

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

PFIZER, INC.,
Petitioner,

v.

CHUGAI PHARMACEUTICAL CO. LTD.,
Patent Owner.

Case IPR2017-01357
Patent 7,332,289 B2

Before GRACE KARAFFA OBERMANN, RAMA G. ELLURU, and
JACQUELINE T. HARLOW, *Administrative Patent Judges*.

HARLOW, *Administrative Patent Judge*.

FINAL WRITEN DECISION
35 U.S.C. § 318(a) and 37 C.F.R. § 42.73

I. INTRODUCTION

Pfizer, Inc. (“Petitioner”) filed a Petition requesting an *inter partes* review of claims 1–8 and 13 of U.S. Patent No. 7,332,289 B2 (Ex. 1001, “the ’289 patent”). Paper 2 (“Pet.”). Chugai Pharmaceutical Co. Ltd. (“Patent Owner”) filed a Preliminary Response. Paper 6 (“Prelim. Resp.”). On December 1, 2017, we instituted an *inter partes* review of all challenged claims on all grounds asserted. Paper 7 (“Institution Decision” or “Inst. Dec.”). Following our institution, Patent Owner filed a Response to the Petition (Paper 19, “PO Resp.”), and Petitioner filed a Reply to Patent Owner’s Response (Paper 28, “Reply”).

Petitioner also filed a Motion to Exclude Evidence (Paper 36), to which Patent Owner filed an Opposition (Paper 44), and Petitioner filed a Reply (Paper 47). In addition, Patent Owner filed an Authorized Statement Regarding Petitioner’s Reply Papers Beyond the Proper Scope and Improper Incorporation By Reference (Paper 40), to which Petitioner filed a Response (Paper 42). Patent Owner also filed a Motion for Observations on Cross Examination (Paper 41), to which Petitioner filed a Response (Paper 43).

An oral hearing was held on August 2, 2018. The transcript of that hearing has been entered into the record. Paper 55 (“Tr.”).

We issue this Final Written Decision pursuant to 35 U.S.C. § 318(a) and 37 C.F.R. § 42.73. Having considered the record before us, we determine that Petitioner has not shown by a preponderance of the evidence that claims 1–8 and 13 of the ’289 patent are unpatentable. *See* 35 U.S.C. § 316(e).

A. Related Matters

The parties inform us of no related pending litigations. Pet. 3–4; Paper 5, 2–3. In addition to the instant proceeding, Petitioner has challenged, and we have instituted *inter partes* review of, U.S. Patent No. 7,927,815 B2, which is related to the '289 patent. *See Pfizer, Inc. v. Chugai Pharma. Co. Ltd*, Case IPR2017-01358 (PTAB Dec. 1, 2017) (Paper 7). We issue our decision determining that Petitioner has not shown the unpatentability of the challenged claims of the '815 patent concurrently with this Decision.

B. The '289 Patent

The '289 patent, titled “Method of Purifying Protein,” issued February 19, 2008, from U.S. Patent Application No. 10/471,374, which is the U.S. National Stage Application of International Application No. PCT/JP02/02248, filed on March 11, 2002. Ex. 1001, at [54], [45], [21], [22], [86]. The '289 patent claims priority to Japanese Patent Application No. 2001-067111, filed on March 9, 2001. *Id.* at [30].

The '289 patent describes a “method for purifying proteins, more specifically [] a method for removing contaminant DNA from a sample containing a physiologically active protein such as antibody molecules.” Ex. 1001, 1:5–8. The '289 patent recognizes that methods for removing contaminant DNA from recombinant antibody drug formulations were known in the art. *See, e.g., id.* at 1:35–43. The '289 patent states, however, that the chromatographic processes associated with known purification

methods were “time-, labor- and cost-consuming, as well as being complicated. Moreover, they fail to provide stable results.” *Id.* at 1:44–47.

To address these shortcomings, the ’289 patent discloses the “surprising finding that contaminant DNA can be efficiently removed from a sample containing a physiologically active protein without using complicated chromatographic processes.” *Id.* at 1:59–62. In particular, the ’289 patent teaches that such a sample can be “converted into an acidic aqueous solution of low conductivity, neutralized by addition of a buffer to raise the pH to a neutral level, and then filtered through a filter to remove the resulting particles.” *Id.* at 1:59–66. The ’289 patent goes on to state that “[w]ithout being bound by any particular theory, the inventors of the present invention estimate that each of the [aforementioned] particles is a conjugate formed between physiologically active protein and DNA.” *Id.* at 6:16–19.

The ’289 patent explains that “[a]s used herein, ‘an acidic aqueous solution of low conductivity’ generally refers to an aqueous solution of pH 1.5 to pH 3.9, . . . which has a molarity of 0 to 100 mM, . . . or has an ionic strength of 0 to 0.2, . . . or has a conductivity of 0 to 300 mS/m” *Id.* at 5:29–35. The ’289 patent further discloses that “[t]he acidic aqueous solution may be selected from aqueous solutions of hydrochloric acid, citric acid, acetic acid and other acids.” *Id.* at 5:35–37. The ’289 patent also states that “[t]he type, conductivity and pH of acidic aqueous solution of low conductivity will vary depending on the type of physiologically active protein or antibody to be purified. Those skilled in the art will readily determine optimal conditions for these parameters in preliminary experiments as described herein.” *Id.* at 5:37–42.

With regard to the neutralization and particle removal steps of the above-described purification procedure, the '289 patent teaches that neutralization of the solution containing a physiologically active protein to a "neutral pH level" "in turn, produces particles (i.e., becomes clouded). These particles may be removed by filtration through a filter to ensure efficient removal of contaminant DNA." *Id.* at 6:4–8. The '289 patent exemplifies a "1.0–0.2 μm Cellulose Acetate Filter System (Corning) or TFF" as filters available for particle filtration. *Id.* at 6:10–15.

C. Illustrative Claim

Claim 1, reproduced below, is the sole independent claim in the '289 patent, and is illustrative of the claimed subject matter.

1. A method for removing contaminant DNA in an antibody-containing sample, which comprises the followings [sic] steps:

- 1) applying the antibody-containing sample to affinity chromatography on Protein A or Protein G;
- 2) eluting the antibody with an acidic aqueous solution of low conductivity having a molarity of 100 mM or less;
- 3) neutralizing the eluate from step (2) to form particles by addition of a buffer to raise the pH to 4 to 8, wherein the molarity of the neutralized eluate is 100 mM or less; and
- 4) removing the particles to thereby remove contaminant DNA from the antibody-containing sample.

Ex. 1001, 12:45–58.

D. Evidence Relied Upon

Petitioner relies upon the following as a prior art reference (Pet. 5):

Shadle WO 95/22389 Aug. 24, 1995 (Ex. 1003)

Petitioner also relies on declarations submitted by Todd M. Przybycien, Ph.D. (Ex. 1002 (Dr. Przybycien’s Opening Declaration); Ex. 1036 (Dr. Przybycien’s Reply Declaration)).

Patent Owner relies on the Declarations of Steven M. Cramer, Ph.D. (Ex. 2015), Kirston Koths, Ph.D. (Ex. 2016), and Harry G. Brittain, Ph.D. (Ex. 2019).

E. Instituted Challenges

We instituted trial based on each challenge to the patentability of the ’289 patent presented in the Petition (Pet. 5):

Claim(s)	Basis	References
1–8 and 13	§ 102(b)	Shadle
1–8 and 13	§ 103(a)	Shadle

II. ANALYSIS

A. Level of Ordinary Skill in the Art

The level of skill in the art is a factual determination that provides a primary guarantee of objectivity in an obviousness analysis. *Al-Site Corp. v. VSI Int’l Inc.*, 174 F.3d 1308, 1324 (Fed. Cir. 1999) (citing *Graham v. John Deere Co.*, 383 U.S. 1, 17–18 (1966); *Ryko Mfg. Co. v. Nu-Star, Inc.*, 950 F.2d 714, 718 (Fed. Cir. 1991)).

According to Petitioner, a person of ordinary skill in the art at the time of the invention would have had “at least a graduate degree, such as a Ph.D.,

and several years of postgraduate training or practical experience in a relevant discipline such as biochemistry, process chemistry, protein chemistry, chemical engineering and/or biochemical engineering, among others.” Pet. 6 (citing Ex. 1002 ¶¶ 28–29). Petitioner further contends that “[s]uch a person would also understand that protein purification is a multidisciplinary field, and could take advantage of the specialized skills of others using a collaborative approach.” *Id.* (citing Ex. 1002 ¶¶ 28–29). Patent Owner does not address Petitioner’s position on this matter and does not propose its own description for a person of ordinary skill in the art at the time of the invention.

We agree with Petitioner, and adopt Petitioner’s description of the level of ordinary skill in the art at the time of invention of the ’289 patent. We also note that the applied prior art reflects a level of skill at the time of the claimed invention consistent with our determination. *See Okajima v. Bourdeau*, 261 F.3d 1350, 1355 (Fed. Cir. 2001).

In addition, we have reviewed the credentials of Petitioner’s and Patent Owner’s declarants, and recognize each of them as qualified to provide the proffered opinions on the level of skill and the knowledge of a person of ordinary skill in the art at the time of the invention. *See* Ex. 1004, Attachment A; Ex. 2015, Appendix A; Ex. 2016, Appendix A; Ex. 2019, Appendix A. The relative weight that we assign such testimony, however, is subject to additional factors. *See, e.g.*, 37 C.F.R. § 42.65(a) (“Expert testimony that does not disclose the underlying facts or data on which the opinion is based is entitled to little or no weight.”); Office Patent Trial Practice Guide, 77 Fed. Reg. 48,756, 48,763 (Aug. 14, 2012) (same).

B. Claim Construction

The broadest reasonable interpretation standard applies to the construction of the challenged claims in this proceeding. 37 C.F.R. § 42.100(b) (2016); *Cuozzo Speed Techs., LLC v. Lee*, 136 S.Ct. 2131, 2142 (2016). Under that standard, and absent any special definitions, we give claim terms their ordinary and customary meaning, as would be understood by one of ordinary skill in the art at the time of the invention, in the context of the entire disclosure. *In re Translogic Tech., Inc.*, 504 F.3d 1249, 1257 (Fed. Cir. 2007). Any special definitions for claim terms must be set forth with reasonable clarity, deliberateness, and precision. *In re Paulsen*, 30 F.3d 1475, 1480 (Fed. Cir. 1994).

1. “molarity”

In the Institution Decision, we concluded that the broadest reasonable interpretation of the claim term “molarity,” as it is used in the ’289 patent, is the “total concentration of solute present in the solution.” Inst. Dec. 12.

Patent Owner agrees that “molarity” refers to the total concentration of solute in a solution, and avers that this construction is supported by the claims, specification, and file history of the ’289 patent, which address the “contributions of *multiple* solutes to the solution’s molarity.” PO Resp. 16. Patent Owner further asserts that the molarity calculations for Shadle’s neutralized eluate provided by Petitioner’s declarant, Dr. Przybycien, which account for contributions from different solutes (Ex. 1002 ¶¶ 79–82) are inconsistent with Petitioner’s contention that molarity refers to the concentration of a single solute in a solution. PO Resp. 17.

Petitioner does not directly address the construction of “molarity” in its Reply. However, during oral argument, Petitioner explained that it maintains its position that “molarity” refers to the concentration of a single solute in solution. Tr. 13:1–7. In the Petition, Petitioner asserts that “molarity” refers to “[a] measure of the concentration of a given solute within a solution in terms of the moles of that solute contained per liter of solution.” Pet. 24. As we observed in the Institution Decision, however, despite its proposed construction, Petitioner nevertheless appears to recognize that

“molarity” may take account of multiple solutes present in a solution. Specifically, Petitioner proposes that the term “molarity,” as it is used in the greater claim phrase “an acidic aqueous solution of low conductivity having a molarity of 100 mM or less,” should be understood to mean “that the molarity of the acidic aqueous solution is 100 mM or less, without considering any effects of the contaminant DNA or protein from the sample.”

Inst. Dec. 8 (quoting Pet. 24–25).

Consistent with the Institution Decision, we determine that “the plain language of the claims, as well as the specification of the ’289 patent, indicates that the term ‘molarity’ refers to the total concentration of solute present in the solution, rather than the concentration of one particular solute.” *Id.* at 10. The claims of the ’289 patent refer consistently to the overall molarity of solutions, and not of any particular solute in a given solution. For example, claim 1 requires “an *acidic aqueous solution* of low conductivity having a molarity of 100 mM or less” (Ex. 1001, 12:50–51 (emphasis added)), and further recites that the “molarity of the *neutralized*

eluate is 100 mM or less” (*id.* at 12:54 (emphasis added)). Claim 2 similarly limits the molarity of the “acidic aqueous solution.” *Id.* at 12:59–61.

Notably, these claims do not identify any particular solute to which the term molarity refers; rather, the claims of the ’289 patent describe solutions having certain characteristics, of which solution molarity is one. As we stated in our Institution Decision:

Similarly, the specification of the ’289 patent refers to the molarity of the complete solution, rather than one solute in that solution. *See, e.g., id.* at 4:61–66 (“As used herein, a ‘neutral aqueous solution . . .’ generally refers to an-aqueous [sic] solution . . . which has a molarity of 0 to 100mM”), 5:28–31 (“an ‘acidic aqueous solution of low conductivity’ generally refers to an aqueous solution . . . , which has a molarity of 0 to 100 mM”). The prosecution history of the ’289 patent likewise references the molarity of the solution, rather than of a given solute in the solution. *See e.g., Ex.1005*, 12 (“an important feature of the present invention is to adjust pH value of the solution, the eluate, to from 4 to 8 while maintaining the molarity of the solution at 100mM or less.”), 37 (“0.1 M buffer was used as an eluent, and 1 M Tris-HCl was used to adjust the pH of the eluted fraction, that is, the fact that 0.1 M and 1 M solutions were used means that the molarity of the eluted fration [sic] must be over 0.1 M (100 mM)”).

Inst. Dec. 10–11.

In the Institution Decision, because the parties did not identify any controversy concerning whether any effects of contaminant DNA or protein from the sample should be taken into account when calculating the molarity of the acidic aqueous solution, we declined to decide that issue. Inst. Dec. 11–12 (citing *Nidex Motor Corp. v. Zhongshan Broad Ocean Motor Co. Ltd.*, 868 F.3d 1013, 1017 (Fed. Cir. 2017) (“we need only construe

terms ‘that are in controversy, and only to the extent necessary to resolve the controversy’” (quoting *Vivid Techs., Inc. v. Am. Sci. & Eng’g, Inc.*, 200 F.3d 795, 803 (Fed. Cir. 1999))). Nor is resolution of that issue necessary at this juncture. In particular, we observe that our anticipation and obviousness analyses below remain the same regardless of whether any effects from contaminant DNA and/or protein from the sample are included in the molarity calculation.

2. *Other Claim Terms*

In the Institution Decision, we concluded that the claim phrase “an acidic aqueous solution of low conductivity having a molarity of 100 mM or less” encompasses an aqueous solution of pH 1.5 to pH 3.9, which has a molarity of 0 to 100 mM. Inst. Dec. 12. Neither Petitioner nor Patent Owner challenges this construction. *See* Pet. 30; PO Resp. 12–23. Accordingly, for the reasons set forth in the Institution Decision, we determine that “an acidic aqueous solution of low conductivity having a molarity of 100 mM or less” encompasses an aqueous solution of pH 1.5 to pH 3.9, which has a molarity of 0 to 100 mM.

In its Response, Patent Owner proposes constructions for several additional terms. Namely, Patent Owner contends that the preamble of claim 1 should be construed as limiting (PO Resp. 12), “to form particles” should be construed to require that the solution becomes clouded (*id.* at 17), and “the treated sample containing an antibody,” as recited in claim 5, should be interpreted to mean “the sample resulting from performing the method of claim 1, which concludes with removing particles in step 4” (*id.*

at 18). Because interpretation of these claim terms is not necessary to our anticipation or obviousness analyses, we need not construe them. *See Nidec*, 868 F.3d at 1017.

*C. Prior Art Relied Upon:
Overview of Shadle*

Petitioner relies on the teachings of Shadle (Ex. 1003) as the basis for its patentability challenges in this proceeding.

Shadle discloses methods for the “purification of antibody molecule proteins” that employ “sequential steps of Protein A affinity chromatography, ion exchange chromatography, and hydrophobic interaction chromatography.” Ex. 1003, Abstract. In this regard, Shadle teaches that a “purification protocol should not only provide a protein product that is essentially free of other proteins, . . . but also eliminate or reduce to acceptable levels other host cell contaminants, DNA, RNA, potential pyrogens and the like.” *Id.* at 9:12–16. In particular, Shadle discloses:

The purified antibodies obtained by practicing the process of this invention have the following properties: 1) greater than 97% antibody protein by weight; 2) stable to proteolytic degradation at 4°C for at least three months; 3) low (< 0.1 E.U./mg protein) endotoxin; 4) low (< 1 pg/mg protein) DNA; 5) non-antibody protein < 5% by weight; and 6) virally inactive.

Id. at 14:21–27.

Shadle exemplifies the disclosed protein purification method by describing a procedure “for the isolation and purification of a monoclonal antibody against Respiratory Syncytial Virus (RSV),” identified as

“RSHZ 19.” *Id.* at 15:3–7. Shadle explains that this “process is designed to prepare RSHZ-19 of >95% purity while removing contaminants derived from the host cell, cell culture medium, or other raw materials.” *Id.* at 15:7–9.

In Example IA, Shadle teaches the application of 100 liters of conditioned culture medium containing 0.8 grams per liter of RSHZ-19 monoclonal antibody to a previously equilibrated ProSep A affinity column. *Id.* at 21:4–8. Subsequent to washing with 15 liters of PBS/glycine, the “IgG was eluted by applying 15–20 liters of ProSep A elution buffer. Fractions of the non-bound peak and the elution peak were collected and assayed for IgG content using an HPLC assay. The eluate was approximately 15 liters in volume, and contained approximately 5 milligrams protein per milliliter.” *Id.* at 21:9–13. Shadle identifies the “ProSep Elution Buffer” as being composed of 25 mM citrate, and having pH 3.5. *Id.* at 20:10. Shadle additionally explains that “[t]he eluate fractions from the Protein A capture . . . are pooled based on the UV tracing on the chromatogram, and the entire peak is collected.” *Id.* at 19:3–5.

Shadle further discloses that

[i]mmediately after elution, the sample was adjusted to pH 3.5 by the addition of 2.5 M hydrochloric acid, held for approximately 30 minutes, and adjusted to pH 5.5 by the addition of approximately 350 milliliters of 1 M Tris base. After neutralizing to pH 5.5, the sample was filtered through a 0.1 micron Polygard CR filter in tandem with a sterile 0.2 micron Millipak 200, into a sterile container.

Id. at 21:15–19. Subsequently, the filtered sample was subject to cation exchange chromatography and hydrophobic interaction chromatography. *Id.* at 21:26–22:29.

D. Anticipation Based on Shadle

Petitioner asserts that claims 1–8 and 13 are anticipated under § 102(b) by Shadle. Pet. 26–43. Patent Owner disagrees that Shadle anticipates the challenged claims. PO Resp. 23–52.

To anticipate a claim, a prior art reference must disclose every limitation of the claimed invention, either expressly or inherently. *Blue Calypso, LLC v. Groupon, Inc.*, 815 F.3d 1331, 1341 (Fed. Cir. 2016). “To establish that a prior art reference inherently—rather than expressly—discloses a claim limitation, ‘the limitation at issue necessarily must be present, or [is] the natural result of the combination of elements explicitly disclosed by the prior art.’” *Endo Pharm. Sols., Inc. v. Custopharm Inc.*, 894 F.3d 1374, 1381 (Fed. Cir. 2018) (alteration in original) (quoting *PAR Pharm., Inc. v. TWI Pharm., Inc.*, 773 F.3d 1186, 1196 (Fed. Cir. 2014)).

Inherency, however, may not be established by probabilities or possibilities. The mere fact that a certain thing *may* result from a given set of circumstances is not sufficient. [Citations omitted.] If, however, the disclosure is sufficient to show that the natural result flowing from the operation as taught would result in the performance of the questioned function, it seems to be well settled that the disclosure should be regarded as sufficient.

Cont’l Can Co. USA, Inc. v. Monsanto Co., 948 F.2d 1264, 1269 (Fed. Cir. 1991) (alteration in original) (quoting *In re Oelrich*, 666 F.2d 578, 581 (CCPA 1981)).

The challenged claims of the '289 patent each require eluting antibody with an "acidic aqueous solution of low conductivity having a molarity of 100 mM or less," neutralizing the resulting antibody-containing eluate "to form particles by addition of a buffer to raise the pH to 4 to 8, wherein the molarity of the neutralized eluate is 100 mM or less," and "removing the particles to thereby remove contaminant DNA from the antibody-containing sample." Ex. 1001, 12:50–57. To support its contention that Shadle anticipates the challenged claims, Petitioner asserts, *inter alia*, that each of the initial eluent and the antibody-containing eluate disclosed by Shadle has a molarity of 100 mM or less, and that Shadle discloses neutralizing the antibody-containing eluate to form particles. Pet. 29–37. For the reasons set forth below, however, we determine that Petitioner has not met its burden to establish, by a preponderance of the evidence, that Shadle discloses each of these claim requirements.¹

The antibody purification protocol set forth in Example IA of Shadle discloses the use of ProSep A Elution Buffer to elute antibody from a ProSep A column. Ex. 1003, 21:9–10. Table 1 of Shadle, which discloses the formulations for relevant buffers, characterizes the "ProSep Elution

¹ Because we determine that Petitioner has not met its burden with respect to the aforementioned claim steps, we need not address whether the preamble of claim 1 is limiting, or whether Petitioner has established that it is disclosed by Shadle. Neither must we address whether Shadle discloses "removing the particles to thereby remove contaminant DNA from the antibody-containing sample" (Ex. 1001, 12:56–57).

Buffer” as “25 mM citrate, pH 3.5.” *Id.* at 20:10. Shadle does not further describe the composition of the ProSep A Elution Buffer.

Relying on the concentration of citrate alone present in Shadle’s elution buffer, Petitioner argues that the claim requirement for eluting antibody with an aqueous solution of 100 mM or less is satisfied. Applying Petitioner’s interpretation of “molarity” as referring to the “concentration of a *given solute* within a solution” (Pet. 24), the Petition asserts that Shadle’s disclosure of using 25 mM citrate, pH 3.5 buffer for antibody elution expressly satisfies the initial eluent molarity requirement of the challenged claims. Pet. 29; *see also* Ex. 1002 ¶¶ 71–73.

Petitioner does not, however, adduce evidence sufficient to show that Shadle expressly discloses that its ProSep A Elution Buffer has 100 mM or less *total solute* present in the solution, as required under our interpretation of “molarity,” initially set forth in the Institution Decision (Inst. Dec. 12). *See Reply, passim.* Shadle does not describe how its ProSep A Elution Buffer is prepared, or otherwise define the composition of that buffer, beyond specifying the concentration of citrate and pH. *See* Ex. 1003, 20–21 (identifying the conditions of “ProSep Elution Buffer” as “25 mM citrate, pH 3.5”). Standing alone, Shadle’s characterization of its elution buffer as including 25 mM *citrate* and having pH 3.5 is insufficient to establish that Shadle expressly discloses an elution buffer with 100 mM or less *total solute*. *See* Ex. 2014, 90:23–92:19 (testimony by Dr. Przybycien explaining his reliance on molarity calculations for four different citrate buffer preparations to determine the total molarity of Shadle’s elution buffer).

Indeed, neither Petitioner nor its declarant, Dr. Przybycien, contends that Shadle describes any particular method for preparing ProSep A Elution Buffer, or that Shadle specifies the total concentration of solute present in its elution buffer. *See* Reply 6 (“[S]tarting with any of four conventional buffer preparations, Shadle meets the claimed molarity limitation.”); Ex. 1026, 2 (“Thus, total molarity of [Shadle’s] 25 mM Citrate elution buffer is 25 mM, 30.73 mM, or 44.08 mM, depending on the method of preparation used.”). Moreover, to the extent Petitioner asserts that Shadle expressly discloses an elution buffer having 25 mM *total solute* (*see* Tr. 14:13–20), Dr. Przybycien’s testimony that an ordinarily skilled artisan “would have understood that there were four conventional, most common methods for preparing such a buffer” (Ex. 1036 ¶ 26), and that the molarities of the resulting solutions varied according to which method was used (*id.* at ¶ 37; Ex. 1026, 1–2), belies any such contention. *See also* Ex. 2014, 88:22–89:16 (testimony by Dr. Przybycien acknowledging that Shadle’s 25 mM citrate elution buffer could be prepared a fifth way, but characterizing that method of preparation as unconventional).

On Reply, Petitioner recasts its theory of the case, shifting its contention that Shadle *expressly* discloses “eluting the antibody with an acidic aqueous solution of low conductivity having a molarity of 100 mM or less” (Ex. 1001, 12:50–51) to an assertion that Shadle *inherently* discloses that claim step. *Compare* Pet. 29 (“WO ’389 [Shadle] explicitly discloses step 2 of the claimed purification process.”), *with* Reply 8 (“Dr. Przybycien recalculated molarity under the Board’s construction, and confirmed that, regardless of the construction, Shadle inherently meets the molarity

limitations and anticipates the claims.”). To support its new inherency argument, Petitioner relies on updated molarity calculations submitted by Dr. Przybycien after institution, but prior to Patent Owner’s filing of its Response. Reply 8; Ex. 1026. In describing his updated calculations, Dr. Przybycien explains that he calculated the molarity of four different elution buffers that satisfy Shadle’s conditions of 25 mM citrate and pH 3.5, and that were prepared according to the most common and conventional methods. Ex. 1026, 1–2; Ex. 1036 ¶ 26. The molarities of the four buffer formulations evaluated by Dr. Przybycien range from 25 mM to 44 mM total solute. Ex. 1026, 2. Dr. Przybycien testifies that “for each of the four proposed conventional ProSep A citrate buffer elution solutions . . . it is clear that even when considering the ‘total molarity’ of the Shadle ProSep A citrate buffer, it would still inherently be below 100 mM (and below 50 mM).” Ex. 1036 ¶ 37.

We do not find Petitioner’s inherency arguments persuasive. First, Petitioner’s belated change of course and argument on Reply that Shadle *inherently* discloses “eluting the antibody with an acidic aqueous solution of low conductivity having a molarity of 100 mM or less” (Ex. 1001, 12:50–51) is improper. “Unlike district court litigation—where parties have greater freedom to revise and develop their arguments over time and in response to newly discovered material—the expedited nature of IPRs bring with it an obligation for petitioners to make their case in their petition to institute.” *Intelligent Bio-Sys., Inc. v. Illumina Cambridge Ltd.*, 821 F.3d 1359, 1369 (Fed. Cir. 2016). In an *inter partes* review, the petitioner has the burden from the onset to show with particularity why the patent it challenges is

unpatentable. *Harmonic Inc. v. Avid Tech., Inc.*, 815 F.3d 1356, 1363 (Fed. Cir. 2016). Specifically, the petition must identify “with particularity . . . the grounds on which the challenge to each claim is based, and the evidence that supports the grounds for the challenge to each claim.” 35 U.S.C.

§ 312(a)(3). Thus, although “the introduction of new evidence in the course of the trial is to be expected in *inter partes* review trial proceedings,” *Genzyme Therapeutic Prods. LP v. Biomarin Pharm. Inc.*, 825 F.3d 1360, 1366 (Fed. Cir. 2016), the shifting of arguments is not, *Wasica Fin. GmbH v. Cont’l Auto. Sys., Inc.*, 853 F.3d 1272, 1286 (Fed. Cir. 2017). Petitioner’s inherency argument concerning the total molarity of Shadle’s elution buffer is an impermissible shift of its anticipation theory because “[r]ather than explaining how its original petition was correct,” *id.*, i.e., how Shadle’s elution buffer expressly satisfied the recited molarity requirement, Petitioner’s “subsequent arguments amount to an entirely new theory of [inherent anticipation] absent from the petition,” *id.* For this reason alone, Petitioner has not shown that the challenged claims are unpatentable as anticipated by Shadle.

Second, even if Petitioner’s inherency argument were timely, it is nevertheless insufficient to support a finding of anticipation. Petitioner has not shown that the total concentration of solute present in Shadle’s elution buffer is necessarily 100 mM or less, as required by the challenged claims. As explained above, Shadle does not disclose the total molarity of its elution buffer, or describe how that buffer is prepared, beyond stating that it includes 25 mM citrate and is of pH 3.5. Ex. 1003, 20–21. Petitioner does not argue either that there is only a *single method* for making Shadle’s

elution buffer, which always yields a buffer having 100 mM or less total solute, or that *every method* for producing Shadle’s buffer results in a buffer with 100 mM or less total solute. *See* Ex. 1036 ¶ 26 (“Based on Shadle’s disclosure of using a ProSep A elution buffer of 25 mM citrate, pH 3.5, a POSA would have understood that there were four conventional, most common methods for preparing such a buffer.”). Rather, Petitioner contends that “starting with any of four conventional buffer preparations, Shadle meets the claimed molarity limitation.” Reply 6; *see also* Ex. 1036 ¶ 37 (“I have prepared updated calculations of the ‘total molarity’ for each of the four proposed conventional ProSep A citrate buffer elution solutions Based on these calculations, it is clear that even when considering the ‘total molarity’ of the Shadle ProSep A citrate buffer, it would still inherently be below 100 mM (and below 50 mM).”). That four common and conventional ways of preparing Shadle’s buffer each satisfy the elution buffer molarity requirement of the challenged claims is insufficient to establish that Shadle’s elution buffer *necessarily* meets that requirement. The probability or possibility that an ordinarily skilled artisan would have used one of the four methods described by Dr. Przybycien to prepare Shadle’s elution buffer is insufficient to establish inherency. *See Cont’l Can.*, 948 at 1269 (“Inherency, however, may not be established by probabilities or possibilities. The mere fact that a certain thing *may* result from a given set of circumstances is not sufficient.” (alteration in original) (quoting *In re Oelrich*, 666 F.2d at 581)).

Indeed, Patent Owner's declarant, Dr. Cramer, identifies a fifth way to prepare Shadle's 25 mM citrate, pH 3.5 ProSep A elution buffer. PO Resp. 31; Ex. 2015 ¶ 59. Specifically, Dr. Cramer testifies that "Shadle's ProSep A elution buffer (25 mM citrate, pH 3.5) could very well have been prepared using 25 mM trisodium citrate and hydrochloric acid—a method well known to [artisans of ordinary skill] at the time of Shadle and at the time of the '289 [patent]." Ex. 2015 ¶ 59. Dr. Cramer further testifies that such an artisan would have considered a trisodium citrate and hydrochloric acid "buffer preparation as among the numerous readily-available and reasonable choices for Shadle in preparing its citrate buffer of Example 1A [sic]" (*id.*). *See also* Ex. 1036 ¶ 27 (testimony by Dr. Przybycien "agree[ing] that it is theoretically possible to make the 25 mM citrate, pH 3.5 ProSep A elution buffer using trisodium citrate and HCl," but disputing that would be the normal and usual way of making the buffer); Ex. 2014, 88:22–89:8 (testimony by Dr. Przybycien acknowledging 25 mM citrate buffer could be prepared using trisodium citrate and hydrochloric acid, but stating that is not a conventional manner of preparing the buffer). Dr. Cramer additionally testifies that an ordinarily skilled artisan "reading Shadle would not know how the 25 millimolar citric buffer was made and, therefore — therefore, could have made that buffer in a wide variety of ways and one of them is with a trisodium citrate." Ex. 1034, 103:21–104:5. According to Dr. Cramer, factors such as the reagents available in the lab, personal preferences, and level of experience in making buffers, rather than any disclosure or suggestion in Shadle, would have motivated an ordinarily

skilled artisan to select one buffer formulation over another. *Id.* at 105:3–107:23. For purposes of this Decision, we credit Dr. Cramer’s testimony.

Petitioner takes issue with Patent Owner’s contention that a fifth elution buffer could be used to practice Shadle’s antibody purification process, but Petitioner’s arguments are unpersuasive on this record. The purported disadvantages of making Shadle’s elution buffer with trisodium citrate and hydrochloric acid identified by Petitioner (*see* Reply 12–13 (citing Ex. 1036 ¶¶ 26–38)) do not override the undisputed fact that an ordinarily skilled artisan *could* “make the 25 mM citrate, pH 3.5 ProSep A elution buffer using trisodium citrate and HCl” (Ex. 1036 ¶ 27 (testimony by Petitioner’s declarant, Dr. Przybycien); *see also* Ex. 2014, 88:22–89:8). In addition, we credit Dr. Cramer’s testimony that preparation of Shadle’s elution buffer with trisodium citrate and hydrochloric acid would have been a “well known[,] . . . readily-available and reasonable choice” (Ex. 2015 ¶ 59). For example, as Dr. Cramer testifies (*see id.*), Roth discloses preparing two elution buffers using trisodium citrate, and adjusting the pH of those buffers to 3.07 and 4.25 with hydrochloric acid (Ex. 2005, 1). Furthermore, we agree with Dr. Cramer that an ordinarily skilled artisan having trisodium citrate in the lab and interested in Shadle’s purification method could reasonably elect to prepare Shadle’s elution buffer using trisodium citrate and hydrochloric acid rather than by one of the four methods identified by Dr. Przybycien. Ex. 1034, 102:24–107:23. Finally, we observe that Patent Owner’s representation to the European Patent Office (“EPO”) that “the molarity of [Shadle’s] eluent can be calculated to *at least*: $(375 + 350)/15.35 = 47.2 \text{ mM}$ ” (Ex. 1006, 28 (emphasis added)) is

consistent with Dr. Cramer's testimony, as this calculation does not account for how the elution buffer was prepared or the contribution of solutes other than citrate present in the elution buffer to the molarity of the eluate (or the contribution of hydrochloric acid). *Id.* Furthermore, the calculation provided by Patent Owner to the EPO sets a floor, rather than a ceiling, for molarity of the neutralized eluate.

This is not a case where the prior art discloses a machine or process, which in normal operation or practice, would have produced a result required by a claim. *See In re Ackenbach*, 45 F.2d 437, 439 (CCPA 1930) (“[I]f a previously patented device, in its normal and usual operation, will perform the function which an appellant claims in a subsequent application for process patent, then such application for process patent will be considered, to have been anticipated by the former patented device.”). On the contrary, because Shadle's disclosure does not define the total concentration of solute present in its elution buffer, or restrict how that buffer is prepared beyond characterizing the concentration of citrate and pH, here the ordinarily skilled artisan would have had at her disposal a number of buffer formulations from which to choose—and selecting among them would not necessarily have resulted in a method that meets the claims. Even if we accept that the four options, identified by Dr. Przybycien, would have been more probable choices than the fifth, identified by Dr. Cramer, this is still a case that is based on odds or probabilities, rather than a result that flows naturally from the disclosure of the prior art. *See Perricone v. Medicis Pharm. Corp.*, 432 F.3d 1368, 1378 (Fed. Cir. 2005) (“[W]hen considering a prior art method, the anticipation doctrine examines the natural and inherent

results *in that method* without regard to the full recognition of those benefits or characteristics within the art field at the time of the prior art disclosure.” (emphasis added)). Petitioner’s position is based on the very kind of probability that precludes a finding of inherent disclosure. *See Endo*, 894 F.3d at 1381–82 (“the prior art was replete with *potential* co-solvents such that a skilled artisan, reviewing the Articles [the cited art], *would not have necessarily recognized* that the Articles’ authors used *benzyl benzoate as a co-solvent* for their reported clinical studies.” (emphases added)). Stated somewhat differently, the record before us does not adequately support Petitioner’s contention that Shadle’s elution buffer would necessarily be made using one of the four methods proposed by Dr. Przybycien.

The cases Petitioner relies on do not support its contention that the “law looks to the ‘normal and usual’ way a POSA *would practice* the prior art” (Reply 10–11) to fill gaps in the disclosure. Rather, these cases embody the above-stated principle that if a prior art method or device in its “normal and usual operation” *will perform the function claimed* in the challenged patent, “then such [patent] will be considered to have been anticipated by the [prior art].” *King Pharm., Inc. v. Eon Labs, Inc.*, 616 F.3d 1267, 1275–1276 (Fed. Cir. 2010) (internal quotations omitted); *see also In re King*, 801 F.2d 1324, 1327 (Fed. Cir. 1986) (“[T]he law is, and long has been, that ‘if a previously patented device, in its normal and usual operation, will perform the function which an appellant claims in a subsequent application for process patent, then such application for process patent will be considered to have been anticipated by the former patented device.’” (quoting *Ackenbach*, 45 F.2d at 439)). As explained above, that principle does not apply here.

The challenged claims of the '289 patent require the use of an elution buffer having a molarity of 100 mM or less *total solute* in order to perform the recited methods. Ex. 1001, 12:50–51. Shadle, in contrast, discloses only that the elution buffer includes 25 mM *citrate* and has a pH of 3.5. Ex. 1003, 20–21. Furthermore, the elution buffer of the '289 patent and Shadle are not *products resulting from* the performance of each of those methods. Rather, the elution buffers disclosed by the '289 patent and Shadle are *reagents for use in* each of the relevant methods. Thus, neither elution buffer can be said to be the inherent result of the relevant method.

Petitioner's analysis "goes astray because it assumes what [Shadle] neither disclosed nor rendered inherent." *Perricone*, 432 F.3d at 1379. Specifically, beyond the stated requirements for citrate concentration and pH, Shadle is agnostic as to how its elution buffer is prepared, or what additional solutes it includes. Ex. 1003, 20–21. Accordingly, it is not accurate to say that performance of Shadle's antibody purification method in its normal and usual way discloses an elution buffer having a total solute concentration of 100 mM or less. *See Endo*, 894 F.3d at 1381–82 ("First, Custopharm has not demonstrated that a skilled artisan could extrapolate the vehicle formulation used in the Articles from pharmacokinetic performance data. . . . Second, the prior art was replete with potential co-solvents such that a skilled artisan, reviewing the Articles, would not have necessarily recognized that the Articles' authors used benzyl benzoate as a co-solvent for their reported clinical studies."); *Perricone*, 432 F.3d at 1379 ("Because Pereira does not disclose topical application to skin sunburn, this court

reverses the district court's holding that Pereira anticipates claims 1–4 and 7 of the '693 patent.”).

Our rejection of Petitioner's inherency analysis does not turn on whether Petitioner has established that it would be “impossible” to practice Shadle without practicing the claimed invention, but rather, Petitioner's failure to prove that Shadle's disclosure is sufficient to show that the natural result flowing from performance of the method as taught would result in the claimed invention. *See SmithKline Beecham Corp. v. Apotex Corp.*, 403 F.3d 1331, 1343 (Fed. Cir. 2005) (“Apotex did not need to prove that it was impossible to make PHC anhydrate . . . that contained no PHC hemihydrate, but merely that ‘the disclosure [of the prior art] is sufficient to show that the natural result flowing from the operation as taught [in the prior art] would result in’ the claimed product.” (quoting *Oelrich*, 666 F.3d at 581)); *see also Atlas Powder Co. v. Ireco, Inc.*, 190 F.3d 1342, 1349 (Fed. Cir. 1999) (finding inherent disclosure of a claim limitation even though “the record showed that special mixing techniques – such as grinding and screening the AN particles – remove interstitial air from the blasting compositions” because the asserted reference “did not teach or suggest any such techniques”). In this regard, we highlight our finding, set forth above, that practice of the antibody purification method of Shadle in its normal and usual way does not require use of one of the four elution buffer preparations identified by Dr. Przybycien as common and conventional. Instead, we find, in view of Shadle's silence concerning how its elution buffer is prepared and how much total solute it includes (Ex. 1003, 20–21), as well as Dr. Cramer's testimony that an ordinarily skilled artisan reasonably could have employed

buffer formulations other than those described by Dr. Przybycien (Ex. 2015 ¶ 59; Ex. 1034, 103:21–104:5, 105:3–107:23), that Shadle does not necessarily require use of one of Dr. Przybycien’s four buffer formulations, and does not inherently disclose an elution buffer with 100 mM total solute. *See MEHL/Biophile Int’l Corp. v. Milgraum*, 192 F.3d 1362, 1365 (Fed. Cir. 1999) (“Occasional results are not inherent.”).

Petitioner’s further reliance on inherency to prove that Shadle discloses “neutralizing the eluate from step (2) to form particles by addition of a buffer to raise the pH to 4 to 8, wherein the molarity of the neutralized eluate is 100 mM or less” suffers from the same defects described above, and by layering deficient inherency arguments atop each other, magnifies those defects. Petitioner acknowledges that Shadle does not expressly disclose the molarity of the neutralized eluate of Example IA, but contends that it can be calculated based on other disclosures in that reference. Pet. 33. In Example IA, Shadle explains that subsequent to loading with IgG, i.e., antibody, the ProSep A affinity column was washed with “approximately 15 liters of PBS/glycine.” *Id.* Next, the “IgG was eluted by applying 15–20 liters of ProSep A elution buffer. Fractions of the non-bound peak and the elution peak were collected and assayed for IgG content using an HPLC assay. The eluate was approximately 15 liters in volume, and contained approximately 5 milligrams protein per milliliter.” *Id.* Shadle further explains that “[i]mmediately after elution, the sample was adjusted to pH 3.5 by the addition of 2.5 M hydrochloric acid, held for approximately 30 minutes, and adjusted to pH 5.5 by the addition of approximately 350 milliliters of 1 M Tris base.” *Id.* In order to determine the molarity of

total solute present in Shadle's neutralized eluate, it is necessary to account for contributions from the solutions used in each of these steps. *See* Ex. 1026 (calculating molarity based on contributions from elution buffer, hydrochloric acid, and Tris base); Ex. 1047 (additionally accounting for contribution from wash buffer).

As explained previously, however, Shadle does not describe how to prepare its elution buffer, or define the total concentration of solute present in that buffer. *See* Ex. 1003, 21. Neither does Shadle expressly describe the amount of hydrochloric acid added to the eluate, or state how much, if any, wash buffer is included in the neutralized eluate. *See id.* As such, it cannot be said that Shadle's neutralized eluate *necessarily* has 100 mM or less total solute, or that a neutralized eluate with 100 mM or less total solute is the "natural result of the combination of elements *explicitly disclosed* by the prior art." *Endo*, 894 F.3d at 1381 (emphasis added) (quoting *PAR*, 773 F.3d at 1196).

Petitioner's arguments to the contrary do not merit a different result. Absent disclosure, either express or inherent, of the molarity of Shadle's elution buffer, it is impossible to discern the molarity of Shadle's neutralized eluate, or to determine whether it satisfies the challenged claims. For example, Dr. Przybycien and Dr. Cramer testify regarding the effect of different ProSep A elution buffers, each prepared according to the requirements of Shadle, on the molarity of Shadle's neutralized eluate, and their combined calculations indicate that, even excluding any contribution of wash buffer from the analysis, only some—not all—elution buffers result in a neutralized eluate with 100 mM or less total solute. Ex. 2015 ¶¶ 60–64

(determining that the total molarity of the neutralized eluate obtained using elution buffer prepared with trisodium citrate and hydrochloric acid would be 102–108 mM); Ex. 1026, 2–5 (determining that the total molarity of the neutralized eluate obtained using elution buffer prepared according to one of the four methods identified by Dr. Przybycien would be 50.8–69.4 mM); *see also* PO Resp. 31–32 (discussing Dr. Cramer’s calculations). Although Petitioner bears the burden of proving inherent disclosure of any claim step, Patent Owner’s evidence, in the form of Dr. Cramer’s calculations, that at least one preparation of Shadle’s ProSep A elution buffer fails to satisfy the eluate molarity requirement recited in the challenged claims highlights the shortcomings of Petitioner’s inherency argument. The fact that Shadle’s neutralized eluate may additionally contain an unknown amount of wash buffer further undercuts Petitioner’s inherency analysis. *See* Ex. 1036 ¶ 50 (acknowledging that up to 0.582 L of wash buffer may be present in Shadle’s neutralized eluate); Ex. 2015 ¶ 71 (stating that by “conservative estimate,” 1L of wash buffer could be present in Shadle’s eluate); Ex. 2001, 5–6 (EPO examination division finding that Shadle’s eluate adjusted to pH 3.5 included 3.75–4.5 L of wash buffer).

Finally, because Petitioner has established neither that Shadle discloses an elution buffer with 100 mM or less total solute, nor an eluate with 100 mM or less total solute, its assertion that Shadle inherently discloses “neutralizing the eluate from step (2) *to form particles* by addition of a buffer to raise the pH to 4 to 8, wherein the molarity of the neutralized eluate is 100 mM or less” (Ex. 1001, 12:52–54 (emphasis added)) necessarily fails. *See Cont’l Can.*, 948 F.2d at 1268 (“To serve as an

anticipation when the reference is silent about the asserted inherent characteristic, such gap in the reference may be filled with recourse to extrinsic evidence. Such evidence must make clear that the missing descriptive matter is necessarily present in the thing described in the reference, and that it would be so recognized by persons of ordinary skill.”). Even assuming that adherence to the steps of the challenged claims would inherently result in particle formation, it cannot be said that particle formation inherently results from the performance of Shadle’s antibody purification process, because Petitioner has not established the molarity either of Shadle’s eluent or of Shadle’s eluate.

Accordingly, we find that Petitioner has not established by a preponderance of the evidence that claims 1–8 and 13 of the ’289 patent are anticipated by Shadle.

E. Obviousness Based on Shadle

Petitioner’s second ground challenges the same set of claims over the same reference as challenged in the first ground, except on obviousness under 35 U.S.C. § 103(a). Pet. 44. Patent Owner disagrees that Shadle renders the challenged claims obvious. PO Resp. 53–62.

A patent claim is unpatentable under 35 U.S.C. § 103(a) if the differences between the claimed subject matter and the prior art are such that the subject matter, as a whole, would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. *KSR Int’l Co. v. Teleflex Inc.*, 550 U.S. 398, 406 (2007). The question of obviousness is resolved on the basis of underlying

factual determinations including: (1) the scope and content of the prior art; (2) any differences between the claimed subject matter and the prior art; (3) the level of ordinary skill in the art; and (4) objective evidence of nonobviousness. *Graham v. John Deere Co.*, 383 U.S. 1, 17–18 (1966). “To satisfy its burden of proving obviousness, a petitioner cannot employ mere conclusory statements. The petitioner must instead articulate specific reasoning, based on evidence of record, to support the legal conclusion of obviousness.” *In re Magnum Oil Tools Int’l, Ltd.*, 829 F.3d 1364, 1380 (Fed. Cir. 2016).

The Petition addresses obviousness with only perfunctory assertions. For example, with regard to its contention that Shadle teaches or suggests the elution buffer molarity, neutralized eluate molarity, and particle formation requirements of the challenged claims, the Petition states:

In view of the disclosures of WO ’389 [Shadle] as discussed above for Ground I, all limitations of claims 1–8 and 13 were expressly or inherently disclosed. Thus, for the reasons explained above, it would also have been at least obvious for a POSA, based on the purification process disclosed in WO ’389, to arrive at and perform the method steps of claims 1–8 and 13— with a reasonable expectation of success. Ex. 1002 ¶¶ 103–106.

As discussed above for anticipation, WO ’389 discloses an antibody purification process that falls within the scope of claims 1–8 and 13 in the ’289 patent. *Id.* There is no patentable difference between the prior art antibody purification process of Example IA in and the claimed invention. *Id.* In light of these circumstances, the single prior art reference WO ’389 renders the claims obvious. In particular, a POSA would understand from the teachings of WO ’389 that DNA contaminants would be removed from an antibody sample by applying the sample to Protein A affinity chromatography column, eluting the antibody

sample with an acidic citrate solution of 25 mM and pH of 3.5, and then neutralizing the solution by raising the pH to 5.5 using a Tris buffer while the molarity of the solution remains below 100 mM. *Id.* The neutralized buffer solution is then filtered using a 0.1 micron and a 0.2 micron filter. *Id.*

Pet. 45–46. The Petition does not further elaborate on its assertion that Shadle teaches or suggests these claim requirements. *See id.* at 44–48. Indeed, the only claim step specifically addressed in the obviousness analysis set forth in the Petition is “removing the particles to thereby remove contaminant DNA from the antibody-containing sample” (Ex. 1001, 12:57–58). Pet. 46–47. Petitioner’s Reply is likewise superficial in its obviousness analysis, simply stating that “[e]ven if any limitation were not disclosed by Shadle at least inherently, it would have been obvious to a POSA” (Reply 23), without explaining why that would be the case for the elution buffer molarity, neutralized eluate molarity, or particle formation requirements of the challenged claims (*see id.* at 23–25).

Accordingly, in view of the deficiencies in Petitioner’s obviousness analysis, and for the reasons set forth above concerning anticipation by Shadle, we determine that Petitioner has not established, by a preponderance of the evidence, that claims 1–8 and 13 of the ’289 patent would have been obvious Shadle.²

² In light of the above described shortcomings in Petitioner’s obviousness analysis, we do not address Patent Owner’s objective evidence of nonobviousness. *See* PO Resp. 56–62.

III. MOTION TO EXCLUDE

Petitioner moves to exclude Patent Owner's Exhibits 2201–2207. Paper 36, 1. Patent Owner opposes the motion. Paper 44. As the moving party, Petitioner has the burden of proof to establish that it is entitled to the requested relief.

Petitioner asserts that Patent Owner untimely introduced Exhibits 2201–2207 for the first time during the deposition of Petitioner's Reply declarant, Dr. Przybycien. Paper 36, 4. Petitioner additionally asserts that the challenged exhibits are inadmissible because they are irrelevant. *Id.* Patent Owner responds that the objected-to exhibits were timely introduced because they were used to challenge opinions presented by Dr. Przybycien for the first time in conjunction with Petitioner's Reply. Paper 44, 2. Patent Owner further asserts that the exhibits are relevant because they illuminate gaps in Dr. Przybycien's Reply testimony. *Id.* at 3.

Because we have not relied on Exhibits 2201–2207, or Dr. Przybycien's testimony regarding those exhibits in this Final Written Decision, we dismiss Petitioner's Motion to Exclude Exhibits 2201–2207 as moot.

IV. CONCLUSION

For the foregoing reasons, we determine that Petitioner has not shown by a preponderance of the evidence that claims 1–8 and 13 of the '289 patent are unpatentable.

V. ORDER

It is

ORDERED that claims 1–8 and 13 of the '289 patent are not held unpatentable;

FURTHER ORDERED that Petitioner's Motion to Exclude is *dismissed* as moot; and

FURTHER ORDERED that, because this is a Final Written Decision, parties to the proceeding seeking judicial review of the Decision must comply with the notice and service requirements of 37 C.F.R. § 90.2.

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