to promote a mature phenotype in the human breast cancer cell lines AU-565 and MDA-MB453 (which overexpress ErbB2) as well as MCF-7 (containing low levels of the receptor). Bacus et al. saw a correlation between tumor inhibition in vivo and cellular differentiation; the tumor-stimulatory antibody N28 had no effect on differentiation, and the tumor inhibitory action of the N12, N29 and N24 antibodies correlated with the extent of differentiation they induced.

Xu et al. *Int. J. Cancer* 53:401-408 (1993) evaluated a panel of anti-ErbB2 antibodies for their epitope binding specificities, as well as their ability to inhibit anchorage-independent and anchorage-dependent growth of SKBR3 cells (by individual antibodies and in combinations), modulate cell-surface ErbB2, and inhibit ligand stimulated anchorage-independent growth. See also WO94/00136 published Jan. 6, 1994 and Kasprzyk et al. *Cancer Research* 52:2771-2776 (1992) concerning anti-ErbB2 antibody combinations. Other anti-ErbB2 antibodies are discussed in Hancock et al. *Cancer Res.* 51:4575-4580 (1991); Shawver et al. *Cancer Res.* 54:1367-1373 (1994); Arteaga et al. *Cancer Res.* 54:3758-3765 (1994); and Harwerth et al. *J. Biol. Chem.* 267:15160-15167 (1992).

A recombinant humanized anti-ErbB2 monoclonal antibody (a humanized version of the murine anti-ErbB2 antibody 4D5, referred to as rhuMab HER2 or HERCEPTIN®) has been clinically active in patients with ErbB2-overexpressing metastatic breast cancers that had received extensive prior anti-cancer therapy (Baselga et al., *J. Clin. Oncol.* 14:737-744 [1996]).

ErbB2 overexpression is commonly regarded as a predictor of a poor prognosis, especially in patients with primary disease that involves axillary lymph nodes (Slamon et al., [1987] and [1989], supra Ravdin and Chamness, *Gene* 159:19-27 [1995]; and Hynes and Stern, *Biochim Biophys Acta* 1198: 165-184 [1994]), and has been linked to sensitivity and/or resistance to hormone therapy and chemotherapeutic regimens, including CMF (cyclophosphamide, methotrexate, and fluoruracil) and anthracyclines (Baselga et al., *Oncology* 11(3 Suppl 2):43-48 [1997]). However, despite the association of ErbB2 overexpression with poor prognosis, the odds of HER2-positive patients responding clinically to treatment with taxanes were greater than three times those of HER2-negative patients (Ibid). rhuMab HER2 was shown to enhance the activity of paclitaxel (TAXOL®) and doxorubicin against breast cancer xenografts in nude mice injected with BT-474 human breast adenocarcinoma cells, which express high levels of HER2 (Baselga et al., *Breast Cancer, Proceedings of ASCO*, Vol. 13, Abstract 53 [1994]).

#### SUMMARY OF THE INVENTION

The present invention concerns the treatment of disorders characterized by overexpression of ErbB2, and is based on the recognition that while treatment with anti-ErbB2 antibodies

4

markedly enhances the clinical benefit of the use of chemotherapeutic agents in general, a syndrome of myocardial dysfunction that has been observed as a side-effect of anthracycline derivatives is increased by the administration of anti-ErbB2 antibodies.

Accordingly, the invention concerns a method for the treatment of a human patient susceptible to or diagnosed with a disorder characterized by overexpression of ErbB2 receptor comprising administering a therapeutically effective amount of a combination of an anti-ErbB2 antibody and a chemotherapeutic agent other than an anthracycline derivative, e.g. doxorubicin or epirubicin, in the absence of an anthracycline derivative, to the human patient.

The disorder preferably is a benign or malignant tumor characterized by the overexpression of the ErbB2 receptor, e.g. a cancer, such as, breast cancer, squamous cell cancer, small-cell lung cancer, non-small cell lung cancer, gastrointestinal cancer, pancreatic cancer, glioblastoma, cervical cancer, ovarian cancer, liver cancer, bladder cancer, hepatoma, colon cancer, colorectal cancer, endometrial carcinoma, salivary gland carcinoma, kidney cancer, liver cancer, prostate cancer, vulval cancer, thyroid cancer, hepatic carcinoma and various types of head and neck cancer. The chemotherapeutic agent preferably is a taxoid, such as TAXOL® (paclitaxel) or a TAXOL® derivative.

Although an antiproliferative effect is sufficient, in a preferred embodiment, the anti-ErbB2 antibody is capable of inducing cell death or is capable of inducing apoptosis. Preferred anti-ErbB2 antibodies bind the extracellular domain of the ErbB2 receptor, and preferably bind to the epitope 4D5 or 3H4 within the ErbB2 extracellular domain sequence. More preferably, the antibody is the antibody 4D5, most preferably in a humanized form.

The method of the present invention is particularly suitable for the treatment of breast or ovarian cancer, characterized by the overexpression of the ErbB2 receptor.

In another aspect, the invention concerns an article of manufacture, comprising a container, a composition within the container comprising an anti-ErbB2 antibody, optionally a label on or associated with the container that indicates that the composition can be used for treating a condition characterized by overexpression of ErbB2 receptor, and a package insert containing instructions to avoid the use of anthracycline-type chemotherapeutics in combination with the composition.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows epitope-mapping of the extracellular domain of ErbB2 as determined by truncation mutant analysis and site-directed mutagenesis (Nakamura et al. *J. of Virology* 67(10):6179-6191 [October 1993]; Renz et al. *J. Cell Biol.* 125(6):1395-1406 [June 1994]). The anti-proliferative MAbs 4D5 and 3H4 bind adjacent to the transmembrane domain. The various ErbB2-ECD truncations or point mutations were prepared from cDNA using polymerase chain reaction technology. The ErbB2 mutants were expressed as gD fusion proteins in a mammalian expression plasmid. This expression plasmid uses the cytomegalovirus promoter/enhancer with SV40 termination and polyadenylation signals located downstream of the inserted cDNA. Plasmid DNA was transfected into 293S cells. One day following transfection, the cells were metabolically labeled overnight in methionine and cysteine-free, low glucose DMEM containing 1% dialyzed fetal bovine serum and 25 µCi each of <sup>35</sup>S methionine and <sup>35</sup>S cysteine. Supernatants were harvested either the ErbB2 MAbs or control antibodies were added to the supernatant



US 7,892,549 B2

5

and incubated 2-4 hours at 4° C. The complexes were precipitated, applied to a 10-20% Tricine SDS gradient gel and electrophoresed at 100 V. The gel was electroblotted onto a membrane and analyzed by autoradiography. SEQ ID NOs:8 and 9 depict the 3H4 and 4D5 epitopes, respectively.

FIG. 2 depicts with underlining the amino acid sequence of Domain 1 of ErbB2 (SEQ ID NO: 1). Bold amino acids indicate the location of the epitope recognized by MAb 7C2 and 7F3 as determined by deletion mapping, i.e. the "7C2/7F3 epitope" (SEQ ID NO:2).

#### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

##### I. Definitions

The terms "HER2", "ErbB2" "c-Erb-B2" are used interchangeably. Unless indicated otherwise, the terms "ErbB2" "c-Erb-B2" and "HER2" when used herein refer to the human protein and "her2", "erbB2" and "c-erb-B2" refer to human gene. The human erbB2 gene and ErbB2 protein are, for example, described in Semba et al., *PNAS* (USA) 82:6497-6501 (1985) and Yamamoto et al. *Nature* 319:230-234 (1986) (Genebank accession number X03363). ErbB2 comprises four domains (Domains 1-4).

The "epitope 4D5" is the region in the extracellular domain of ErbB2 to which the antibody 4D5 (ATCC CRL 10463) binds. This epitope is close to the transmembrane region of ErbB2. To screen for antibodies which bind to the 4D5 epitope, a routine cross-blocking assay such as that described in *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory, Ed Harlow and David Lane (1988), can be performed. Alternatively, epitope mapping can be performed (see FIG. 1) to assess whether the antibody binds to the 4D5 epitope of ErbB2 (i.e. any one or more residues in the region from about residue 529, e.g. about residue 561 to about residue 625, inclusive).

The "epitope 3H4" is the region in the extracellular domain of ErbB2 to which the antibody 3H4 binds. This epitope is shown in FIG. 1, and includes residues from about 541 to about 599, inclusive, in the amino acid sequence of ErbB2 extracellular domain.

The "epitope 7C2/7F3" is the region at the N terminus of the extracellular domain of ErbB2 to which the 7C2 and/or 7F3 antibodies (each deposited with the ATCC, see below) bind. To screen for antibodies which bind to the 7C2/7F3 epitope, a routine cross-blocking assay such as that described in *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory, Ed Harlow and David Lane (1988), can be performed. Alternatively, epitope mapping can be performed to establish whether the antibody binds to the 7C2/7F3 epitope on ErbB2 (i.e. any one or more of residues in the region from about residue 22 to about residue 53 of ErbB2; SEQ ID NO:2).

The term "induces cell death" or "capable of inducing cell death" refers to the ability of the antibody to make a viable cell become nonviable. The "cell" here is one which expresses the ErbB2 receptor, especially where the cell overexpresses the ErbB2 receptor. A cell which "overexpresses" ErbB2 has significantly higher than normal ErbB2 levels compared to a noncancerous cell of the same tissue type. Preferably, the cell is a cancer cell, e.g. a breast, ovarian, stomach, endometrial, salivary gland, lung, kidney, colon, thyroid, pancreatic or bladder cell. In vitro, the cell may be a SKBR3, BT474, Calu 3, MDA-MB-453, MDA-MB-361 or SKOV3 cell. Cell death in vitro may be determined in the absence of complement and immune effector cells to distin-

6

guish cell death induced by antibody dependent cellular cytotoxicity (ADCC) or complement dependent cytotoxicity (CDC). Thus, the assay for cell death may be performed using heat inactivated serum (i.e. in the absence of complement) and in the absence of immune effector cells. To determine whether the antibody is able to induce cell death, loss of membrane integrity as evaluated by uptake of propidium iodide (PI), trypan blue (see Moore et al. *Cytotechnology* 17:1-11 [1995]) or 7AAD can be assessed relative to untreated cells. Preferred cell death-inducing antibodies are those which induce PI uptake in the "PI uptake assay in BT474 cells".

The phrase "induces apoptosis" or "capable of inducing apoptosis" refers to the ability of the antibody to induce programmed cell death as determined by binding of annexin V, fragmentation of DNA, cell shrinkage, dilation of endoplasmic reticulum, cell fragmentation, and/or formation of membrane vesicles (called apoptotic bodies). The cell is one which overexpresses the ErbB2 receptor. Preferably the "cell" is a tumor cell, e.g. a breast, ovarian, stomach, endometrial, salivary gland, lung, kidney, colon, thyroid, pancreatic or bladder cell. In vitro, the cell may be a SKBR3, BT474, Calu 3 cell, MDA-MB-453, MDA-MB-361 or SKOV3 cell. Various methods are available for evaluating the cellular events associated with apoptosis. For example, phosphatidyl serine (PS) translocation can be measured by annexin binding; DNA fragmentation can be evaluated through DNA laddering as disclosed in the example herein; and nuclear/chromatin condensation along with DNA fragmentation can be evaluated by any increase in hypodiploid cells. Preferably, the antibody which induces apoptosis is one which results in about 2 to 50 fold, preferably about 5 to 50 fold, and most preferably about 10 to 50 fold, induction of annexin binding relative to untreated cell in an "annexin binding assay using BT474 cells" (see below).

Sometimes the pro-apoptotic antibody will be one which blocks HRG binding/activation of the ErbB2/ErbB3 complex (e.g. 7F3 antibody). In other situations, the antibody is one which does not significantly block activation of the ErbB2/ErbB3 receptor complex by HRG (e.g. 7C2). Further, the antibody may be one like 7C2 which, while inducing apoptosis, does not induce a large reduction in the percent of cells in S phase (e.g. one which only induces about 0-10% reduction in the percent of these cells relative to control).

The antibody of interest may be one like 7C2 which binds specifically to human ErbB2 and does not significantly cross-react with other proteins such as those encoded by the erbB1, erbB3 and/or erbB4 genes. Sometimes, the antibody may not significantly cross-react with the rat neu protein, e.g., as described in Schechter et al. *Nature* 312:513 (1984) and Drebin et al., *Nature* 312:545-548 (1984). In such embodiments, the extent of binding of the antibody to these proteins (e.g., cell surface binding to endogenous receptor) will be less than about 10% as determined by fluorescence activated cell sorting (FACS) analysis or radioimmunoprecipitation (RIA).

"Heregulin" (HRG) when used herein refers to a polypeptide which activates the ErbB2-ErbB3 and ErbB2-ErbB4 protein complexes (i.e. induces phosphorylation of tyrosine residues in the complex upon binding thereto). Various heregulin polypeptides encompassed by this term are disclosed in Holmes et al., *Science*, 256:1205-1210 (1992); WO 92/20798; Wen et al., *Mol. Cell. Biol.*, 14(3):1909-1919 (1994); and Marchionni et al., *Nature*, 362:312-318 (1993), for example. The term includes biologically active fragments and/or variants of a naturally occurring HRG polypeptide, such as an EGF-like domain fragment thereof (e.g. HRGβ1<sub>177-244</sub>).



US 7,892,549 B2

7

The "ErbB2-ErbB3 protein complex" and "ErbB2-ErbB4 protein complex" are noncovalently associated oligomers of the ErbB2 receptor and the ErbB3 receptor or ErbB4 receptor, respectively. The complexes form when a cell expressing both of these receptors is exposed to HRG and can be isolated by immunoprecipitation and analyzed by SDS-PAGE as described in Sliwkowski et al., *J. Biol. Chem.*, 269(20): 14661-14665 (1994).

"Antibodies" (Abs) and "immunoglobulins" (Igs) are glycoproteins having the same structural characteristics. While antibodies exhibit binding specificity to a specific antigen, immunoglobulins include both antibodies and other antibody-like molecules which lack antigen specificity. Polypeptides of the latter kind are, for example, produced at low levels by the lymph system and at increased levels by myelomas.

"Native antibodies" and "native immunoglobulins" are usually heterotetrameric glycoproteins of about 150,000 daltons, composed of two identical light (L) chains and two identical heavy (H) chains. Each light chain is linked to a heavy chain by one covalent disulfide bond, while the number of disulfide linkages varies among the heavy chains of different immunoglobulin isotypes. Each heavy and light chain also has regularly spaced intrachain disulfide bridges. Each heavy chain has at one end a variable domain ( $V_H$ ) followed by a number of constant domains. Each light chain has a variable domain at one end ( $V_L$ ) and a constant domain at its other end; the constant domain of the light chain is aligned with the first constant domain of the heavy chain, and the light-chain variable domain is aligned with the variable domain of the heavy chain. Particular amino acid residues are believed to form an interface between the light- and heavy-chain variable domains.

The term "variable" refers to the fact that certain portions of the variable domains differ extensively in sequence among antibodies and are used in the binding and specificity of each particular antibody for its particular antigen. However, the variability is not evenly distributed throughout the variable domains of antibodies. It is concentrated in three segments called complementarity-determining regions (CDRs) or hypervariable regions both in the light-chain and the heavy-chain variable domains. The more highly conserved portions of variable domains are called the framework region (FR). The variable domains of native heavy and light chains each comprise four FR regions, largely adopting a  $\beta$ -sheet configuration, connected by three CDRs, which form loops connecting, and in some cases forming part of, the  $\beta$ -sheet structure. The CDRs in each chain are held together in close proximity by the FRs and, with the CDRs from the other chain, contribute to the formation of the antigen-binding site of antibodies (see Kabat et al., *NIH Publ. No.* 91-3242, Vol. 1, pages 647-669 [1991]). The constant domains are not involved directly in binding an antibody to an antigen, but exhibit various effector functions, such as participation of the antibody in antibody dependent cellular cytotoxicity.

Papain digestion of antibodies produces two identical antigen-binding fragments, called "Fab" fragments, each with a single antigen-binding site, and a residual "Fc" fragment, whose name reflects its ability to crystallize readily. Pepsin treatment yields an  $F(ab')_2$  fragment that has two antigen-combining sites and is still capable of cross-linking antigen.

"Fv" is the minimum antibody fragment which contains a complete antigen-recognition and -binding site. This region consists of a dimer of one heavy- and one light-chain variable domain in tight, non-covalent association. It is in this configuration that the three CDRs of each variable domain interact to define an antigen-binding site on the surface of the  $V_H$ - $V_L$  dimer. Collectively, the six CDRs confer antigen-

8

binding specificity to the antibody. However, even a single variable domain (or half of an Fv comprising only three CDRs specific for an antigen) has the ability to recognize and bind antigen, although at a lower affinity than the entire binding site.

The Fab fragment also contains the constant domain of the light chain and the first constant domain (CH1) of the heavy chain. Fab' fragments differ from Fab fragments by the addition of a few residues at the carboxy terminus of the heavy chain CH1 domain including one or more cysteines from the antibody hinge region. Fab'-SH is the designation herein for Fab' in which the cysteine residue(s) of the constant domains bear a free thiol group.  $F(ab')_2$  antibody fragments originally were produced as pairs of Fab' fragments which have hinge cysteines between them. Other chemical couplings of antibody fragments are also known.

The "light chains" of antibodies (immunoglobulins) from any vertebrate species can be assigned to one of two clearly distinct types, called kappa ( $\kappa$ ) and lambda ( $\lambda$ ), based on the amino acid sequences of their constant domains.

Depending on the amino acid sequence of the constant domain of their heavy chains, immunoglobulins can be assigned to different classes. There are five major classes of immunoglobulins: IgA, IgD, IgE, IgG, and IgM, and several of these may be further divided into subclasses (isotypes), e.g., IgG1, IgG2, IgG3, IgG4, IgA, and IgA2. The heavy-chain constant domains that correspond to the different classes of immunoglobulins are called  $\alpha$ ,  $\delta$ ,  $\epsilon$ ,  $\gamma$ , and  $\mu$ , respectively. The subunit structures and three-dimensional configurations of different classes of immunoglobulins are well known.

The term "antibody" is used in the broadest sense and specifically covers intact monoclonal antibodies, polyclonal antibodies, multispecific antibodies (e.g. bispecific antibodies) formed from at least two intact antibodies, and antibody fragments so long as they exhibit the desired biological activity.

"Antibody fragments" comprise a portion of an intact antibody, preferably the antigen binding or variable region of the intact antibody. Examples of antibody fragments include Fab, Fab',  $F(ab')_2$ , and Fv fragments; diabodies; linear antibodies (Zapata et al. *Protein Eng.* 8(10):1057-1062 [1995]); single-chain antibody molecules; and multispecific antibodies formed from antibody fragments.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. In addition to their specificity, the monoclonal antibodies are advantageous in that they are synthesized by the hybridoma culture, uncontaminated by other immunoglobulins. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler et al., *Nature*, 256:495 (1975), or may be made by recombinant DNA methods (see, e.g., U.S. Pat. No. 4,816,567). The "monoclonal antibodies"



US 7,892,549 B2

9

may also be isolated from phage antibody libraries using the techniques described in Clackson et al., *Nature*, 352:624-628 (1991) and Marks et al., *J. Mol. Biol.*, 222:581-597 (1991), for example.

The monoclonal antibodies herein specifically include "chimeric" antibodies (immunoglobulins) in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (U.S. Pat. No. 4,816,567; Morrison et al., *Proc. Natl. Acad. Sci. USA*, 81:6851-6855 [1984]).

"Humanized" forms of non-human (e.g., murine) antibodies are chimeric immunoglobulins, immunoglobulin chains or fragments thereof (such as Fv, Fab, Fab', F(ab')<sub>2</sub> or other antigen-binding subsequences of antibodies) which contain minimal sequence derived from non-human immunoglobulin. For the most part, humanized antibodies are human immunoglobulins (recipient antibody) in which residues from a complementarity determining region (CDR) of the recipient are replaced by residues from a CDR of a non-human species (donor antibody) such as mouse, rat or rabbit having the desired specificity, affinity, and capacity. In some instances, Fv framework region (FR) residues of the human immunoglobulin are replaced by corresponding non-human residues. Furthermore, humanized antibodies may comprise residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. These modifications are made to further refine and maximize antibody performance. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the CDRs correspond to those of a non-human immunoglobulin and all or substantially all of the FRs are those of a human immunoglobulin sequence. The humanized antibody optimally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. For further details, see Jones et al., *Nature*, 321:522-525 (1986); Reichmann et al., *Nature*, 332:323-329 (1988); and Presta, *Curr. Op. Struct. Biol.*, 2:593-596 (1992). The humanized antibody includes a PRIMATIZED™ antibody wherein the antigen-binding region of the antibody is derived from an antibody produced by immunizing macaque monkeys with the antigen of interest.

"Single-chain Fv" or "sFv" antibody fragments comprise the V<sub>H</sub> and V<sub>L</sub> domains of antibody, wherein these domains are present in a single polypeptide chain. Preferably, the Fv polypeptide further comprises a polypeptide linker between the V<sub>H</sub> and V<sub>L</sub> domains which enables the sFv to form the desired structure for antigen binding. For a review of sFv see Plückthun in *The Pharmacology of Monoclonal Antibodies*, vol. 113, Rosenberg and Moore eds., Springer-Verlag, New York, pp. 269-315 (1994).

The term "diabodies" refers to small antibody fragments with two antigen-binding sites, which fragments comprise a heavy-chain variable domain (V<sub>H</sub>) connected to a light-chain variable domain (V<sub>L</sub>) in the same polypeptide chain (V<sub>H</sub>-V<sub>L</sub>). By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites. Diabodies are described more fully in, for example, EP 404,097; WO 93/11161; and Hollinger et al., *Proc. Natl. Acad. Sci. USA*, 90:6444-6448 (1993).

10

An "isolated" antibody is one which has been identified and separated and/or recovered from a component of its natural environment. Contaminant components of its natural environment are materials which would interfere with diagnostic or therapeutic uses for the antibody, and may include enzymes, hormones, and other proteinaceous or nonproteinaceous solutes. In preferred embodiments, the antibody will be purified (1) to greater than 95% by weight of antibody as determined by the Lowry method, and most preferably more than 99% by weight, (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by use of a spinning cup sequenator, or (3) to homogeneity by SDS-PAGE under reducing or nonreducing conditions using Coomassie blue or, preferably, silver stain. Isolated antibody includes the antibody in situ within recombinant cells since at least one component of the antibody's natural environment will not be present. Ordinarily, however, isolated antibody will be prepared by at least one purification step.

As used herein, the term "salvage receptor binding epitope" refers to an epitope of the Fc region of an IgG molecule (e.g., IgG<sub>1</sub>, IgG<sub>2</sub>, IgG<sub>3</sub>, or IgG<sub>4</sub>) that is responsible for increasing the in vivo serum half-life of the IgG molecule.

"Treatment" refers to both therapeutic treatment and prophylactic or preventative measures. Those in need of treatment include those already with the disorder as well as those in which the disorder is to be prevented.

"Mammal" for purposes of treatment refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, sports, or pet animals, such as dogs, horses, cats, cows, etc. Preferably, the mammal is human.

A "disorder" is any condition that would benefit from treatment with the anti-ErbB2 antibody. This includes chronic and acute disorders or diseases including those pathological conditions which predispose the mammal to the disorder in question. Non-limiting examples of disorders to be treated herein include benign and malignant tumors; leukemias and lymphoid malignancies; neuronal, glial, astrocytic, hypothalamic and other glandular, macrophagal, epithelial, stromal and blastocoele disorders; and inflammatory, angiogenic and immunologic disorders.

The term "therapeutically effective amount" is used to refer to an amount having antiproliferative effect. Preferably, the therapeutically effective amount has apoptotic activity, or is capable of inducing cell death, and preferably death of benign or malignant tumor cells, in particular cancer cells. Efficacy can be measured in conventional ways, depending on the condition to be treated. For cancer therapy, efficacy can, for example, be measured by assessing the time to disease progression (TTP), or determining the response rates (RR) (see the Example below).

The terms "cancer" and "cancerous" refer to or describe the physiological condition in mammals that is typically characterized by unregulated cell growth. Examples of cancer include, but are not limited to, carcinoma, lymphoma, blastoma, sarcoma, and leukemia. More particular examples of such cancers include squamous cell cancer, small-cell lung cancer, non-small cell lung cancer, gastrointestinal cancer, pancreatic cancer, glioblastoma, cervical cancer, ovarian cancer, liver cancer, bladder cancer, hepatoma, breast cancer, colon cancer, colorectal cancer, endometrial carcinoma, salivary gland carcinoma, kidney cancer, liver cancer, prostate cancer, vulval cancer, thyroid cancer, hepatic carcinoma and various types of head and neck cancer.

The term "cytotoxic agent" as used herein refers to a substance that inhibits or prevents the function of cells and/or causes destruction of cells. The term is intended to include radioactive isotopes (e.g. I<sup>131</sup>, I<sup>125</sup>, Y<sup>90</sup> and Re<sup>186</sup>), chemo-



US 7,892,549 B2

11

therapeutic agents, and toxins such as enzymatically active toxins of bacterial, fungal, plant or animal origin, or fragments thereof.

A "chemotherapeutic agent" is a chemical compound useful in the treatment of cancer. Examples of chemotherapeutic agents include adriamycin, doxorubicin, epirubicin, 5-fluorouracil, cytosine arabinoside ("Ara-C"), cyclophosphamide, thiopeta, busulfan, cytosine, taxoids, e.g. paclitaxel (TAXOL®, Bristol-Myers Squibb Oncology, Princeton, N.J.) and docetaxel (TAXOTERE®, Rhône-Poulenc Rorer, Antony, France), methotrexate, cisplatin, melphalan, vinblastine, bleomycin, etoposide, ifosfamide, mitomycin C, mitoxantrone, vincristine, vinorelbine, carboplatin, teniposide, daunomycin, carminomycin, aminopterin, dactinomycin, mitomycins, esperamicins (see U.S. Pat. No. 4,675,187), melphalan and other related nitrogen mustards. Also included in this definition are hormonal agents that act to regulate or inhibit hormone action on tumors such as tamoxifen and onapristone.

A "growth inhibitory agent" when used herein refers to a compound or composition which inhibits growth of a cell, especially an ErbB2-overexpressing cancer cell either in vitro or in vivo. Thus, the growth inhibitory agent is one which significantly reduces the percentage of ErbB2 overexpressing cells in S phase. Examples of growth inhibitory agents include agents that block cell cycle progression (at a place other than S phase), such as agents that induce G1 arrest and M-phase arrest. Classical M-phase blockers include the vincas (vincristine and vinblastine), TAXOL®, and topo II inhibitors such as doxorubicin, epirubicin, daunorubicin, etoposide, and bleomycin. Those agents that arrest G1 also spill over into S-phase arrest, for example, DNA alkylating agents such as tamoxifen, prednisone, dacarbazine, mechlorethamine, cisplatin, methotrexate, 5-fluorouracil, and ara-C. Further information can be found in *The Molecular Basis of Cancer*, Mendelsohn and Israel, eds., Chapter 1, entitled "Cell cycle regulation, oncogenes, and antineoplastic drugs" by Murakami et al. (WB Saunders: Philadelphia, 1995), especially p. 13. The 4D5 antibody (and functional equivalents thereof) can also be employed for this purpose.

"Doxorubicin" is an anthracycline antibiotic. The full chemical name of doxorubicin is (8S-cis)-10-[(3-amino-2,3,6-trideoxy- $\alpha$ -L-lyxo-hexopyranosyl)oxy]-7,8,9,10-tetrahydro-6,8,11-trihydroxy-8-(hydroxyacetyl)-1-methoxy-5,12-naphthacenedione.

The term "cytokine" is a generic term for proteins released by one cell population which act on another cell as intercellular mediators. Examples of such cytokines are lymphokines, monokines, and traditional polypeptide hormones. Included among the cytokines are growth hormone such as human growth hormone, N-methionyl human growth hormone, and bovine growth hormone; parathyroid hormone; thyroxine; insulin; proinsulin; relaxin; prolaxin; glycoprotein hormones such as follicle stimulating hormone (FSH), thyroid stimulating hormone (TSH), and luteinizing hormone (LH); hepatic growth factor; fibroblast growth factor; prolactin; placental lactogen; tumor necrosis factor- $\alpha$  and - $\beta$ ; mulierian-inhibiting substance; mouse gonadotropin-associated peptide; inhibin; activin; vascular endothelial growth factor; integrin; thrombopoietin (TPO); nerve growth factors such as NGF- $\beta$ ; platelet-growth factor; transforming growth factors (TGFs) such as TGF- $\alpha$  and TGF- $\beta$ ; insulin-like growth factor-I and -II; erythropoietin (EPO); osteoinductive factors; interferons such as interferon- $\alpha$ , - $\beta$ , and - $\gamma$ ; colony stimulating factors (CSFs) such as macrophage-CSF (M-CSF); granulocyte-macrophage-CSF (GM-CSF); and granulocyte-CSF (G-CSF); interleukins (ILs) such as IL-1, IL-1a, IL-2,

12

IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-11, IL-12; a tumor necrosis factor such as TNF- $\alpha$  or TNF- $\beta$ ; and other polypeptide factors including LIF and kit ligand (KL). As used herein, the term cytokine includes proteins from natural sources or from recombinant cell culture and biologically active equivalents of the native sequence cytokines.

The term "prodrug" as used in this application refers to a precursor or derivative form of a pharmaceutically active substance that is less cytotoxic to tumor cells compared to the parent drug and is capable of being enzymatically activated or converted into the more active parent form. See, e.g., Wilman, "Prodrugs in Cancer Chemotherapy" *Biochemical Society Transactions*, 14, pp. 375-382, 615th Meeting Belfast (1986) and Stella et al., "Prodrugs: A Chemical Approach to Targeted Drug Delivery," *Directed Drug Delivery*, Borchardt et al., (ed.), pp. 247-267, Humana Press (1985). The prodrugs of this invention include, but are not limited to, phosphate-containing prodrugs, thiophosphate-containing prodrugs, sulfate-containing prodrugs, peptide-containing prodrugs, D-amino acid-modified prodrugs, glycosylated prodrugs,  $\beta$ -lactam-containing prodrugs, optionally substituted phenoxyacetamide-containing prodrugs or optionally substituted phenylacetamide-containing prodrugs, 5-fluorocytosine and other 5-fluorouridine prodrugs which can be converted into the more active cytotoxic free drug. Examples of cytotoxic drugs that can be derivatized into a prodrug form for use in this invention include, but are not limited to, those chemotherapeutic agents described above.

By "solid phase" is meant a non-aqueous matrix to which the antibodies used in accordance with the present invention can adhere. Examples of solid phases encompassed herein include those formed partially or entirely of glass (e.g., controlled pore glass), polysaccharides (e.g., agarose), polyacrylamides, polystyrene, polyvinyl alcohol and silicones. In certain embodiments, depending on the context, the solid phase can comprise the well of an assay plate; in others it is a purification column (e.g., an affinity chromatography column). This term also includes a discontinuous solid phase of discrete particles, such as those described in U.S. Pat. No. 4,275,149.

A "liposome" is a small vesicle composed of various types of lipids, phospholipids and/or surfactant which is useful for delivery of a drug (such as the anti-ErbB2 antibodies disclosed herein and, optionally, a chemotherapeutic agent) to a mammal. The components of the liposome are commonly arranged in a bilayer formation, similar to the lipid arrangement of biological membranes.

The term "package insert" is used to refer to instructions customarily included in commercial packages of therapeutic products, that contain information about the indications, usage, dosage, administration, contraindications and/or warnings concerning the use of such therapeutic products.

## II. Production of anti-ErbB2 Antibodies

A description follows as to exemplary techniques for the production of the antibodies used in accordance with the present invention. The ErbB2 antigen to be used for production of antibodies may be, e.g., a soluble form of the extracellular domain of ErbB2 or a portion thereof, containing the desired epitope. Alternatively, cells expressing ErbB2 at their cell surface (e.g. NIH-3T3 cells transformed to overexpress ErbB2; or a carcinoma cell line such as SKBR3 cells, see Stancovski et al. *PNAS* (USA) 88:8691-8695 [1991]) can be used to generate antibodies. Other forms of ErbB2 useful for generating antibodies will be apparent to those skilled in the art.



US 7,892,549 B2

13

## (i) Polyclonal Antibodies

Polyclonal antibodies are preferably raised in animals by multiple subcutaneous (sc) or intraperitoneal (ip) injections of the relevant antigen and an adjuvant. It may be useful to conjugate the relevant antigen to a protein that is immunogenic in the species to be immunized, e.g., keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, or soybean trypsin inhibitor using a bifunctional or derivatizing agent, for example, maleimidobenzoyl sulfosuccinimide ester (conjugation through cysteine residues), N-hydroxysuccinimide (through lysine residues), glutaraldehyde, succinic anhydride,  $\text{SOCl}_2$ , or  $\text{R}_1\text{N}=\text{C}=\text{NR}$ , where R and  $\text{R}^1$  are different alkyl groups.

Animals are immunized against the antigen, immunogenic conjugates, or derivatives by combining, e.g., 100  $\mu\text{g}$  or 5  $\mu\text{g}$  of the protein or conjugate (for rabbits or mice, respectively) with 3 volumes of Freund's complete adjuvant and injecting the solution intradermally at multiple sites. One month later the animals are boosted with  $\frac{1}{5}$  to  $\frac{1}{10}$  the original amount of peptide or conjugate in Freund's complete adjuvant by subcutaneous injection at multiple sites. Seven to 14 days later the animals are bled and the serum is assayed for antibody titer. Animals are boosted until the titer plateaus. Preferably, the animal is boosted with the conjugate of the same antigen, but conjugated to a different protein and/or through a different cross-linking reagent. Conjugates also can be made in recombinant cell culture as protein fusions. Also, aggregating agents such as alum are suitably used to enhance the immune response.

## (ii) Monoclonal Antibodies

Monoclonal antibodies are obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except for possible naturally occurring mutations that may be present in minor amounts. Thus, the modifier "monoclonal" indicates the character of the antibody as not being a mixture of discrete antibodies.

For example, the monoclonal antibodies may be made using the hybridoma method first described by Kohler et al., *Nature*, 256:495 (1975), or may be made by recombinant DNA methods (U.S. Pat. No. 4,816,567).

In the hybridoma method, a mouse or other appropriate host animal, such as a hamster, is immunized as hereinabove described to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the protein used for immunization. Alternatively, lymphocytes may be immunized in vitro. Lymphocytes then are fused with myeloma cells using a suitable fusing agent, such as polyethylene glycol, to form a hybridoma cell (Goding, *Monoclonal Antibodies: Principles and Practice*, pp. 59-103 [Academic Press, 1986]).

The hybridoma cells thus prepared are seeded and grown in a suitable culture medium that preferably contains one or more substances that inhibit the growth or survival of the unfused, parental myeloma cells. For example, if the parental myeloma cells lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT or HPRT), the culture medium for the hybridomas typically will include hypoxanthine, aminopterin, and thymidine (HAT medium), which substances prevent the growth of HGPRT-deficient cells.

Preferred myeloma cells are those that fuse efficiently, support stable high-level production of antibody by the selected antibody-producing cells, and are sensitive to a medium such as HAT medium. Among these, preferred myeloma cell lines are murine myeloma lines, such as those derived from MOPC-21 and MPC-11 mouse tumors available from the Salk Institute Cell Distribution Center, San Diego,

14

Calif. USA, and SP-2 or X63-Ag8-653 cells available from the American Type Culture Collection, Rockville, Md. USA. Human myeloma and mouse-human heteromyeloma cell lines also have been described for the production of human monoclonal antibodies (Kozbor, *J. Immunol.*, 133:3001 (1984); Brodeur et al., *Monoclonal Antibody Production Techniques and Applications*, pp. 51-63 [Marcel Dekker, Inc., New York, 1987]).

Culture medium in which hybridoma cells are growing is assayed for production of monoclonal antibodies directed against the antigen. Preferably, the binding specificity of monoclonal antibodies produced by hybridoma cells is determined by immunoprecipitation or by an in vitro binding assay, such as radioimmunoassay (RIA) or enzyme-linked immunoabsorbent assay (ELISA).

The binding affinity of the monoclonal antibody can, for example, be determined by the Scatchard analysis of Munson et al., *Anal. Biochem.*, 107:220 (1980).

After hybridoma cells are identified that produce antibodies of the desired specificity, affinity, and/or activity, the clones may be subcloned by limiting dilution procedures and grown by standard methods (Goding, *Monoclonal Antibodies: Principles and Practice*, pp. 59-103 [Academic Press, 1986]). Suitable culture media for this purpose include, for example, D-MEM or RPMI-1640 medium. In addition, the hybridoma cells may be grown in vivo as ascites tumors in an animal.

The monoclonal antibodies secreted by the subclones are suitably separated from the culture medium, ascites fluid, or serum by conventional immunoglobulin purification procedures such as, for example, protein A-Sepharose, hydroxylapatite chromatography, gel electrophoresis, dialysis, or affinity chromatography.

DNA encoding the monoclonal antibodies is readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). The hybridoma cells serve as a preferred source of such DNA. Once isolated, the DNA may be placed into expression vectors, which are then transfected into host cells such as *E. coli* cells, simian COS cells, Chinese Hamster Ovary (CHO) cells, or myeloma cells that do not otherwise produce immunoglobulin protein, to obtain the synthesis of monoclonal antibodies in the recombinant host cells. Review articles on recombinant expression in bacteria of DNA encoding the antibody include Skerra et al., *Curr. Opinion in Immunol.*, 5:256-262 (1993) and Pluckthun, *Immunol. Revs.*, 130: 151-188 (1992).

In a further embodiment, antibodies or antibody fragments can be isolated from antibody phage libraries generated using the techniques described in McCafferty et al., *Nature*, 348: 552-554 (1990). Clackson et al., *Nature*, 352:624-628 (1991) and Marks et al., *J. Mol. Biol.*, 222:581-597 (1991) describe the isolation of murine and human antibodies, respectively, using phage libraries. Subsequent publications describe the production of high affinity (nM range) human antibodies by chain shuffling (Marks et al., *Bio/Technology*, 10:779-783 [1992]), as well as combinatorial infection and in vivo recombination as a strategy for constructing very large phage libraries (Waterhouse et al., *Nuc. Acids. Res.*, 21:2265-2266 [1993]). Thus, these techniques are viable alternatives to traditional monoclonal antibody hybridoma techniques for isolation of monoclonal antibodies.

The DNA also may be modified, for example, by substituting the coding sequence for human heavy- and light-chain constant domains in place of the homologous murine sequences (U.S. Pat. No. 4,816,567; Morrison, et al., *Proc.*



US 7,892,549 B2

15

*Natl. Acad. Sci. USA*, 81:6851 [1984]), or by covalently joining to the immunoglobulin coding sequence all or part of the coding sequence for a non-immunoglobulin polypeptide.

Typically such non-immunoglobulin polypeptides are substituted for the constant domains of an antibody, or they are substituted for the variable domains of one antigen-combining site of an antibody to create a chimeric bivalent antibody comprising one antigen-combining site having specificity for an antigen and another antigen-combining site having specificity for a different antigen.

#### (iii) Humanized and Human Antibodies

Methods for humanizing non-human antibodies are well known in the art. Preferably, a humanized antibody has one or more amino acid residues introduced into it from a source which is non-human. These non-human amino acid residues are often referred to as "import" residues, which are typically taken from an "import" variable domain. Humanization can be essentially performed following the method of Winter and co-workers (Jones et al., *Nature*, 321:522-525 (1986); Riechmann et al., *Nature*, 332:323-327 (1988); Verhoeven et al., *Science*, 239:1534-1536 [1988]), by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Accordingly, such "humanized" antibodies are chimeric antibodies (U.S. Pat. No. 4,816,567) wherein substantially less than an intact human variable domain has been substituted by the corresponding sequence from a non-human species. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies.

The choice of human variable domains, both light and heavy, to be used in making the humanized antibodies is very important to reduce antigenicity. According to the so-called "bestfit" method, the sequence of the variable domain of a rodent antibody is screened against the entire library of known human variable-domain sequences. The human sequence which is closest to that of the rodent is then accepted as the human framework region (FR) for the humanized antibody (Sims et al., *J. Immunol.*, 151:2296 (1993); Chothia et al., *J. Mol. Biol.*, 196:901 [1987]). Another method uses a particular framework region derived from the consensus sequence of all human antibodies of a particular subgroup of light or heavy chains. The same framework may be used for several different humanized antibodies (Carter et al., *Proc. Natl. Acad. Sci. USA*, 89:4285 (1992); Presta et al., *J. Immunol.*, 151:2623 [1993]).

It is further important that antibodies be humanized with retention of high affinity for the antigen and other favorable biological properties. To achieve this goal, according to a preferred method, humanized antibodies are prepared by a process of analysis of the parental sequences and various conceptual humanized products using three-dimensional models of the parental and humanized sequences. Three-dimensional immunoglobulin models are commonly available and are familiar to those skilled in the art. Computer programs are available which illustrate and display probable three-dimensional conformational structures of selected candidate immunoglobulin sequences. Inspection of these displays permits analysis of the likely role of the residues in the functioning of the candidate immunoglobulin sequence, i.e., the analysis of residues that influence the ability of the candidate immunoglobulin to bind its antigen. In this way, FR residues can be selected and combined from the recipient and import sequences so that the desired antibody characteristic, such as increased affinity for the target antigen(s), is achieved. In general, the CDR residues are directly and most substantially involved in influencing antigen binding.

16

Alternatively, it is now possible to produce transgenic animals (e.g., mice) that are capable, upon immunization, of producing a full repertoire of human antibodies in the absence of endogenous immunoglobulin production. For example, it has been described that the homozygous deletion of the antibody heavy-chain joining region ( $J_H$ ) gene in chimeric and germ-line mutant mice results in complete inhibition of endogenous antibody production. Transfer of the human germ-line immunoglobulin gene array in such germ-line mutant mice will result in the production of human antibodies upon antigen challenge. See, e.g., Jakobovits et al., *Proc. Natl. Acad. Sci. USA*, 90:2551 (1993); Jakobovits et al., *Nature*, 362:255-258 (1993); Bruggermann et al., *Year in Immuno.*, 7:33 (1993). Human antibodies can also be derived from phage-display libraries (Hoogenboom et al., *J. Mol. Biol.*, 227:381 (1991); Marks et al., *J. Mol. Biol.*, 222:581-597 [1991]).

#### (iv) Antibody Fragments

Various techniques have been developed for the production of antibody fragments. Traditionally, these fragments were derived via proteolytic digestion of intact antibodies (see, e.g., Morimoto et al., *Journal of Biochemical and Biophysical Methods* 24:107-117 (1992) and Brennan et al., *Science*, 229:81 [1985]). However, these fragments can now be produced directly by recombinant host cells. For example, the antibody fragments can be isolated from the antibody phage libraries discussed above. Alternatively, Fab'-SH fragments can be directly recovered from *E. coli* and chemically coupled to form  $F(ab')_2$  fragments (Carter et al., *Bio/Technology* 10:163-167 [1992]). According to another approach,  $F(ab')_2$  fragments can be isolated directly from recombinant host cell culture. Other techniques for the production of antibody fragments will be apparent to the skilled practitioner. In other embodiments, the antibody of choice is a single chain Fv fragment (scFv). See WO 93/16185.

#### (v) Bispecific Antibodies

Bispecific antibodies are antibodies that have binding specificities for at least two different epitopes. Exemplary bispecific antibodies may bind to two different epitopes of the ErbB2 protein. For example, one arm may bind an epitope in Domain 1 of ErbB2 such as the 7C2/7F3 epitope, the other may bind a different ErbB2 epitope, e.g. the 4D5 epitope. Other such antibodies may combine an ErbB2 binding site with binding site(s) for EGFR, ErbB3 and/or ErbB4. Alternatively, an anti-ErbB2 arm may be combined with an arm which binds to a triggering molecule on a leukocyte such as a T-cell receptor molecule (e.g. CD2 or CD3), or Fc receptors for IgG (FcγR), such as FcγRI (CD64), FcγRII (CD32) and FcγRIII (CD16) so as to focus cellular defense mechanisms to the ErbB2-expressing cell. Bispecific antibodies may also be used to localize cytotoxic agents to cells which express ErbB2. These antibodies possess an ErbB2-binding arm and an arm which binds the cytotoxic agent (e.g. saporin, anti-interferon-α, vinca alkaloid, ricin A chain, methotrexate or radioactive isotope hapten). Bispecific antibodies can be prepared as full length antibodies or antibody fragments (e.g.  $F(ab')_2$  bispecific antibodies).

Methods for making bispecific antibodies are known in the art. Traditional production of full length bispecific antibodies is based on the coexpression of two immunoglobulin heavy chain-light chain pairs, where the two chains have different specificities (Millstein et al., *Nature*, 305:537-539 [1983]). Because of the random assortment of immunoglobulin heavy and light chains, these hybridomas (quadromas) produce a potential mixture of 10 different antibody molecules, of which only one has the correct bispecific structure. Purification of the correct molecule, which is usually done by affinity



US 7,892,549 B2

17

chromatography steps, is rather cumbersome, and the product yields are low. Similar procedures are disclosed in WO 93/08829, and in Traunecker et al., *EMBO J.*, 10:3655-3659 (1991).

According to a different approach, antibody variable domains with the desired binding specificities (antibody-antigen combining sites) are fused to immunoglobulin constant domain sequences. The fusion preferably is with an immunoglobulin heavy chain constant domain, comprising at least part of the hinge, CH2, and CH3 regions. It is preferred to have the first heavy-chain constant region (CH1) containing the site necessary for light chain binding, present in at least one of the fusions. DNAs encoding the immunoglobulin heavy chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host organism. This provides for great flexibility in adjusting the mutual proportions of the three polypeptide fragments in embodiments when unequal ratios of the three polypeptide chains used in the construction provide the optimum yields. It is, however, possible to insert the coding sequences for two or all three polypeptide chains in one expression vector when the expression of at least two polypeptide chains in equal ratios results in high yields or when the ratios are of no particular significance.

In a preferred embodiment of this approach, the bispecific antibodies are composed of a hybrid immunoglobulin heavy chain with a first binding specificity in one arm, and a hybrid immunoglobulin heavy chain-light chain pair (providing a second binding specificity) in the other arm. It was found that this asymmetric structure facilitates the separation of the desired bispecific compound from unwanted immunoglobulin chain combinations, as the presence of an immunoglobulin light chain in only one half of the bispecific molecule provides for a facile way of separation. This approach is disclosed in WO 94/04690. For further details of generating bispecific antibodies see, for example, Suresh et al., *Methods in Enzymology*, 121:210 (1986).

According to another approach described in WO96/27011, the interface between a pair of antibody molecules can be engineered to maximize the percentage of heterodimers which are recovered from recombinant cell culture. The preferred interface comprises at least a part of the C<sub>H</sub>3 domain of an antibody constant domain. In this method, one or more small amino acid side chains from the interface of the first antibody molecule are replaced with larger side chains (e.g., tyrosine or tryptophan). Compensatory "cavities" of identical or similar size to the large side chain(s) are created on the interface of the second antibody molecule by replacing large amino acid side chains with smaller ones (e.g., alanine or threonine). This provides a mechanism for increasing the yield of the heterodimer over other unwanted end-products such as homodimers.

Bispecific antibodies include cross-linked or "heteroconjugate" antibodies. For example, one of the antibodies in the heteroconjugate can be coupled to avidin, the other to biotin. Such antibodies have, for example, been proposed to target immune system cells to unwanted cells (U.S. Pat. No. 4,676,980), and for treatment of HIV infection (WO 91/00360, WO 92/200373, and EP 03089). Heteroconjugate antibodies may be made using any convenient cross-linking methods. Suitable cross-linking agents are well known in the art, and are disclosed in U.S. Pat. No. 4,676,980, along with a number of cross-linking techniques.

Techniques for generating bispecific antibodies from antibody fragments have also been described in the literature. For example, bispecific antibodies can be prepared using chemical linkage. Brennan et al., *Science*, 229:81 (1985) describe a

18

procedure wherein intact antibodies are proteolytically cleaved to generate F(ab')<sub>2</sub> fragments. These fragments are reduced in the presence of the dithiol complexing agent sodium arsenite to stabilize vicinal dithiols and prevent intermolecular disulfide formation. The Fab' fragments generated are then converted to thionitrobenzoate (TNB) derivatives. One of the Fab'-TNB derivatives is then reconverted to the Fab'-thiol by reduction with mercaptoethylamine and is mixed with an equimolar amount of the other Fab'-TNB derivative to form the bispecific antibody. The bispecific antibodies produced can be used as agents for the selective immobilization of enzymes.

Recent progress has facilitated the direct recovery of Fab'-SH fragments from *E. coli*, which can be chemically coupled to form bispecific antibodies. Shalaby et al., *J. Exp. Med.*, 175:217-225 (1992) describe the production of a fully humanized bispecific antibody F(ab')<sub>2</sub> molecule. Each Fab' fragment was separately secreted from *E. coli* and subjected to directed chemical coupling in vitro to form the bispecific antibody. The bispecific antibody thus formed was able to bind to cells overexpressing the ErbB2 receptor and normal human T cells, as well as trigger the lytic activity of human cytotoxic lymphocytes against human breast tumor targets.

Various techniques for making and isolating bispecific antibody fragments directly from recombinant cell culture have also been described. For example, bispecific antibodies have been produced using leucine zippers. Kostelny et al., *J. Immunol.*, 148(5):1547-1553 (1992). The leucine zipper peptides from the Fos and Jun proteins were linked to the Fab' portions of two different antibodies by gene fusion. The antibody homodimers were reduced at the hinge region to form monomers and then re-oxidized to form the antibody heterodimers. This method can also be utilized for the production of antibody homodimers. The "diabody" technology described by Hollinger et al., *Proc. Natl. Acad. Sci. USA*, 90:6444-6448 (1993) has provided an alternative mechanism for making bispecific antibody fragments. The fragments comprise a heavy-chain variable domain (V<sub>H</sub>) connected to a light-chain variable domain (V<sub>L</sub>) by a linker which is too short to allow pairing between the two domains on the same chain. Accordingly, the V<sub>H</sub> and V<sub>L</sub> domains of one fragment are forced to pair with the complementary V<sub>L</sub> and V<sub>H</sub> domains of another fragment, thereby forming two antigen-binding sites. Another strategy for making bispecific antibody fragments by the use of single-chain Fv (sFv) dimers has also been reported. See Gruber et al., *J. Immunol.*, 152:5368 (1994).

Antibodies with more than two valencies are contemplated. For example, trispecific antibodies can be prepared. Tutt et al. *J. Immunol.* 147:60 (1991).

(vi) Screening for Antibodies with the Desired Properties

Techniques for generating antibodies have been described above. Those antibodies having the characteristics described herein are selected.

To select for antibodies which induce cell death, loss of membrane integrity as indicated by, e.g., PI, trypan blue or 7AAD uptake is assessed relative to control. The preferred assay is the "PI uptake assay using BT474 cells". According to this assay, BT474 cells (which can be obtained from the American Type Culture Collection [Rockville, Md.]) are cultured in Dulbecco's Modified Eagle Medium (D-MEM): Ham's F-12 (50:50) supplemented with 10% heat-inactivated FBS (Hyclone) and 2 mM L-glutamine. (Thus, the assay is performed in the absence of complement and immune effector cells). The BT474 cells are seeded at a density of 3×10<sup>6</sup> per dish in 100×20 mm dishes and allowed to attach overnight. The medium is then removed and replaced with fresh medium alone or medium containing 10 µg/ml of the appropriate



US 7,892,549 B2

19

MAB. The cells are incubated for a 3 day time period. Following each treatment, monolayers are washed with PBS and detached by trypsinization. Cells are then centrifuged at 1200 rpm for 5 minutes at 4° C., the pellet resuspended in 3 ml ice cold Ca<sup>2+</sup> binding buffer (10 mM Hepes, pH 7.4, 140 mM NaCl, 2.5 mM CaCl<sub>2</sub>) and aliquoted into 35 mm strainer-capped 12x75 tubes (1 ml per tube, 3 tubes per treatment group) for removal of cell clumps. Tubes then receive PI (10 µg/ml). Samples may be analyzed using a FACSCAN™ flow cytometer and FACSCONVERT™ CellQuest software (Becton Dickinson). Those antibodies which induce statistically significant levels of cell death as determined by PI uptake are selected.

In order to select for antibodies which induce apoptosis, an "annexin binding assay using BT474 cells" is available. The BT474 cells are cultured and seeded in dishes as discussed in the preceding paragraph. The medium is then removed and replaced with fresh medium alone or medium containing 10 µg/ml of the MAB. Following a three day incubation period, monolayers are washed with PBS and detached by trypsinization. Cells are then centrifuged, resuspended in Ca<sup>2+</sup> binding buffer and aliquoted into tubes as discussed above for the cell death assay. Tubes then receive labeled annexin (e.g. annexin V-FITC) (1 µg/ml). Samples may be analyzed using a FACSCAN™ flow cytometer and FACSCONVERT™ CellQuest software (Becton Dickinson). Those antibodies which induce statistically significant levels of annexin binding relative to control are selected as apoptosis-inducing antibodies.

In addition to the annexin binding assay, a "DNA staining assay using BT474 cells" is available. In order to perform this assay, BT474 cells which have been treated with the antibody of interest as described in the preceding two paragraphs are incubated with 9 µg/ml HOECHST 33342™ for 2 hr at 37° C., then analyzed on an EPICS ELITE™ flow cytometer (Coulter Corporation) using MODFIT LT™ software (Verity Software House). Antibodies which induce a change in the percentage of apoptotic cells which is 2 fold or greater (and preferably 3 fold or greater) than untreated cells (up to 100% apoptotic cells) may be selected as pro-apoptotic antibodies using this assay.

To screen for antibodies which bind to an epitope on ErbB2 bound by an antibody of interest, a routine cross-blocking assay such as that described in *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory, Ed Harlow and David Lane (1988), can be performed. Alternatively, epitope mapping can be performed by methods known in the art.

To identify anti-ErbB2 antibodies which inhibit growth of SKBR3 cells in cell culture by 50-100%, the SKBR3 assay described in WO89/06692 can be performed. According to this assay, SKBR3 cells are grown in a 1:1 mixture of F12 and DMEM medium supplemented with 10% fetal bovine serum, glutamine and penicillin/streptomycin. The SKBR3 cells are plated at 20,000 cells in a 35 mm cell culture dish (2 mls/35 mm dish). 2.5 µg/ml of the anti-ErbB2 antibody is added per dish. After six days, the number of cells, compared to untreated cells are counted using an electronic COULTER™ cell counter. Those antibodies which inhibit growth of the SKBR3 cells by 50-100% are selected for combination with the apoptotic antibodies as desired.

#### (vii) Effector Function Engineering

It may be desirable to modify the antibody of the invention with respect to effector function, so as to enhance the effectiveness of the antibody in treating cancer, for example. For example cysteine residue(s) may be introduced in the Fc region, thereby allowing interchain disulfide bond formation in this region. The homodimeric antibody thus generated may have improved internalization capability and/or increased

20

complement-mediated cell killing and antibody-dependent cellular cytotoxicity (ADCC). See Caron et al., *J. Exp. Med.* 176:1191-1195 (1992) and Shopes, B. *J. Immunol.* 148:2918-2922 (1992). Homodimeric antibodies with enhanced antitumor activity may also be prepared using heterobifunctional cross-linkers as described in Wolff et al. *Cancer Research* 53:2560-2565 (1993). Alternatively, an antibody can be engineered which has dual Fc regions and may thereby have enhanced complement lysis and ADCC capabilities. See Stevenson et al. *Anti-Cancer Drug Design* 3:219-230 (1989).

#### (viii) Immunoconjugates

The invention also pertains to immunoconjugates comprising the antibody described herein conjugated to a cytotoxic agent such as a chemotherapeutic agent, toxin (e.g. an enzymatically active toxin of bacterial, fungal, plant or animal origin, or fragments thereof), or a radioactive isotope (i.e., a radioconjugate).

Chemotherapeutic agents useful in the generation of such immunoconjugates have been described above. Enzymatically active toxins and fragments thereof which can be used include diphtheria A chain, nonbinding active fragments of diphtheria toxin, exotoxin A chain (from *Pseudomonas aeruginosa*), ricin A chain, abrin A chain, modeccin A chain, alpha-sarcin, *Aleurites fordii* proteins, dianthin proteins, *Phytolacca americana* proteins (PAPI, PAPII, and PAP-S), momordica charantia inhibitor, curcin, croton, sapaonaria officinalis inhibitor, gelonin, mitogellin, restrictocin, phenomycin, enomycin and the tricothecenes. A variety of radionuclides are available for the production of radioconjugated anti-ErbB2 antibodies. Examples include <sup>212</sup>Bi, <sup>131</sup>I, <sup>131</sup>In, <sup>90</sup>Y and <sup>186</sup>Re.

Conjugates of the antibody and cytotoxic agent are made using a variety of bifunctional protein coupling agents such as N-succinimidyl-3-(2-pyridyldithiol) propionate (SPDP), iminothiolane (IT), bifunctional derivatives of imidoesters (such as dimethyl adipimidate HCL), active esters (such as disuccinimidyl suberate), aldehydes (such as glutaraldehyde), bis-azido compounds (such as bis (p-azidobenzoyl) hexanediamine), bis-diazonium derivatives (such as bis-(p-diazoniumbenzoyl)-ethylenediamine), diisocyanates (such as tolyene 2,6-diisocyanate), and bis-active fluorine compounds (such as 1,5-difluoro-2,4-dinitrobenzene). For example, a ricin immunotoxin can be prepared as described in Vitetta et al. *Science* 238:1098 (1987). Carbon-14-labeled 1-isothiocyanatobenzyl-3-methyldiethylene triaminepentacetic acid (MXDTPA) is an exemplary chelating agent for conjugation of radionucleotide to the antibody. See WO94/11026.

In another embodiment, the antibody may be conjugated to a "receptor" (such streptavidin) for utilization in tumor pre-targeting wherein the antibody-receptor conjugate is administered to the patient, followed by removal of unbound conjugate from the circulation using a clearing agent and then administration of a "ligand" (e.g. avidin) which is conjugated to a cytotoxic agent (e.g. a radionucleotide).

#### (ix) Immunoliposomes

The anti-ErbB2 antibodies disclosed herein may also be formulated as immunoliposomes. Liposomes containing the antibody are prepared by methods known in the art, such as described in Epstein et al., *Proc. Natl. Acad. Sci. USA*, 82:3688 (1985); Hwang et al., *Proc. Natl. Acad. Sci. USA*, 77:4030 (1980); and U.S. Pat. Nos. 4,485,045 and 4,544,545. Liposomes with enhanced circulation time are disclosed in U.S. Pat. No. 5,013,556.

Particularly useful liposomes can be generated by the reverse phase evaporation method with a lipid composition comprising phosphatidylcholine, cholesterol and PEG-de-



US 7,892,549 B2

21

rivatized phosphatidylethanolamine (PEG-PE). Liposomes are extruded through filters of defined pore size to yield liposomes with the desired diameter. Fab' fragments of the antibody of the present invention can be conjugated to the liposomes as described in Martin et al. *Biol. Chem.* 257: 286-288 (1982) via a disulfide interchange reaction. A chemotherapeutic agent is optionally contained within the liposome. See Gabizon et al. *J. National Cancer Inst.* 81(19):1484 (1989).

(x) Antibody Dependent Enzyme Mediated Prodrug Therapy (ADEPT)

The antibodies of the present invention may also be used in ADEPT by conjugating the antibody to a prodrug-activating enzyme which converts a prodrug (e.g. a peptidyl chemotherapeutic agent, see WO81/01145) to an active anti-cancer drug. See, for example, WO. 88/07378 and U.S. Pat. No. 4,975,278.

The enzyme component of the immunoconjugate useful for ADEPT includes any enzyme capable of acting on a prodrug in such a way so as to convert it into its more active, cytotoxic form.

Enzymes that are useful in the method of this invention include, but are not limited to, alkaline phosphatase useful for converting phosphate-containing prodrugs into free drugs; arylsulfatase useful for converting sulfate-containing prodrugs into free drugs; cytosine deaminase useful for converting non-toxic 5-fluorocytosine into the anti-cancer drug, 5-fluorouracil; proteases, such as serratin protease, thermolysin, subtilisin, carboxypeptidases and cathepsins (such as cathepsins B and L), that are useful for converting peptide-containing prodrugs into free drugs; D-alanylcarboxypeptidases, useful for converting prodrugs that contain D-amino acid substituents; carbohydrate-cleaving enzymes such as  $\beta$ -galactosidase and neuraminidase useful for converting glycosylated prodrugs into free drugs;  $\beta$ -lactamase useful for converting drugs derivatized with  $\beta$ -lactams into free drugs; and penicillin amidases, such as penicillin V amidase or penicillin G amidase, useful for converting drugs derivatized at their amine nitrogens with phenoxyacetyl or phenylacetyl groups, respectively, into free drugs. Alternatively, antibodies with enzymatic activity, also known in the art as "abzymes", can be used to convert the prodrugs of the invention into free active drugs (see, e.g., Massey, *Nature* 328:457-458 [1987]). Antibody-abzyme conjugates can be prepared as described herein for delivery of the abzyme to a tumor cell population.

The enzymes of this invention can be covalently bound to the anti-ErbB2 antibodies by techniques well known in the art such as the use of the heterobifunctional crosslinking reagents discussed above. Alternatively, fusion proteins comprising at least the antigen binding region of an antibody of the invention linked to at least a functionally active portion of an enzyme of the invention can be constructed using recombinant DNA techniques well known in the art (see, e.g., Neuberger et al., *Nature*, 312:604-608 [1984]).

(xi) Antibody-salvage Receptor Binding Epitope Fusions

In certain embodiments of the invention, it may be desirable to use an antibody fragment, rather than an intact antibody, to increase tumor penetration, for example. In this case, it may be desirable to modify the antibody fragment in order to increase its serum half life. This may be achieved, for example, by incorporation of a salvage receptor binding epitope into the antibody fragment (e.g. by mutation of the appropriate region in the antibody fragment or by incorporating the epitope into a peptide tag that is then fused to the antibody fragment at either end or in the middle, e.g., by DNA or peptide synthesis).

22

A systematic method for preparing such an antibody variant having an increased in vivo half-life comprises several steps. The first involves identifying the sequence and conformation of a salvage receptor binding epitope of an Fc region of an IgG molecule. Once this epitope is identified, the sequence of the antibody of interest is modified to include the sequence and conformation of the identified binding epitope. After the sequence is mutated, the antibody variant is tested to see if it has a longer in vivo half-life than that of the original antibody. If the antibody variant does not have a longer in vivo half-life upon testing, its sequence is further altered to include the sequence and conformation of the identified binding epitope. The altered antibody is tested for longer in vivo half-life, and this process is continued until a molecule is obtained that exhibits a longer in vivo half-life.

The salvage receptor binding epitope being thus incorporated into the antibody of interest is any suitable such epitope as defined above, and its nature will depend, e.g., on the type of antibody being modified. The transfer is made such that the antibody of interest still possesses the biological activities described herein.

The epitope preferably constitutes a region wherein any one or more amino acid residues from one or two loops of a Fc domain are transferred to an analogous position of the antibody fragment. Even more preferably, three or more residues from one or two loops of the Fc domain are transferred. Still more preferred, the epitope is taken from the CH2 domain of the Fc region (e.g., of an IgG) and transferred to the CH1, CH3, or  $V_H$  region, or more than one such region, of the antibody. Alternatively, the epitope is taken from the CH2 domain of the Fc region and transferred to the  $C_L$  region or  $V_L$  region, or both, of the antibody fragment.

In one most preferred embodiment, the salvage receptor binding epitope comprises the sequence (5' to 3'): PKNS-SMISNTP (SEQ ID NO:3), and optionally further comprises a sequence selected from the group consisting of HQSLGTQ (SEQ ID NO:4), HQNLSDGK (SEQ ID NO:5), HQNISDGK (SEQ ID NO:6), or VISSHLGQ (SEQ ID NO:7), particularly where the antibody fragment is a Fab or F(ab')<sub>2</sub>. In another most preferred embodiment, the salvage receptor binding epitope is a polypeptide containing the sequence(s) (5' to 3'): HQNLSDGK (SEQ ID NO:5), HQNISDGK (SEQ ID NO:6), or VISSHLGQ (SEQ ID NO:7) and the sequence: PKNS-SMISNTP (SEQ ID NO:3).

(xii) Purification of Anti-ErbB2 Antibody

When using recombinant techniques, the antibody can be produced intracellularly, in the periplasmic space, or directly secreted into the medium. If the antibody is produced intracellularly, as a first step, the particulate debris, either host cells or lysed fragments, is removed, for example, by centrifugation or ultrafiltration. Carter et al., *Bio/Technology* 10:163-167 (1992) describe a procedure for isolating antibodies which are secreted to the periplasmic space of *E. coli*. Briefly, cell paste is thawed in the presence of sodium acetate (pH 3.5), EDTA, and phenylmethylsulfonylfluoride (PMSF) over about 30 min. Cell debris can be removed by centrifugation. Where the antibody is secreted into the medium, supernatants from such expression systems are preferably first concentrated using a commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit. A protease inhibitor such as PMSF may be included in any of the foregoing steps to inhibit proteolysis and antibiotics may be included to prevent the growth of adventitious contaminants.

The antibody composition prepared from the cells can be purified using, for example, hydroxylapatite chromatography, gel electrophoresis, dialysis, and affinity chromatography.



US 7,892,549 B2

23

phy, with affinity chromatography being the preferred purification technique. The suitability of protein A as an affinity ligand depends on the species and isotype of any immunoglobulin Fc domain that is present in the antibody. Protein A can be used to purify antibodies that are based on human  $\gamma 1$ ,  $\gamma 2$ , or  $\gamma 4$  heavy chains (Lindmark et al., *J. Immunol. Meth.* 62:1-13 [1983]). Protein G is recommended for all mouse isotypes and for human  $\gamma 3$  (Guss et al., *EMBO J.* 5:15671575 [1986]). The matrix to which the affinity ligand is attached is most often agarose, but other matrices are available. Mechanically stable matrices such as controlled pore glass or poly(styrenedivinyl)benzene allow for faster flow rates and shorter processing times than can be achieved with agarose. Where the antibody comprises a  $C_H3$  domain, the Bakerbond ABX™ resin (J. T. Baker, Phillipsburg, N.J.) is useful for purification. Other techniques for protein purification such as fractionation on an ion-exchange column, ethanol precipitation, Reverse Phase HPLC, chromatography on silica, chromatography on heparin SEPHAROSE™ chromatography on an anion or cation exchange resin (such as a polyaspartic acid column), chromatofocusing, SDS-PAGE, and ammonium sulfate precipitation are also available depending on the antibody to be recovered.

Following any preliminary purification step(s), the mixture comprising the antibody of interest and contaminants may be subjected to low pH hydrophobic interaction chromatography using an elution buffer at a pH between about 2.5-4.5, preferably performed at low salt concentrations (e.g. from about 0-0.25M salt).

### III. Pharmaceutical Formulations

Therapeutic formulations of the antibodies used in accordance with the present invention are prepared for storage by mixing an antibody having the desired degree of purity with optional pharmaceutically acceptable carriers, excipients or stabilizers (*Remington's Pharmaceutical Sciences* 16th edition, Osol, A. Ed. [1980]), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e.g. Zn-protein complexes); and/or non-ionic surfactants such as TWEEN™, PLURONICS™ or polyethylene glycol (PEG).

The formulation herein may also contain more than one active compound as necessary for the particular indication being treated, preferably those with complementary activities that do not adversely affect each other. For example, it may be desirable to further provide antibodies which bind to EGFR, ErbB2 (e.g. an antibody which binds a different epitope on ErbB2), ErbB3, ErbB4, or vascular endothelial factor (VEGF) in the one formulation. Alternatively, or in addition,

24

the composition may comprise a cytotoxic agent, cytokine or growth inhibitory agent, provided that the cytotoxic agent is other than an anthracycline derivative, e.g. doxorubicin, or epirubicin. Such molecules are suitably present in combination in amounts that are effective for the purpose intended.

The active ingredients may also be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsules and poly-(methylmethacrylate) microcapsules, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nanoparticles and nanocapsules) or in macroemulsions. Such techniques are disclosed in *Remington's Pharmaceutical Sciences* 16th edition, Osol, A. Ed. (1980).

The formulations to be used for in vivo administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes.

Sustained-release preparations may be prepared. Suitable examples of sustained-release preparations include semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, e.g. films, or microcapsules. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919), copolymers of L-glutamic acid and  $\gamma$  ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOT™ (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods. When encapsulated antibodies remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at 37° C., resulting in a loss of biological activity and possible changes in immunogenicity. Rational strategies can be devised for stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be intermolecular S—S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

### IV. Treatment with the Anti-ErbB2 Antibodies

It is contemplated that, according to the present invention, the anti-ErbB2 antibodies may be used to treat various conditions characterized by overexpression and/or activation of the ErbB2 receptor. Exemplary conditions or disorders include benign or malignant tumors (e.g. renal, liver, kidney, bladder, breast, gastric, ovarian, colorectal, prostate, pancreatic, lung, vulval, thyroid, hepatic carcinomas; sarcomas; glioblastomas; and various head and neck tumors); leukemias and lymphoid malignancies; other disorders such as neuronal, glial, astrocytic, hypothalamic and other glandular, macrophagal, epithelial, stromal and blastocoelic disorders; and inflammatory, angiogenic and immunologic disorders.

The antibodies of the invention are administered to a human patient, in accord with known methods, such as intravenous administration as a bolus or by continuous infusion over a period of time, by intramuscular, intraperitoneal, intracerebrospinal, subcutaneous, intra-articular, intrasynovial, intrathecal, oral, topical, or inhalation routes. Intravenous administration of the antibody is preferred.



US 7,892,549 B2

25

The treatment of the present invention involved the combined administration of an anti-ErbB2 antibody and a chemotherapeutic agent, other than an anthracycline derivative. The combined administration includes coadministration, using separate formulations or a single pharmaceutical formulation, and consecutive administration in either order, wherein preferably there is a time period while both (or all) active agents simultaneously exert their biological activities. Preparation and dosing schedules for such chemotherapeutic agents may be used according to manufacturers' instructions or as determined empirically by the skilled practitioner. Preparation and dosing schedules for such chemotherapy are also described in *Chemotherapy Service Ed.*, M. C. Perry, Williams & Wilkins, Baltimore, Md. (1992). The chemotherapeutic agent may precede, or follow administration of the antibody or may be given simultaneously therewith. The antibody may be combined with an anti-estrogen compound such as tamoxifen or an anti-progesterone such as onapristone (see, EP 616 812) in dosages known for such molecules.

It may be desirable to also administer antibodies against other tumor associated antigens, such as antibodies which bind to the EGFR, ErbB3, ErbB4, or vascular endothelial factor (VEGF). Alternatively, or in addition, two or more anti-ErbB2 antibodies may be co-administered to the patient. Sometimes, it may be beneficial to also administer one or more cytokines to the patient. In a preferred embodiment, the ErbB2 antibody is co-administered with a growth inhibitory agent. For example, the growth inhibitory agent may be administered first, followed by the ErbB2 antibody. However, simultaneous administration or administration of the ErbB2 antibody first is also contemplated. Suitable dosages for the growth inhibitory agent are those presently used and may be lowered due to the combined action (synergy) of the growth inhibitory agent and anti-ErbB2 antibody.

For the prevention or treatment of disease, the appropriate dosage of antibody will depend on the type of disease to be treated, as defined above, the severity and course of the disease, whether the antibody is administered for preventive or therapeutic purposes, previous therapy, the patient's clinical history and response to the antibody, and the discretion of the attending physician. The antibody is suitably administered to the patient at one time or over a series of treatments.

Depending on the type and severity of the disease, about 1 µg/kg to 15 mg/kg (e.g. 0.1-20 mg/kg) of antibody is an initial candidate dosage for administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. A typical daily dosage might range from about 1 µg/kg to 100 mg/kg or more, depending on the factors mentioned above. For repeated administrations over several days or longer, depending on the condition, the treatment is sustained until a desired suppression of disease symptoms occurs. However, other dosage regimens may be useful. The progress of this therapy is easily monitored by conventional techniques and assays.

Further information about suitable dosages is provided in the Example below.

#### V. Articles of Manufacture

In another embodiment of the invention, an article of manufacture containing materials useful for the treatment of the disorders described above is provided. The article of manufacture comprises a container, a label and a package insert. Suitable containers include, for example, bottles, vials, syringes, etc. The containers may be formed from a variety of materials such as glass or plastic. The container holds a composition which is effective for treating the condition and may

26

have a sterile access port (for example the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection needle). At least one active agent in the composition is an anti-ErbB2 antibody. The label on, or associated with, the container indicates that the composition is used for treating the condition of choice. The article of manufacture may further comprise a second container comprising a pharmaceutically-acceptable buffer, such as phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, and syringes. In addition, the article of manufacture comprises a package insert with instructions for use, including a warning that the composition is not to be used in combination with anthracycline-type chemotherapeutic agent, e.g. doxorubicin, or epirubicin.

#### DEPOSIT OF MATERIALS

The following hybridoma cell lines have been deposited with the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Md., USA (ATCC):

| Antibody Designation | ATCC No.       | Deposit Date  |
|----------------------|----------------|---------------|
| 7C2                  | ATCC HB-12215  | Oct. 17, 1996 |
| 7F3                  | ATCC HB-12216  | Oct. 17, 1996 |
| 4D5                  | ATCC CRL 10463 | May 24, 1990  |

Further details of the invention are illustrated by the following non-limiting Example.

#### EXAMPLE

##### Materials and Methods

**Anti-ErbB2 monoclonal antibody** The anti-ErbB2 IgG<sub>1</sub>K murine monoclonal antibody 4D5, specific for the extracellular domain of ErbB2, was produced as described in Fendly et al., *Cancer Research* 50:1550-1558 (1990) and WO89/06692. Briefly, NIH 3T3HER2-3<sub>400</sub> cells (expressing approximately 1×10<sup>5</sup> ErbB2 molecules/cell) produced as described in Hudziak et al. *Proc. Natl. Acad. Sci. (USA)* 84:7159 (1987) were harvested with phosphate buffered saline (PBS) containing 25 mM EDTA and used to immunize BALB/c mice. The mice were given injections i.p. of 10<sup>7</sup> cells in 0.5 ml PBS on weeks, 0, 2, 5 and 7. The mice with antisera that immunoprecipitated <sup>32</sup>P-labeled ErbB2 were given i.p. injections of a wheat germ agglutinin-Sepharose (WGA) purified ErbB2 membrane extract on weeks 9 and 13. This was followed by an i.v. injection of 0.1 ml of the ErbB2 preparation and the splenocytes were fused with mouse myeloma line X63-Ag8.653. Hybridoma supernatants were screened for ErbB2-binding by ELISA and radioimmuno-precipitation. MOPC-21 (IgG1), (Cappell, Durham, N.C.), was used as an isotype-matched control.

The treatment was performed with a humanized version of the murine 4D5 antibody (HERCEPTIN®). The humanized antibody was engineered by inserting the complementarity determining regions of the murine 4D5 antibody into the framework of a consensus human immunoglobulin IgG<sub>1</sub> (IgG<sub>1</sub>) (Carter et al., *Proc. Natl. Acad. Sci. USA* 89:4285-4289 [1992]). The resulting humanized anti-ErbB2 monoclonal antibody has high affinity for p185<sup>HER2</sup> (Dilohiation constant [K<sub>d</sub>]=0.1 nmol/L), markedly inhibits, in vitro and in human xenografts, the growth of breast cancer cells that con-



US 7,892,549 B2

27

tain high levels of p185<sup>HER2</sup>, induces antibody-dependent cellular cytotoxicity (ADCC), and has been found clinically active, as a single agent, in patients with ErbB2-overexpressing metastatic breast cancers that had received extensive prior therapy. HERCEPTIN® is produced by a genetically engineered Chinese Hamster Ovary (CHO) cell line, grown in large scale, that secretes the antibody into the culture medium. The antibody is purified from the CHO culture media using standard chromatographic and filtration methods. Each lot of antibody used in this study was assayed to verify identity, purity, and potency, as well as to meet Food and Drug Administration requirements for sterility and safety.

Eligibility Criteria Patients had to fulfill all of the following criteria to be eligible for study admission:

Metastatic breast cancer

Overexpression of the ErbB2 (HER2) oncogene (2+ to 3+ as determined by immunohistochemistry or fluorescence in situ hybridization (FISH). [Tumor expression of ErbB2 can be determined by immunohistochemical analysis, as previously described (Slamon et al., [1987] and [1989], supra), of a set of thin sections prepared from the patient's paraffin-archived tumor blocks. The primary detecting antibody used is murine 4D5 MAb, which has the same CDRs as the humanized antibody used for the treatment. Tumors are considered to over-express ErbB2 if at least 25% of tumor cells exhibit characteristic membrane staining for p185<sup>HER2</sup>].

Bidimensionally measurable disease (including lytic bone lesions) by radiographic means, physical examination, or photographs

Measurable disease was defined as any mass reproducibly measurable in two perpendicular diameters by physical examination, X-ray (plain films), computerized tomography (CT), magnetic resonance imaging (MRI), ultrasound, or photographs.

Osteoblastic metastases, pleural effusions, or ascites were not considered to be measurable. Measurable lesions must be at least 1 cm in greatest dimension. Enumeration of evaluable sites of metastatic disease and number of lesions in an evaluable site (e.g. lung) had to be recorded on the appropriate Case Report Form (CRF). If a large number of pulmonary or hepatic lesions were present, the six largest lesions per site were followed.

The ability to understand and willingness to sign a written informed consent form

Women ≥ 18 years

Suitable candidates for receiving concomitant cytotoxic chemotherapy as evidenced by screening laboratory assessments of hematologic, renal, hepatic, and metabolic functions.

Exclusion Criteria Patients with any of the following were excluded from study entry:

Prior cytotoxic chemotherapy for metastatic breast cancer  
Patients may have received prior hormonal therapy (e.g. tamoxifen) for metastatic disease or cytotoxic therapy in the adjuvant setting.

Concomitant malignancy that has not been curatively treated

A performance status of <60% on the Karnofsky scale

Pregnant or nursing women; women of childbearing potential, unless using effective contraception as determined by the investigator

Bilateral breast cancer (either both primary tumors must have 2+ to 3+HER2 overexpression, or the metastatic site must have 2+ to 3+HER2 overexpression)

28

Use of investigational or unlicensed agents within 30 days prior to study entry

Clinically unstable or untreated metastases to the brain (e.g. requiring radiation therapy)

Based upon the foregoing criteria, 469 patients were chosen, and enrolled in the study. Half the patients (stratified by chemotherapy) were randomized to additionally receive the HERCEPTIN® antibody (see below).

Administration and Dosage

Anti-ErbB2 Antibody

On day 0, a 4 mg/kg dose of humanized anti-ErbB2 antibody (HERCEPTIN®, H) was administered intravenously, over a 90-minute period. Beginning on day 7, patients received weekly administration of 2 mg/kg antibody (i.v.) over a 90-minute period.

Chemotherapy

The patients received one of two chemotherapy regimens for a minimum of six cycles, provided their disease was not progressing: a) cyclophosphamide and doxorubicin or epirubicin (AC), if patients have not received anthracycline therapy in the adjuvant setting, or b) paclitaxel (T, TAXOL®), if patients have received any anthracycline therapy in the adjuvant setting. The initial dose of the HERCEPTIN® antibody preceded the first cycle of either chemotherapy regimen by 24 hours. Subsequent doses of the antibody were given immediately before chemotherapy administration, if the initial dose of the antibody was well tolerated. If the first dose of the antibody was not well tolerated, subsequent infusions continued to precede chemotherapy administration by 24 hours. Patients were permitted to continue receiving chemotherapy beyond six cycles if, in the opinion of the treating physician, they were continuing to receive treatment benefit.

Cyclophosphamide (600 mg/m<sup>2</sup>) was given either by iv push over a minimum period of 3 minutes or by infusion over a maximum period of 2 hours.

Doxorubicin (60 mg/m<sup>2</sup>) or epirubicin (75 mg/m<sup>2</sup>) were given either by slow iv push over a minimum period of 3-5 minutes or by infusion over a maximum period of 2 hours, according to institutional protocol.

Paclitaxel (TAXOL®) was given at a dose of 175 mg/m<sup>2</sup> over 3 hours by intravenous administration. All patients receiving paclitaxel were premedicated with dexamethasone (or its equivalent) 20 mg×2, administered orally 12 and 6 hours prior to paclitaxel; diphenhydramine (or its equivalent) 50 mg, iv, administered 30 minutes prior to paclitaxel, and dimetidine (or another H2 blocker) 300 mg, iv, administered 30 minutes prior to paclitaxel.

Response Criteria

Progressive Disease Objective evidence of an increase of 25% or more in any measurable lesion. Progressive disease also includes those instances when new lesions have appeared. For bone lesions, progression is defined as a 25% increase in objective measurement by plain film, CT, MRI; symptomatic new lesions not due to fracture; or requirement for palliative radiotherapy.

Complete Response Disappearance of all radiographically and/or visually apparent tumor for a minimum of 4 weeks. Skin and chest wall complete responses had to be confirmed by biopsy.

Partial Response A reduction of at least 50% in the sum of the products of the perpendicular diameters of all measurable lesions for a minimum period of 4 weeks. No new lesions may have appeared, nor may any lesions have progressed in size.

Minor Response A reduction of 25% to 49% in the sum of the products of the perpendicular diameters of all measurable lesions. No new lesions may have appeared, nor may any lesions have progressed in size.

US 7,892,549 B2

29

Stable Disease No change of greater than 25% in the size of measurable lesions. No lesions may have appeared.

Time to disease progression (TTP) was calculated from the beginning of therapy to progression. Confidence limits for response rates were calculated using the exact method for a single proportion. (Fleiss, J L, *Statistical Methods for Rates and Proportions* (ed.2), New York, N.Y., Wiley, 1981, pp 13-17).

## RESULTS

At a median follow-up of 10.5 months, assessments of time to disease progression (TTP in months) and response rates (RR) showed a significant augmentation of the chemotherapeutic effect by HERCEPTIN®, without increase in overall severe adverse events (AE):

|         | Enrolled | TTP(months) | RR(%)   | AE(%) |
|---------|----------|-------------|---------|-------|
| CRx     | 234      | 5.5         | 36.2    | 66    |
| CRx + H | 235      | 8.6*        | 62.00** | 69    |
| AC      | 145      | 6.5         | 42.1    | 71    |
| AC + H  | 146      | 9.0         | 64.9    | 68    |

30

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|       | Enrolled | TTP(months) | RR(%) | AE(%) |
|-------|----------|-------------|-------|-------|
| T     | 89       | 4.2         | 25.0  | 59    |
| T + H | 89       | 7.1         | 57.3  | 70    |

\*p &lt; 0.001 by log-rank test

\*\*p < 0.001 by X<sup>2</sup> test

CRx: chemotherapy

10 AC: anthracycline/cyclophosphamide treatment

H: HERCEPTIN ®

T: TAXOL ®

A syndrome of myocardial dysfunction similar to that observed with anthracyclines was reported more commonly with a combined treatment of AC+H (18% Grade 3/4) than with AC alone (3%), T (0%), or T+H (2%).

These data indicate that the combination of anti-ErbB2 antibody treatment with chemotherapy markedly increases the clinical benefit, as assessed by response rates and the evaluation of disease progression. However, due to the increased cardiac side-effects of doxorubicin or epirubicin, the combined use of anthracyclines with anti-ErbB2 antibody therapy is contraindicated. The results, taking into account risk and benefit, favor the combined treatment with HERCEPTIN® and paclitaxel (TAXOL®).

The disclosures of all citations in the specification are expressly incorporated herein by reference.

## SEQUENCE LISTING

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Val Gln Gly Asn Leu Glu Leu Thr Tyr Leu Pro Thr Asn Ala Ser  
35 40 45

Leu Ser Phe Leu Gln Asp Ile Gln Glu Val Gln Gly Tyr Val Leu  
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Ile Ala His Asn Gln Val Arg Gln Val Pro Leu Gln Arg Leu Arg  
65 70 75

Ile Val Arg Gly Thr Gln Leu Phe Glu Asp Asn Tyr Ala Leu Ala  
80 85 90

Val Leu Asp Asn Gly Asp Pro Leu Asn Asn Thr Thr Pro Val Thr  
95 100 105

Gly Ala Ser Pro Gly Gly Leu Arg Glu Leu Gln Leu Arg Ser Leu  
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Thr Glu Ile Leu Lys Gly Gly Val Leu Ile Gln Arg Asn Pro Gln  
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Leu Cys Tyr Gln Asp Thr Ile Leu Trp Lys Asp Ile Phe His Lys  
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Asn Asn Gln Leu Ala Leu Thr Leu Ile Asp Thr Asn Arg Ser Arg  
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US 7,892,549 B2

31

32

-continued

Ala  
166

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| Ser | Thr | Gln | Val | Cys | Thr | Gly | Thr | Asp | Met | Lys | Leu | Arg | Leu | Pro |
| 1   |     |     |     | 5   |     |     |     |     | 10  |     |     |     |     | 15  |

|     |     |     |     |     |     |     |     |     |     |     |     |     |     |     |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Ala | Ser | Pro | Glu | Thr | His | Leu | Asp | Met | Leu | Arg | His | Leu | Tyr | Gln |
|     |     |     |     | 20  |     |     |     |     | 25  |     |     |     |     | 30  |

|     |     |
|-----|-----|
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|     | 32  |

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US 7,892,549 B2

33

34

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Lys Asp Pro Pro Phe Cys Val Ala Arg Cys Pro Ser Gly Val Lys
 35            40            45
Pro Asp Leu Ser Tyr Met Pro Ile Trp Lys Phe Pro Asp Glu Glu
 50            55            60
Gly Ala Cys Gln Pro
 65

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The invention claimed is:

1. A method for the treatment of a human patient with breast cancer that overexpresses ErbB2 receptor, comprising administering a combination of an antibody that binds ErbB2, a taxoid, and a further growth inhibitory agent to the human patient in an amount effective to extend the time to disease progression in the human patient, wherein the antibody binds to epitope 4D5 within the ErbB2 extracellular domain sequence.
2. The method of claim 1 wherein the antibody is a humanized 4D5 anti-ErbB2 antibody.
3. The method of claim 1 wherein the antibody cross-blocks binding of 4D5 to the ErbB2 extracellular domain sequence.
4. The method of claim 1 wherein the antibody binds to amino acid residues in the region from about residue 529 to about residue 625 of the ErbB2 extracellular domain sequence.
5. A method for the treatment of a human patient with breast cancer characterized by overexpression of ErbB2 receptor, comprising administering an effective amount of a combination of an anti-ErbB2 antibody which binds epitope 4D5 within the ErbB2 extracellular domain sequence, a taxoid, and a further therapeutic agent, to the human patient.
6. The method of claim 5 wherein the breast cancer is metastatic breast carcinoma.
7. The method of claim 5 wherein the antibody is a humanized 4D5 anti-ErbB2 antibody.
8. The method of claim 7 wherein the antibody is administered as a 4 mg/kg dose and then weekly administration of 2 mg/kg.
9. The method of claim 5 wherein the taxoid is paclitaxel.
10. The method of claim 5 wherein efficacy is measured by determining the time to disease progression or the response rate.
11. The method of claim 5, wherein the further therapeutic agent is selected from the group consisting of: another ErbB2 antibody, EGFR antibody, ErbB3 antibody, ErbB4 antibody, vascular endothelial growth factor (VEGF) antibody, cytokine, and growth inhibitory agent.
12. The method of claim 5 wherein the further therapeutic agent is another ErbB2 antibody.
13. The method of claim 5 wherein the further therapeutic agent is a vascular endothelial growth factor (VEGF) antibody.
14. The method of claim 5 wherein the further therapeutic agent is a growth inhibitory agent.
15. The method of claim 14 wherein the growth inhibitory agent is a DNA alkylating agent.
16. A method for the treatment of a human patient with ErbB2 overexpressing breast cancer, comprising administering a combination of an antibody that binds epitope 4D5 within the ErbB2 extracellular domain sequence, a taxoid and a further growth inhibitory agent, in the absence of an anthracycline derivative, to the human patient in an amount effective to extend the time to disease progression in the human patient.
17. The method of claim 16 wherein the breast cancer is metastatic breast carcinoma.

\* \* \* \* \*

### **CERTIFICATE OF SERVICE**

I hereby certify that, on this 9th day of July, 2019 I filed the foregoing Non-Confidential Brief for Appellant Genentech, Inc. with the Clerk of the United States Court of Appeals for the Federal Circuit via the CM/ECF system, which will send notice of such filing to all registered CM/ECF users.

/s/ Robert J. Gunther, Jr.

ROBERT J. GUNTHER, JR.

WILMER CUTLER PICKERING

HALE AND DORR LLP

7 World Trade Center

250 Greenwich Street

New York, NY 10007

(212) 230-8800



## **CERTIFICATE OF COMPLIANCE**

Pursuant to Fed. R. App. P. 32(g), the undersigned hereby certifies that this brief complies with the type-volume limitation of Federal Circuit Rule 32(a).

1. Exclusive of the exempted portions of the brief, as provided in Fed. R. App. P. 32(f) and Fed. Cir. R. 32(b), the brief contains 7,298 words.

2. The brief has been prepared in proportionally spaced typeface using Microsoft Word 2016 in 14-point Times New Roman font. As permitted by Fed. R. App. P. 32(g), the undersigned has relied upon the word count feature of this word processing system in preparing this certificate.

/s/ Robert J. Gunther, Jr.

ROBERT J. GUNTHER, JR.

WILMER CUTLER PICKERING

HALE AND DORR LLP

7 World Trade Center

250 Greenwich Street

New York, NY 10007

(212) 230-8800

July 9, 2019